

# Breeding and genomic investigations for quality and nutraceutical traits in vegetable crops-a review

Shrawan Singh, Selvakumar R., Manisha Mangal and Pritam Kalia<sup>\*</sup>

Division of Vegetable Science, ICAR-Indian Agricultural Research Institute, New Delhi 110012

## ABSTRACT

Globally, micronutrient malnutrition is a big burden in pubic health spread across the geopolitical regions and socio-economic strata. The efforts for challenging this 'hidden hunger' are dietary diversity, food supplementation, food fortification and biofortification of common food crops. Human body requires nearly 49 different dietary elements and maintaining their regular intake seems to more effective and sustainable through dietary diversity and biofortified crops, however, their accessibility and affordability in continuum remain issues. For this, vegetable crops have great promise to serve for dietary needs because these crops represent a group of more than thousand plants spread all across world and constituent of regular diets. Although, accounted vegetable production in the world is 1088.8 million tonnes but huge numbers of minor vegetables also serve for dietary minerals in poor or rural communities inhabiting in vulnerable regions. Vegetables are bulky in nature but they are the key sources of health protective dietary constituents such as minerals, vitamins and antioxidants in human diet. Their intake also provides dietary elements and antioxidants to protect body from various immune-related diseases by acting against free radicals. The complex matrix of various constituents and minerals influence consumer preference and absorption pattern also. However, the focus on breeding for yield traits affected quality attributes seriously, hence it become essential to breed varieties having high yield and better quality traits. For this, molecular tools and genome sequencing techniques showed effectiveness to speed up the classical breeding methods for these traits of complex pathways. Besides, genome editing techniques such as transgenics, RNA interference and CRISPR/cas have great prospect in vegetables for enriching health beneficial constituents and also for removal of antinutritional factors. The present review reports significant achievements from attempts on understanding genetic of quality traits and breeding vegetables for quality, nutraceutical, pharmaceutical and other industrial traits.

Key words: Anthocyanin, carotenoids, minerals, molecular markers, bioactive compounds, nutrient, medicinal properties.

### INTRODUCTION

Nutrition is basic prerequisite to sustain human life and activity which depends on quality of food. Essentially, safe and nutritious food is inherent part of food security which means 'all people, at all times, have physical, social, and economic access to sufficient, safe and nutritious food that meets their food preferences and dietary needs for an active and healthy life (World Food Summit, 256). Further, rising awareness of environmental, nutritional and health concerns are leading to changes in consumer behaviour towards demand of quality nutritious foods. Quality food got promise to challenge different Sustainable Development Goals (SDGs), namely SDG-1 (No poverty), SDG-2 (Zero hunger and improved nutrition), SDG-3 (Good health & well being), SDG-4 (Quality education and cognitive development & learning), SDG-5 (Reduced inequalities and gender equality), SDG-7 (Improved work and productivity) and SDG-8 (decent work and economic growth). But, major concern is deficiency of micronutrients in children

and vulnerable population because nearly 60% of the world's people are iron (Fe) deficient, over 30% are zinc (Zn) deficient, 30% are iodine (I) deficient and 15% are selenium (Se) deficient summarised by White and Broadley (250). Their deficiencies are occurring in the form of poor physical growth, intellectual impairments, perinatal complications and increased risk of morbidity and mortality and loss of productivity. Iron (microcytic anemia), iodine (goiter, mental retardation and cognitive function), folate, vitamin A, calcium and zinc are nutrients of major public health concern because around 2 billion people suffer from micronutrient deficiencies globally. In India, malnutrition accounts for 68.2% of total underfive deaths, 21.4% low birthweight, 39.3% stunting, 15.7% wasting and 32.7% underweight, 59.7% anaemia in children (Swaminathan et al., 221). Dietary diversity, food supplementation, food fortification and biofortification are common approaches to handle micronutrient malnutrition. None of them appears as panacea to challenge the complexities of this hidden hunger in population, however their combination have added value to mitigate its incidence.

Corresponding author's Email: pritam.kalia@gmail.com

Food quality is a multidimensional concept which not only depends on the property of the food but also on the consumer choice and preference. Consumer behaviour is mainly influenced by sensory attributes (shape, size, colour, flavour, aroma and taste) but, nutritive value and secondary metabolites determine health and nutritional benefits. The quality is defined as 'sum the all characteristics, properties and attributes of a product or commodity which is aimed at fulfilling the established or presumed customer requirement' (ISO 8402, 1989). This support the common view of 'the customer returns and the product does not'. Welch and Graham (249) listed 49 known essential nutrients for sustaining human life in six categories, namely (i) water and energy (water and carbohydrate), (ii) protein and amino acids (histidine, isoleucin, leucine, lycine, methionine, phenyl alanine, threonine, tryptophan and valine), (iii) macroelements (Na, K, Ca, Mg, S, P, Cl), (iv) microelements (Fe, Zn, Cu, Mn, I, Fe, B, Se, Mo, Ni, Cr, V, Si, As, Sn, Co), (v) fatty acids (lenoleic acid and  $\alpha$ -linolenic acid), and (vi) vitamins (A, D, E, K, C, B<sub>1</sub>, B<sub>2</sub>, B<sub>3</sub>, niacin, B<sub>6</sub>, folate, biotin and B<sub>12</sub>). To meet up this nutrient requirement, it is suggested to intake 400 g fruits and vegetables (excluding potato and starchy food) (FAO/WHO, 61) while ICMR (100) recommended for 300 g vegetables and 100g fruits per capita per day.

