# Molecular cloning and in silico analysis of DREB-like gene in watermelon

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#### ABSTRACT

The objective of this study was to identify and characterize sequences from watermelon related to drought tolerant genes. In order to achieve this, publicly available *Cucumis melo REB* gene (*Cme-DREB1*) sequence was retrieved from NCBI database and gene-specific primers were synthesized. A fragment of 274 bp corresponding to DREB-like gene was amplified from genomic DNA of watermelon. PCR product was cloned in *E. coli* vector (pDrive) and validated through sequencing. The BLAST search showed 99 and 97% homology to *CmeDREB1* gene and *C. sativus DREB3*-like (*CsDREB3*-like) gene, respectively. Furthermore, it showed 87 to 93% homology with rest of DREB genes available in the public database. Full length watermelon *DREB*-like gene (*C. lanatus DREB*-like, *ClaDREB*-like) was identified from cucurbit genomic database and investigated through *in silico* analysis. The intronless structural organization of *ClaDREB*-like gene was found with a conserved AP2/ERF domain.

Key words: Drought tolerance, DREB, in silico analysis, watermelon.

### INTRODUCTION

Watermelon [*Citrullus lanatus* var. *lanatus* (Thunb.) Matsumura & Nakai] is the third most important vegetable fruit crop. It belongs to the Cucurbitaceae family. Watermelon is a diploid (2n = 2x = 22) as well as in certain cases a tetraploid species (seedless) in nature. The genome size of the watermelon is 425 MB with 23,440 predicted genes (Guo *et al.*, 3). The contribution of watermelon in total vegetable production is 7% worldwide. China is the major producer of watermelon in the world. In India, it is grown predominantly in the arid and hot region and river basin areas of the country.

The watermelon and its wild relatives (for example Citrullus collosynthis) have the great potential to withstand severe water deficit conditions. Due to its deep root system and high accumulation of compatible solutes in root and aerial parts, watermelon serves as a suitable model system for understanding drought tolerance mechanism of C3 plants (Yoshimura et al., 14). Several abiotic stress tolerant genes have been isolated and characterized in watermelon. The enhanced expression of molecular chaperones as well as higher accumulation of numerous drought responsive metabolic enzymes was reported in the roots of wild watermelon under drought stress treatment (Yoshimura *et al.*, 14). Watermelon accumulates high concentrations of citrulline, glutamate and arginine (DRIP-1 homologue) in place of proline and glycinebetaine in its aerial parts,

consequently, which leads to the scavenging of hydroxyl radicals and induction of drought tolerant genes (Kawasaki et al., 6; Akashi et al., 2). cDNA sequences encoding COR15b and KIN1 genes were isolated from leaves of watermelon. The expression analysis of these genes revealed their involvement in the dehydration tolerance (Kang et al., 5). The Ccrboh, a drought-responsive gene, encoding the respiratory burst oxidase homologue (rboh) was cloned and investigated in a wild relative of watermelon, Citrullus colocynthis for drought responsiveness (Si et al., 10). Moreover, the transcriptome analysis of *C. colocynthis* revealed an array of differentially expressed drought responsive genes (Wang et al., 13). In addition to watermelon, 131 AP2/ERF gene family was identified and characterized in cucumber (Hu, 4).

Watermelon has been sequenced recently (Guo *et al.*, 3) and is a good source of agronomically important traits, however, the availability of information about genomic resources developed for understanding drought tolerance mechanism is still very less. Therefore, the present study was planned to isolate and characterize the drought tolerance related gene sequences in watermelon, specifically, the *DREB*-like gene from watermelon.

### MATERIALS AND METHODS

Five seeds of watermelon line DRB-669 (red pulp black seeded) were germinated on a moist germination paper at the temperature of 24°C. The seedlings at true two-leaf stage were selected for DNA isolation. The seedlings were immersed in liquid nitrogen and DNA was isolated according to CTAB method with

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some modification. The DNA was treated with *RNase A* for removing of RNA molecules and purified using phenol: chloroform and ammonium acetate method. The integrity of DNA was checked on 0.8% agarose gel followed by quantification using NanoDrop 1000 (Thermo Scientific, USA).