Vegetables are rich in dietary minerals and vitamins along with good amount of natural antioxidants. They are strong agent in improving fight against micronutrient malnutrition because (i) they are traditional part of every household diets, (ii) available worldwide with global production of 1088.8 million tonnes including India (128.2 million tonnes) (FAOSTAT, 62), (iii) successful examples of biofortification in vegetables such as sweet potato, cassava and beans, (iv) epidemiological studies shows association in intake of fruit and vegetables in risk of cancer, diabetes and cardiovascular diseases, (v) diverse vegetable species (1097) grows worldwide to add in dietary diversity, (vi) compound matrix in vegetables favours bioavailability of nutrients, (vii) great prospect for enriching further by use of diverse germplasm, and (viii) wealthier population is willing to include more fruits and vegetables in their daily diet and the regulations are coming from different agencies such as European Union Regulation No. 1151/2012 (Gragnani, 82). Human efficacy trials have shown that diets having biofortified rice (for Fe) and orange fleshed sweet potato (for Vit. A) and sCAX1 carrot (for Ca) significantly improved respective micronutrient status in human.

# Recommended dietary allowance (RDA) of dietary nutrients:

The RDAs are the levels of intake of essential nutrients that, on the basis of scientific knowledge, are judged by the Food and Nutrition Board to be adequate to meet the known nutrient needs (97-98%) of practically all healthy persons. The RDA must not be considered requirements but as levels of intake adequate for maintaining good health. The suggested intake of 300g of vegetables and 100g of fruits are almost enough to meet the RDA of most of nutrients of body as given in Table 1. However, the RDA is different from Dietary Reference Intake (DRI) which in addition to RDA, include maximum intake levels (ULs) and preferred because intake increases above the UL, the risk of adverse effects increases. The ULs have been prescribed for different compounds such as vitamin A (3000 µg/day), vitamin C (2000 mg/day), vitamin D (100 µg/day), vitamin E (1000 mg/day), niacin (35 mg/day) and folate (1000 µg/day).

#### Dietary minerals in vegetables

Minerals play a major role in the functioning of the physiological activities and reproduction. They are component of various body constituents such as (i) Calcium is the essential component of bones and teeth, (ii) Minerals present in blood as electrolytes like Na<sup>+</sup>, K<sup>+</sup>, Ca<sup>++</sup>, Mg<sup>++</sup>, Cl<sup>++</sup>, HCO<sup>-</sup><sub>3</sub>, HPO<sup>-</sup><sub>4</sub>, H2PO<sup>=</sup> etc., (iii) Iron is the important component of haemoglobin. Iron content of crystalline human haemoglobin is 0.34 per cent, (iv) Phosphorus along with C, H, N and O are the components of the DNA. It is also constituent of phospholipids, and (v) Deficiency of iodine interferes with the function of thyroid gland resulting in swelling and cause goiter. Their deficiencies lead to different forms of ailments such as anemia (Iron), impaired gastrointestinal and immune functions and anemia (Zinc), osteoporosis (Calcium), goiter (Iodine). Leafy vegetables are rich source of these mineral elements, such as spinach, amaranth, palak or beet leaf, coriander, Indian spinach, drumstick leaves as reviewed by Natesh et al. (164). Interestingly, minerals have greater stability during kinds of food processing as compared to vitamins and proteins, hence, intake of prescribed quantity of leafy vegetables in daily diet is beneficial for health.

#### Health beneficial nutraceuticals in Vegetables

Certain secondary metabolites in vegetables have strong capacity for quenching free radicals or influencing certain biological processes and thus contribute in health promotion. These are bioactives or phytochemicals or nutraceuticals, of them, latter are certain compounds which have roles in providing nutrition *vis-a-vis* serve as medicines against ailments.

Breeding and Genon	nic Investigations in	Vegetable C	Crops-A review
--------------------	-----------------------	-------------	----------------

Table 1.	. Recommended	nutrient intakes	for	adult males	and	females	(data froi	m FAO/WHO	and	ICMR)
----------	---------------	------------------	-----	-------------	-----	---------	------------	-----------	-----	-------