The available CmeDREB1 (Genbank accession No. AB125974.1) gene was retrieved from NCBI database and gene-specific primer pair (F- 5'CCCAGCCCTGAAATGGCTGCT3' & R- 5' TCGTTCCCAGAGCCGGCAAC3') was designed using Primerquest software of IDT (http://eu.idtdna. com/Primerguest/Home/Index). For amplification of ClaDREB-like gene, PCR assay was performed in 20 µl of reaction containing 1.0 µl DNA (100 ng/µl), 1.0 µl of each forward and reverse primer (10 mM), 1.0 µl dNTPs (10 mM), 0.5 µl Tag DNA polymerase, 2.5 µl 10 × buffer with MgCl<sub>2</sub> (Takara) and 13 µl of nuclease free water (Hi-Media, Mumbai). The thermal cycler machine (Eppendorf) was used for amplification with the following programme: an initial denaturation of 5 min. at 95°C; 35 cycles of 1 min. at 95°C, 30 sec. at 55°C and 2 min. at 72°C followed by a final extension of 10 min. at 72°C. The PCR products were analyzed on 1.0% agarose gel. The desired size products were cloned in pGEM T vector (Promega) and validated through sequencing.

The validation of partial *ClaDREB*-like gene sequence and protein homology was carried out using BLAST programme of NCBI Server (http://www.ncbi. nlm.nih.gov/). The partial sequence of *ClaDREB*-like gene was used as a query search in cucurbit genomic database (http://www.icugi.org/) for mining of full length gene. The full length genomic sequence of *ClaDREB*-like gene was used for structural analysis using Gene Structure Display Server (http://www. gsds.cbi.pku.edu.cn). Multiple protein sequence alignment and phylogenetic relationship of *ClaDREB*- like protein with publicly available database was established using MEGA 6.0 (Tamura et al., 12). The phylogenetic tree of proteins was constructed using the Neighbor-Joining model of MEGA 6.0. The conserved domain AP2/ERF was identified at the NCBI web site (http://www.ncbi.nlm.nih.gov/structure/ cdd/wrpsb.cgi.). Nucleotide sequence alignment between ClaDREB-like gene and other cucurbit DREB genes was carried out using CLUSTALW2 (http://www. ebi.ac.uk/Tools/msa/clustalw2/). The 1.5 kb upstream sequence of the identified gene was retrieved form watermelon database (http://www.icugi.org/cgi-bin/ ICuGI/genome/index.cgi/watermelon) and analyzed for drought stress responsive cis-regulatory elements using PlantCare (http://bioinformatics.psb.ugent.be/ webtools/plantcare/html/).

### **RESULTS AND DISCUSSION**

Based on accessible *Cucumis melo DREB1* (*CmeDREB1*) gene sequence at NCBI, the *DREB-like* gene was amplified in watermelon using designed gene-specific primers. PCR amplification was carried out to amplify a fragment of 274 bp size and was confirmed by gel electrophoresis (Fig. 1). To verify that, the amplified product belongs to *DREB* gene of *C. melo*, the resultant fragment was cloned in pGEM T vector and was sequenced. The obtained putative sequence of *ClaDREB*-like gene was analyzed with BLAST tool for sequence homology confirmation. The *ClaDREB*-like gene showed 99 and 97% sequence homology with *CmeDREB1* gene and *CsDREB3* gene, respectively.

Full length *ClaDREB*-like gene was identified from cucurbit genomic database using the cloned putative *ClaDEB*-like gene sequence as a query search. The BLAST search revealed a locus namely *Cla022212* on chromosome No. 8 showing maximum sequence similarity with putative fragment. The explored 651



Fig. 1. PCR amplification of *DREB*-like gene in watermelon. Lane M indicates standard marker, 1.0 kb DNA ladder. Lane 1-6 indicates amplification of *Osmotin* and *ERF* gene in watermelon. Lane 7 and 8 indicate amplification of *DREB*-like gene in *C. melo* (positive control) and watermelon, respectively. Arrow at the right side of gel indicates fragment of 274 bp corresponding to *DREB*-like gene.

bp sequence encoding ClaDREB-like gene was retrieved from database and deduced into amino acid sequence. The deduced amino acid sequence was further analyzed in BLASTp of NCBI database with default parameters. Twenty two DREB proteins homologous from different plant species matched with ClaDREB-like protein with homology ranging from 83.98% (CsDREB3) to 48.79% (VvDREB3) [Table 1]. Since DREB3 protein matched most with ClaDREBlike protein, therefore, DREB3 protein of other plants were taken for analysis. Additionally, the BLASTp analysis indicated close proximity of ClaDREBlike to CsDREB3 at amino acid based similarity rather than with CmeDREB1 gene as mentioned in initial analysis. The reason could be that the partial ClaDREB-like gene sequence covered only a part of the CmeDREB1gene with maximum homology, while at protein level it was found similar with CsDREB3 protein because of the contained conserved protein domains (AP2/ERF etc.).