Nutrient	Global (FAO/WHO)		Indian (ICMR)	
	Male	Female	Male	Female
Energy (kcal)	2900	2200	2730	2230
Protein (g)	63	50	60	55
Fat(g)	NA	NA	30	25
Vitamin A (µg retinol equivalent)	1000	800	600	600
Vitamin D (µg)	5.0	5.0	10	10
Vitamin E (mg α-tocopherol equivalent)	10	8	7.5	7.5
Vitamin K (µg)	80	65	55	55
Vitamin B <sub>6</sub>	1.3	1.3	2.0	2.0
Vitamin B <sub>12</sub> (µg)	2	2	1.0	1.0
Riboflavin (mg)	1.7	1.3	1.6	1.3
Niacin (mg niacin equivalent)	19	15	18	14
Thiamin (mg)	1.5	1.1	1.4	1.1
Vitamin C (mg)	90	60	40	40
Folate (µg)	200	180	200	200
Calcium (mg)	800	800	600	600
Phosphorus (mg)	800	800	600	600
Magnesium (mg)	350	280	340	310
Sodium (mg)	2000	2000	2100	1900
Potassium (mg)	2000	2000	3750	3225
Iron (mg)	10	15	17	21
lodine (µg)	150	150	150	150
Zinc (mg)	10	15	12	10

NA- Not available.

Pandey et al. (168) classified nutraceuticals as potential and established nutraceutical compounds based on establishing their activity after clinical trials. It is to be noted that much of the nutraceutical products are still in the 'potential' category (Das et al., 42). The common nutraceuticals and their reported health functions are given in Fig. 1. Major attention of consumers is for carotenoids, anthocyanins, flavonoids, glucosinolates, capsaicin, oleoresins and terpenoids due to their potential health benefits. Although, vegetables are inherently rich in such compounds, but some of them lacks one or more dietary/health compounds despite their existence in respective crop germplasm. Hence, breeders explored these germplasm for potential nutraceuticals to enrich commercial varieties through classical or molecular based breeding approaches. Emphasis is towards enrichment of lycopene, vitamin C and anthocyanin in tomato, beta-carotene in cauliflower, anthocyanin and beta-carotene in carrot, glucosinolates and anthocyanin in broccoli, beta-carotene in melons and dietary minerals in leafy vegetables. Similarly,

elimination/reduction of antinutritional factors also successfully attained in some of the legume and leafy vegetables.However, novel techniques in breeding and genomics have great promise in tailoring varieties for specific dietary nutrients.

#### Dietary nutrients for human health

**Dietary fibers:** Dietary fibers mostly include nonstarch polysaccharides (NSP) such as celluloses, hemicelluloses, gums and pectins, lignin, resistant dextrins and resistant starches. Vegetables are rich source of dietary fibers. The soluble components of dietary fiber affect the rate of digestion and the uptake of nutrients, improve glucose tolerance, enhance insulin receptor binding and improve glycaemic response. Intake of high fiber food improves serum lipoprotein values, lowers blood pressure level, improves blood glucose control for diabetes, aids weight loss and promotes regularity. It is recommended that dietary fiber intake for adults generally fall in the range of 20–35 g/day (Pilch,



Fig. 1. Important nutraceuticals in plants (inner whorl) and their role in human health (outer whorl)

179). Curry leaves, drumstick leaves, amaranth and colocasia leaves are among the richest sources of dietary fiber in vegetable crops.

**Polyunsaturated fatty acids (PUFAs):** Although, all fats have important roles in the body but major concern is about those fatty acids which body cannot synthesize and depends on food. The essential fatty acids are linoleic acid (omega-6 group) and alpha-linolenic acid (omega-3 group). The FDA recommends a maximum of 3 g/day intake of EPA and DHA omega-3 fatty acids. Tender leaves of colocasia, drumstick, parsley, spinach, mustard, pumpkin, radish, Amaranth, beet leaf are good source of essential fatty acids. Further, cucurbit seeds, soybean seeds (green), leek, beans are also source for PUFAs.

**Probiotics:** Probiotics also represent a form of nutraceuticals which can be defined as 'live microorganisms which when administered in adequate amounts confer a health benefit on the host' (FAO/WHO, 60). They include *Lactobacilli* and *Bifidobacteria*. Savadogo *et al.* (205) found biofortification of cassava tubers with *Lb. rhamnosus*, *Lb. plantarum, Lb. acidophilus, Lb. reuteri, Lb. fermentum, Lb. brevis* and *Lb. salivarus* as promising option for contributing to folate intake in human body. Saubade *et al.* (203) reported that lactic acid fermentation can be explored as a tool for increasing the folate content of foods.

Most of the nutraceuticals are present across the plant families but there are specific nutraceuticals

which occurs in particular groups of vegetables such as glucosinolates in *Brassicas*, Allylsulfides in *Alliums* and Silymarin in Compositae plants. The most commonly observed nutraceuticals in different vegetable groups are given in Table 2.

**Vitamins in human health:** Essential building block of certain coenzymes which are indispensable for the normal vital activities. All vitamins are essential for human body except vitamin D. Important source vegetables for different vitamin are given in Table 3.

#### Dietary antioxidants in human health

Antioxidants are basically phytochemicals or phytoceuticals naturally present in plant foods like vegetables which neutralize harmful free radicals generated in the body. These phytochemicals are secondary metabolites and formed due to the enzymatic reactions of primary metabolites (amino acids, sugars, vitamins etc.). A 'dietary antioxidant' is a substance in foods that significantly decreases the adverse effects of reactive oxygen species (ROS) and reactive nitrogen species (RNS) on normal physiological function in human beings and prevents from related health ailments. The ROS free radicals are super oxides, hydroxyperoxyl (HO<sub>2</sub>), hydroxyl (OH), peroxyl (RO<sub>2</sub>) and alkoxyl (RO) while common RNS are nitric oxide (NO•) and peroxynitrite (ONOO•). The free radicals are generated either from normal metabolic processes of body or from external sources (X-rays, ozone, smoking, air pollutants). They target Breeding and Genomic Investigations in Vegetable Crops-A review

Nutraceuticals/ bioactive compounds	Vegetables
Allylsulfides, Fructan	Allium vegetables (garlic, onions, chives, leeks)
Glucosinolates	Brassica vegetables (broccoli, cauliflower, cabbage, Brussels sprouts, kale, turnips, kohlrabi)
Lycopene	Tomato, red carrot, water melon, seed aril of Momordica vegetables
Polyphenols	Potato, brinjal, okra, leafy vegetables, onion
Anthocyanin	Black carrot, beet root, red amaranth, red lettuce, red cabbage, brinjal
Carotenoids	Leafy vegetables, carrot, orange cauliflower, orange flesh sweet potato and cassava, muskmelon, pumpkin
Flavonoids, carotenoids, apigenin	Apiaceae (Celery, parsley carrot)
Silymarin	Compositae plants (artichoke)
Folate	Chenopodiaceae (spinach, swiss chard, beet greens)
Flavonoids (isoflavones)	Beans

Table 2.	Vegetable	crops	rich in	nutraceuticals/bioactive	compounds.
----------	-----------	-------	---------	--------------------------	------------

Table 3. Functional/deficiencies of nutrients in human body and their potential source vegetables.

Dietary nutrients	Function/deficiency	Rich vegetable sources (per 100 g edible portion)
Carbohydrate	Source of energy Protein sparing food	Potato (22.6 g), sweet potato (27.0g), cassava (32.4g), elephant foot yam (18.4g), taro (21.1g), pea (15.8g), lima bean (23.5g)
Protein	Body building mainly for tissues, muscles and blood, deficiency leads to kwashiorkor disease.	Pea (7.2g), cowpea (4.3g), broad bean (4.5g), lima bean (7.5g), leaves of fenugreek (4.4g), drumstick (2.5g), Brussels sprout (4.4g)
Fat and oil	Source of energy, part of colloidal complex of cytoplasm	Chilli (0.6g), sweet pepper (0.3g), brinjal (0.3g), Brussels sprouts (0.5g), snake gourd (0.3g), pointed gourd (1.0g), sweet corn (0.3g), Bengal gram leaves (1.4g), bitter gourd (1.0g)
Vitamin A (xerophthol, retinol)	(i) Night blindness (nycalopia), (ii) xerophthalmia in infants and young children causing keratinisation in epithelial cells of eyes (iii) Dryness in skin	Carrot (12000 IU), amaranthus (9200 IU), <i>palak</i> or beet leaf (9770 IU), spinach (9300 IU), fenugreek leaves (3744 IU), mustard leaves (4195 IU), coriander leaves (11168 IU), broccoli (3500 IU), musk melon (3420 IU), kale (7540 IU)
Thiamine	Beri-beri, mascular weakness and loss of weight, neuritis, fluid in body cavities (oedema) and impaired function, dilation of heart and loss of appetite	Potato (0.40 mg), <i>Palak</i> or beet leaf (0.26 mg), pea (0.25 mg), tomato (0.19 mg), chilli (0.19 mg), muskmelon (0.11 mg), garlic (0.16 mg), leek (0.23 mg), coriander leaves (0.50 mg)
Riboflavin	Dark red inflamed tounge, dermatitis (inflammation of skin), loss of hair and dry scaly skin, diarrhoea, cracks in corners of mouths, ulcers in oral cavity, cracked lips	<i>Palak</i> or beet leaf (0.56 mg), chilli (0.19 mg), winged bean (0.12 mg), knol khol (0.10 mg), broccoli (0.12 mg), Brussels sprouts (0.13 mg)
Niacin	Pellagra, nervous breakdown, stomach and intestinal disorder	<i>Palak</i> or beet leaf (3.3 mg), amaranth (1 mg), bitter gourd, giant spine gourd, pointed gourd, bottle gourd, pumpkin (0.5-0.9 mg)
Ascorbic acid	Scurvy (dropsy, anaemia, bleeding gums, and mucus membranes), delayed healing of wounds, reduced resistance to diseases	Sweet potato, chilli, cabbage, broccoli, kale, drumstick, coriander leaves, cauliflower.
Folic acid	Anaemia (megaloblastic) more acute in pregnant women, impaired growth and nervous breakdown	<i>Palak</i> or beet leaf, lettuce, cabbage, spinach, cowpea, French bean