To obtain insights into the structural organization of the *ClaDREB*-like gene, full length CDS and genomic sequences of the gene was used as an input sequence in the Gene Structure Display Server 2.0. The structural analysis by software showed the *ClaDREB*-like gene is intronless. It was also compared with structural organization of *CsDREB3* and *CmeDREB3* gene (Fig. 2A). Though, the *ClaDREB*-like gene of watermelon is comparatively shorter than *CsDREB3* and *CmeDREB3* gene due to occurrence of some deletions at different position in the gene (data not shown), however, the intronless organization of all the three analyzed genes showed conserved evolutionary relationship between them.

To investigate the AP2/ERF domain in *ClaDREB*like gene, predicted protein was analyzed in CDD of NCBI database. A domain with signature motif *YRGVRMRNWGKWVSEIREPRKKSRIWL GTFPSPEMAARAHDVAALSIKGNSAILNFPEL* was identified in *ClaDREB*-like protein sequence (Fig. 2B).

Tab	le 1	. Amino acid	sequence	homology	of	ClaDREB-like	protein	with	DREB3	protein	of other	plants.
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Subject ID	% identity	Alignment length	Mismatches	e-value	Bit score
CsDREB3	83.98	206	24	1e-96	272
CmeDREB3	83.5	206	25	4e-90	255
FvDREB3	60.09	223	49	9e-85	243
PmDREB3	59.35	214	40	2e-79	229
EgDREB3	56.31	222	71	9e-75	218
MnDREB3	59.72	216	51	1e-73	214
CsiDREB3	62.98	208	57	3e-73	212
MdDREB3	57.01	221	74	5e-73	213
CsaDREB3	52.99	234	76	9e-73	212
AtDREB3	52.99	234	76	1e-71	208
NnDREB3	59.9	207	60	3e-71	207
PeDREB3	59.9	197	59	7e-70	205
GmDREB3	56.73	208	63	7e-70	202
BrDREB3	58.25	194	64	1e-69	204
CaDREB3	59.67	181	54	1e-69	201
BnDREB3	60.56	180	53	5e-65	191
VvDREB3	48.79	248	70	7e-64	190
StDREB3	62.01	179	44	2e-63	187
BvDREB3	52.28	241	67	8e-63	187
GsDREB3	61.99	171	46	7e-62	183
SIDREB3	59.36	187	46	1e-61	182
NsDREB3	60.73	191	34	3e-61	181

Abbreviation: Cs (Cucumis sativus); Cme (Cucumis melo); Fv (Fragaria vesca); Pm (Prunus mume); Eg (Eucalyptus grandis); Mn (Morus notabilis); Csi (Citrus sinensis); Md (Malus domestica); Csa (Camelina sativa); At (Arabidopsis thaliana); Nn (Nelumbo nucifera); Pe (Populus euphratica); Gm (Glycine max); Br (Brassica rapa); Ca (Cicer arietinum); Bn (Brassica napus); Vv (Vitis vinifera); St (Solanum tuberosum); Bv (Beta vulgaris); Gs (Glycine soja); Sl (Solanum lycopersicum); Ns (Nicotiana sylvestris).



Fig. 2. Structural analysis of *ClaDREB*-like gene and protein. A, represents the structural organization of *ClaDREB*-like gene and comparison with *CsDREB3* and *CmDREB3* gene. B, represents the AP2/ERF domain present in *ClaDREB*-like protein with consensus sequence.

The characteristics features of *ClaDREB*-like gene, *i.e.*, 3 Beta sheet and a antiparallel alpha helix with two functional conserved amino acids, valine and glutamine, at 14 and 19 positions, respectively in Beta sheet was found similar to reported findings (Sakuma *et al.*, 9; Lata *et al.*, 7).

For evaluating the evolutionary relationships between *ClaDREB*-like protein of watermelon and DREB proteins of other plants, a Neighbor-Joining, unrooted phylogenetic tree with 1,000 bootstrap replicates was generated. The phylogenetic analysis grouped them into three major distinct clusters. Predictably, *ClaDREB*-like protein clustered with other cucurbit DREB protein (Fig. 3).

Question arose that whether ClaDREB-like gene contains same DREB responsive *cis*-regulatory elements as described in other plants. To get insight into this query, a 1.5 kb upstream region as a putative promoter of ClaDREB-like gene was identified in cucurbit genomic database. The putative promoter was subjected to search for *cis*-regulatory elements using PlantCare. The analysis revealed many drought responsive cis-regulatory consensus sequences (TAACTG, CANNTG, WAACAA, CNGTTR, YAACKG, ACTGT, ACGT and CATGTG) in the putative promoter region. The presence of such elements in the promoter indicates its strong possibility in drought-inducibility. The similar drought responsive elements were described in Arabidopsis thaliana (Abe et al., 1; Simpson et al., 11) and rice (Oh et al., 8).