Dietary nutrients	Function/deficiency	Rich vegetable sources (per 100 g edible portion)
Tocopherol	Degeneration of kidney, necrosis of liver, reduction in capacity of reproduction	Zuccini (Green) (3.90 $\mu$ g), Zuccini yellow (1.47 $\mu$ g), Agathi leaves (1.48 $\mu$ g), Spinach (1.27 $\mu$ g), Onion (0.81 $\mu$ g), Cluster bean (0.77 $\mu$ g)
Phylloquinones (Vit K)	Delayed and faulty coagulation of blood cut wounds, hindrance in normal secretion of bile from liver	Drumstick leaves (479 $\mu$ g), <i>Amaranths spinosus</i> (433 $\mu$ g), Fenugreek leaves (428 $\mu$ g), Drumstick (358 $\mu$ g), Spinach (325 $\mu$ g), Parsley (322 $\mu$ g)
Ergocalciferol (Vit D <sub>2</sub> )	Useful against hypoparathyroidism and rickets	Baby corn (31 $\mu$ g), Drumstick leaves (14 $\mu$ g), Cluster bean (13 $\mu$ g), Tomato (12 $\mu$ g), Pea (12 $\mu$ g), Broad bean (11 $\mu$ g)
Lutein	Maintaining eye health and reducing the risk of macular degeneration and cataracts	Agathi (12941 µg), Drumstick leaves (15580 µg), Spinach (3850µg), Parsley (3574 µg), Fenugreek leaves (2275 µg)
Zeaxanthin	Eye health	Agathi leaves (559 $\mu$ g), <i>Amaranthus gangeticus</i> (164 $\mu$ g), Sweet potato (146 $\mu$ g), Fenugreek leaves (28 $\mu$ g), Pumpkin (34 $\mu$ g), Zuccini yellow (25 $\mu$ g)
Lycopene	Antioxidant, protect against sunburns and certain types of cancers	Tomato hybrid (2481 $\mu$ g), tomato green (35 $\mu$ g), watermelon dark skin (1477 $\mu$ g), watermelon pale green (1257 $\mu$ g)

Table 3 contd...

(Source: Longvah et al. 141; Hazra and Som, 215)

DNA, proteins, lipids and carbohydrate and contribute to the development of chronic diseases such as macular degeneration, antherosclerosis, cancer, coronary heart diseases and rheumatoid arthritis. Although, body has its own redox homeostasis mechanism to neutralize free radicals but to a certain extent, afterwards, it need antioxidants from external sources to assist scavenging process. The antioxidants neutralize free radicals through different mechanisms: (i) Decreasing ROS or RNS formation, (ii) binding metal ions needed for catalysis of ROS generation, (iii) scavenging ROS, RNS, or their precursors, (iv) up-regulating endogenous antioxidant enzymes defenses, (v) repairing oxidative damage to biomolecules such as glutathione peroxidases or specific DNA glycosylases, (vi) influencing and upregulating repair enzymes, and (vii) reacting directly with free radical in a non-catalytic manner before the radicals react with other cell components. However, the effectiveness of each dietary antioxidant depends upon (i) which ROS or RNS is being scavenged (ii) how and where they are generated, (iii) the accessibility of antioxidants to this site, and (iv) what oxidizable substrate is involved.

The evidences showed that lycopene, betacarotene, ascorbic acid,  $\alpha$ -tocopherol, flavonoids and selenium are significantly associated with reduced cancer risk. Vitamin E, vitamin C, carotenoids, anthocyanins and phenolics are able to neutralize free radicals and inhibit LDL oxidation and potentially reduce the risk of coronary artery diseases.

**Carotenoids:** Carotenoids play an important role in plant reproduction, through their role in attracting

pollinators and in seed dispersal, and are essential components of human diet. Lycopene,  $\alpha$ -carotene,  $\beta$ -carotene and xanthophylls are common carotenoids from vegetables. Carotenoids provide protection to vision and eye function, and against macular degeneration and cataracts. They promote immune system response and presumed to be associated with inhibition of several types of cancers including cervical, oesophageal, pancreatic, lung, prostate, colorectal and stomach.

**Phenols:** Phenols or phenolics are secondary metabolites and often found in polymeric forms (polyphenols). Phenolics have role in human health due to antioxidant properties (reducing or metal chelating agents, hydrogen donors and singlet oxygen quenchers) and alternative mechanisms (cellular signaling, gene expression and modulation of enzymatic activity). Habauzit and Morand (91) reviewed the evidences for protective effect of phenolics. Besides, phenolics may also affect sensory characteristics of food with impacts on colour, flavour and astringency. Major polyphenols are flavonoids, phenolic acids, tannins (hydrolysable and condensed), stilbenes and lignans.

**Flavonoids:** The basic flavonoid structure is the flavan nucleus, which consists of 15 carbon atoms arranged in three rings (C6-C3-C6), labeled A, B, and C. Important flavonoids in vegetables are quercetin, kaempferol, myricetin and lutein. Onion, kale, broccoli, green beans and celery are rich in flavonoids while tomato, red pepper and broad beans are medium.

Anthocyanins: When the anthocyanidins are found in their glycoside form (bonded to a sugar moiety), they

are known as anthocyanins (Castaneda-Ovando et al., 23). There is a substantial variety of anthocyanins spread in nature. They are water-soluble pigments and impart red (in low pH), purple, and blue (in high pH) colour which are influenced by pH, light, temperature, and structure. Around 600 types of anthocyanin are reported in nature but cyanidin (magenta/purple - red sweet potato, purple corn), delphinidin (blue-reddish or purple; blue hue of flowers), pelargonidin (red to fruits and berries and orange hue to flowers), peonidin (magenta in berries, grapes, and red wines), malvidin (purple in blue-colored flowers, red to dusty red in wine), and petunidin (dark red or purple in blackcurrants) are the most common distributed in the plants. Cyanidin-3-glucoside is the major one found in most of the plants. They possess antioxidative and antimicrobial activities, improve visual and neurological health, and protect against various noncommunicable diseases through different pathways i.e., free-radical scavenging pathway, cyclooxygenase pathway, mitogen-activated protein kinase pathway, and inflammatory cytokines signaling. Black carrot, red cabbage, purple broccoli, cauliflower, beet root, purple potato, purple amaranth and purple lettuce are rich source of anthocyanins. Improvement of anthocyanin through conventional breeding has been done in carrot (Pusa Asita), red cabbage (Kinner Red), purple heading broccoli (Palam Vichitra). Transgenic approach showed potential to enrich anthocyanin(70-100 fold) in tomato by the use of two transcription factors having fruit specific expression from Delila and Rosea1 isolated from Antirrhinum majus (Maligeppagol et al., 145). In tomato, Aft gene causes higher level of functional flavonoids guercetin and kaempferol in and Ant1 gene encodes a Myb transcription factor and a DNA marker showed Aft trait is encoded by a single locus on chromosome 10 fully associated with Ant1 (Sapir et al., 200).

**Organosulphur compounds:** Thiols are sulphur containing compounds in garlic and cruciferous vegetables. Allyl sulfides in onion and garlic are product of enzyme allicin activity on alliins and lowers bad LDL cholesterol and triglycerides and increase good HDL cholesterol in blood. They have antibacterial, antiviral, antifungal and antirheumatic properties and protect against asthma, pneumonia and gastrointestinal disorders. Allyl sulfides possess antimutagenic and anticarcinogenic properties and provide cardiovascular protection.

**Chlorophylls:** Chlorophyll is the most important plant pigment and a 'real life force' that nature uses to explode plants into greenery. The anti-mutagenic properties of chlorophylls have been demonstrated in various assays, and clearly intake of chlorophyll has potential to act as a chemo-preventive compound in humans. Green leafy vegetables, broccoli, cabbage and Brussels sprout are good source of chlorophyll. Glucosinolates: Glucosinolates play an important role in plant defense for diseases and specialist insects (Agrawal and Kurashige, 2) and they also implicate strong influence on human health. Sulforaphane and iberin show stronger anti-carcinogenic activity than other isothiocynates (Faulkner et al., 64). But, few breakdown products such as cynogenic glucosinolates and progoitrin cause goitrogenic effect and reduce meal palatability (Chandra et al., 26; Cartea et al., 22). High content of gluconapin, progoitrin, glucobrassicin and neoglucobrassicin produces bitter or pungent isothiocynates may affect consumer preference (Schonhof et al., 206). Total glucosinolates content in leaves ranges from 46 to 87 µmol/g dw (Menard et al., 151). They also reported wide variation in individual Glucosinolates, such as Sinigrin (5.7 – 12.9 µmol/g dw), Glucoiberin (0.5 – 6.6 mg/100 g fw), Glucoibervirin (0.6 – 2.9 mg/100 g fw) and Indole (15.2 - 24.9 mg/100 g fw), which are comparable with total glucosinolates (0.6 - 35.6 mg/100 g fw and Glucoraphanin (0.8 - 21.7 µmol/g dw) and Indole (0.4 - 6.2 µmol/g dw) in green broccoli (Kushad et al., 127). Already, CYP79 and CYP83 gene families in Arabidopsis and GS-ELONG, GS OX, GS-ALK and GS-OH loci in different Brassicas have been reported to be involved in glucosinolate biosynthesis. Accumulation of aliphatic glucosinolates is mainly regulated by genetic factors while environment and environment x genotype interaction affects indole glucosinolates in broccoli (Brown et al., 15). Efforts were made to develop lines having high glucoraphanin along with agronomically acceptable traits through conventional breeding (Farnham et al., 63; Sarikamis et al., 201). Indoles also bind to chemical carcinogens and activate detoxification enzymes mostly of gastrointestinal tract.