Fig. 3. Phylogenetic tree analysis among ClaDREBlike and DREB3 protein of other plants. For tree construct the Neighbor-Joining method with bootstrap consensus tree inferred 1000 replicates was taken to represent the evolutionary history of the *DREB* genes. The value on braches indicates percent confidence of bootstrap. The ClaDREB-like protein is indicated in rectangular box.

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## REFERENCES

- Abe, H., Urao, T., Ito, T., Seki, M., Shinozaki, K. and Yamaguchi-Shinozaki, K. 2003. *Arabidopsis* AtMYC2 (bHLH) and AtMYB2 (MYB) function as transcriptional activators in abscisic acid signaling. *Plant Cell*, **15**: 63-78.
- Akashi, K., Nishimura, N., Ishida, Y. and Yokota, A. 2004. Potent hydroxyl radical-scavenging activity of drought-induced type-2 metallothionein in wild watermelon. *Biochem. Biophys. Res. Comm.* 323: 72-78.
- Guo, S., Zhang, J., Sun, H., Salse, J., Lucas, W., Zhang, H., Zheng, Y., Mao, L., Ren, Y. and Wang, Z. 2013. The draft genome of watermelon (*Citrullus lanatus*) and resequencing of 20 diverse accessions. *Nature Genet.* 45: 51-58.
- Hu, L. and Liu, S. (2011). Genome-wide identification and phylogenetic analysis of the ERF gene family in cucumbers. *Genet. Mol. Biol.* 34: 624-33.
- Kang, G., Zhu, Z., Guo, T. and Ren, J. 2009. Isolation and expression pattern of *COR15b* and *KIN1* genes in watermelon and pumpkin. *African J. Biotech.* 8: 5666-72.
- Kawasaki, S., Miyake, C., Kouchi, T and Yokota, A. 2000. Responses of wild watermelon to drought stress: Accumulation of an *ArgE* homologue and citrulline in leaves during water deficit. *Plant Cell Physiol.* **41**: 864-73.
- 7. Lata, C., Bhutty, S., Bahadur, R.P., Majee, M. and Prasad, M. 2011. Association of a SNP in a novel *DREB2*-like gene *SiDREB2* with stress tolerance

in foxtail millet [*Setaria italica* (L.)]. *J. Exp. Bot.* **62**: 3387-3401.

- Oh, S.J., Song, S.I., Kim, Y.S., Jang, H.J., Kim, S.Y., Kim, M., Kim, Y.K., Nahm, B.H. and Kim, J.K. 2005. *Arabidopsis CBF3/DREB1A* and *ABF3* in transgenic rice increased tolerance to abiotic stress without stunting growth. *Plant Physiol.* **138**: 341-51.
- Sakuma, Y., Liu, Q., Dubouzet, J.G., Abe, H., Shinozaki, K. and Yamaguchi-Shinozaki, K. 2002. DNA-binding specificity of the *ERF/AP2* domain of *Arabidopsis* DREBs, transcription factors involved in dehydration- and cold-inducible gene expression. *Biochem. Biophys. Res. Comm.* 290: 998-1009.
- Si, Y., Dane, F. Rashotte, A., Kang, K. and Singh, N.K. 2010. Cloning and expression analysis of the *Ccrboh* gene encoding respiratory burst oxidase in *Citrullus colocynthis* and grafting onto *Citrullus lanatus* (watermelon). *J. Exp. Bot.* 61:1635-42.
- Simpson, S.D., Nakashima, K., Narusaka, Y., Seki, M., Shinozaki, K. and Yamaguchi-Shinozaki, K. 2003. Two different novel *cis*-acting elements of *erd1*, a clpA homologous of *Arabidopsis* gene function in induction by dehydration stress and dark-induced senescence. *Plant J.* 33: 259-70.
- Tamura, K., Stecher, G., Peterson, D., Filipski, A. and Kumar, S. 2013. MEGA6: Molecular Evolutionary Genetics Analysis version 6.0. *Mol. Biol. Evol.* **30**: 2725-29.
- Wang, Z., Hu, H., Goertzen, L.R., McElroy, J.S. and Dane, F. 2014. Analysis of the *Citrullus colocynthis* transcriptome during water deficit stress. *PLoS ONE*, **9**: e104657. doi:10.1371/ journal.pone.0104657.
- Yoshimura, K., Masuda, A., Kuwano, M., Yokota, A. and Akashi, K. 2008. Programmed proteome response for drought avoidance/ tolerance in the root of a C3 xerophyte (wild watermelon) under water deficits. *Plant Cell Physiol.* 49: 226-41.

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