#### Germplasm evaluation for nutritional traits

Germplasm screening for donor source is prerequisite for breeding programme which can be done by visual observation, organoleptic scoring, measurements and laboratory analyses. Visual in full daylight or incandescent light and organoleptic selection under optimal condition need expertise of breeder and feasible to improve quality up to certain extend as in case of carrot for carotenoids (up to 120 ppm) while measurement and laboratory analyses are main basis for quality traits. Similarly, selection for different situations such as fresh, processing/ canning items and storage life need specific traits and their measurements. Tomato germplasm ranged from 6.64 to 90.37 mg kg<sup>-1</sup> and 25.7 to 329.9 mg kg<sup>-1</sup>, respectively in Onam (Al Said *et al.*, 3), total titrable acidity varied from 0.20 to 0.64%, whereas the TSS ranged from 3.4 to 9.0% (Panthee et al., 170), β-carotene from 0.11 to 7.79  $\mu$ g g<sup>-1</sup> FW), translycopene (0.19 - 193.17 µg g<sup>-1</sup> FW) (Ruggieri et al., 195). In watermelon, lycopene ranges from 10 to 81 µg. g-1 fresh weight (Davis et al., 44). Carrot germplasm also varied for carotenoids (1.25 - 254.3 mg/100g DW) and β-carotene (0.08 - 136.5 mg/100 g DW) (Jourdan et al., 110). Brinjal fruits are good source of anthocyanin and whole fruit contained anthocyanins in the range of 0.55-88.24 mg/100g (Kumari et al., 126). In Chilli, the pungency was ranged from 272897 to 1037305 SHU in C. chinense, 109508 to 487619 SHU in C. frutescens and 0 to 203731 SHU in C. annuum (Sarpras et al., 202). Glucoraphanin is anti-cancer glucosiolates and its concentration was ranged from 0.8 to 21.7 micromol g<sup>-1</sup> DW in broccoli (Kushad et al., 127). In spinach, wide variation has been shown in germplasm for oxalic acid, nitrate, vitamin C, lutein, carotenoid and phenolic content. Oxalate content in spinach was ranged from 5.3% to 11.6% on dry weight basis (Mou, 161). Vegetable amaranth also had variation for nitrate (1.8 to 8.8 g kg<sup>-1</sup>) and oxalate (5.1 to 19.2 g kg<sup>-1</sup>).

# Quality traits and their genetics in common vegetables

In vegetable crops, the quality attributes are grouped as (i) intrinsic guality attributes which are inherent to the product itself and provide stimuli to consumers such as sensory attributes (flavour, taste, appearance, colour, texture and smell) and health attributes which are concerned with nutritional and health-promoting values, and (ii) extrinsic quality attributes are linked to the production method but not a property of the food itself like pesticides, eco-and animal friendliness, packaging materials, processing technology which can influence the purchasing policy of some consumers. The extrinsic quality traits are not directly related to the product performance or core benefit of the product but contribute in general benefit of product and add value. Further, extrinsic guality can be grouped in narrower sense (characteristics that are perceptible through the product itself *i.e.*, packaging, colour, brand name, price, country of origin) and broader sense (characteristics which are conveyed through marketing instruments such as distribution, communication policies, advertising and pricing). The intrinsic quality traits are generally complex in nature, hence, modern high-throughput biochemical and molecular analytical tools and techniques have great potential to handle complex traits with shorten breeding cycles. The complete changes in the intrinsic traits in short period is

because they are largely governed by genotypic factor and production manner. The breeding objectives for quality traits in major vegetable crops are given in Table 4. The quality attributes and their genetics have been summarized crop wise from available literature hereunder and details of genes involved, type of gene action in respect of quality traits in different vegetable crops are given in Table 5.

Tomato: High lycopene, high TSS (5.5°Brix or higher because high TSS gives mores cases of finished products per tonne of raw fruit and, thus, require less energy in concentrates), lower titrable acidity as percentage of citric acid (0.4-0.5 %), carotene, solid/ acid ratio of 15, sugar /acid ratio of 8.5, pH value (for processing  $\leq 4.35$  because longer time required if pH increases) and flavour (2-isobutylthiazole, methyl salicylate and eugenol). Total acidity and pH are not always closely related due to difference in degree of buffering of unknown volatile constituents of pH by other fruit constituents of unknown volatile constituents). Vitamin C ranges from 10-120 mg/100g fw but high Vitamin C is pleotropic on small fruits. Solanum cheesmanii (15%) and S. chmielewiskii (10%) are sources for total solids while S. peruvianum is source of high sugar and vitamin C while S. pimpinellifolium for high acidity gene. Carotene in tomato is governed by og<sup>c</sup> (old gold crimson gene) and hp (high pigment) gene. The og<sup>c</sup> increase lycopene but reduce carotene by 25% while increases total carotenoids and vitamin A by 25-50%. The B-gene increases  $\beta$ -carotene at the expense of lycopene and result orange fruits with 8-10 times higher than red fruited ones. However, pleotropic effect of high ascorbic acid is pleotropic to size of fruit and of hp gene on slow germination and growth, premature defoliation, yield and fruit size.

**Potato**: Processing ability, uniformity of tuber shape (round long) and size, eye depth and distribution, incidence of growth cracks and growth irregularities, skin smoothness, appearance of final product, texture and flavour (glyalkaloids), flesh colour (white, yellow and cream).

**Brinjal**: Fruit colour (purple colour is dominant over green and cream fruit) is complex trait and monogenic, digenic and trigenic 3 complimentary genes (*Pfb, Pfb1* and *Pfb2*), shape (elongated over round and round over oval by 3 or 4 genes), flavour (glyalkaloids – solasonine and saponin). Tigchelaar *et al.* (229) identified nine genes that determine anthocyanin development and distribution in eggplant, three independent complementary factors *D* (four alleles *D* in order of dominance, *d* inhibits anthocyanin in fruit only, *d<sup>t</sup>* in fruits and dilutes flower color and *dw* in all plant parts), *P*, and *Y* cooperate to affect anthocyanin development in the corolla, hypocotyl

Breeding and Genomic Investigations in Vegetable Crops-A review

Table 4. Objectives specific to quality traits for use in breeding of vegetable crops.

Tomato	Fresh market - Appearance : fruit shape (oblong or square round), size or weight (80-90 g), smoothness (without ribs), stylar or blossom end rot smooth without any depression or scar; Fruit
	colour- uniform, red and deep red; Fruit firmness, a desirable trait for good transportability and long shelf life: Pericarp tissues has more sugars than the locular tissue.
	Acid content in locular tissue has predominant influence on fruit flavour.
	Processing traits: High TSS (5.5 °Brix or higher) because high TSS gives mores cases of finished
	Acidity: pH (below 4.35; because longer time required if pH increases) and low titrable acidity (as
	percentage of citric acid) and 0.4-0.5 %
	Solid/acid ratio of 15, sugar /acid ratio of 8.5
	pleiotropic on small fruits)
Chilli	Fresh market: Fruit shape, size, number of lobes (in sweet potato), colour, pungency, flavour, exocarp thickness, endocarp, seed ratio, vitamin A and C.
	Fresh processing (sauce, paste, canning, pickling): fruit colour, pungency, flavour, pericarp thickness, endocarp: seed ratio.
	Dried shape (whole fruit or powder): colour of dried fruits, pungency, flavour, dry weight recovery, low crude fibre, endocarp: seed ratio. Oleoresin extraction – essential oils (colour, pungency).
Brinjal	Fruit shape, size and colour (glossiness)
	Lower solanine content (i.e. 400 mg is harmful), high anthocyanin, chlorogenic acid.
Potato	Physical - shape (round), size (medium), shallow eyes, colour
	Nutritional – dry matter content, reducing sugar, low glycoalkaloid content (20 mg/100g fw), high vitamin C and protein content
	Potato chips: long-term storage, round or oval shape, 40-75 mm in size. Dry matter (21-25%), starch (16-20%), reducing sugar (<0.25%) uniformly distributed in cross-section.
	French fries: Oblong, or oval in shape, size >70 mm. Dry matter 20-23%, 15-18% starch, reducing sugar (0.3%) distributed equally to avoid 'sugar end' effect in fries.
	Processing ability, uniformity of tuber shape (round long) and size, eye depth and distribution, incidence of growth cracks and growth irregularities, skin smoothness, appearance of final product, texture and flavour (glyalkaloids), flesh colour (white, yellow and cream).
	High specific gravity (dry matter content) suitable for French fries, chips and dehydrated products
Cabbage	Head shape spherical or round (polar and equatorial dia. ratio 0.8-1.0) and drum or flat head (p.e. 0.5-0.6), conical head (p.e. 1.0), medium size head (1.0 kg or 0.8-0.9 kg), head compactness, green heads for fresh market and white leaves are preferred for coleslaw. Short core (< 25% of head diameter).
	Narrow and soft core is desirable for processing. Reduced content of goitrogenic glucosinolates and higher content of desirable GLSs for good flavour and anti-carcinogenic effects
Cauliflower	Curd colour, curd compactness, curd depth, curd shape, self-blanching. Lowering of goitrogenic or adverse glusocinolates and increase of health beneficial Glucosinolates
Knol khol	Round, medium-sized knobs of desired colour-green or purple, low fibres and creamy white to greenish white flesh in tubers. Enriching vitamin A and anthocyanin content. Lowering of goitrogenic or adverse glusocinolates and increase of health beneficial glucosinolates
Broccoli	Fine buds, attractive green or purple heads, compact smooth heads, health beneficial Glucosinolates, increasing vitamin C, anthocyanin and vitamin A and minerals.
Carrot	Colour (red or orange), size, length and girth (diameter), shape (cylindrical, uniformly tapering or stump rooted, broad or narrow shoulder), non-branching and non-forking, smooth surface without fibrous in roots, thick flesh, thin self-coloured core, high carotene and high sugar and dry matter in roots, good flavour and taste.

# Indian Journal of Horticulture, March 2020

Table 4 contd...

Beet root	Uniform size, shape and colour, uniformly coloured roots without roots without internal zoning or white rings.
Turnip	Root shape, size, colour of skin and flesh and their uniformity
Radish	Root length, shape and colour, pungency, taste and edible quality, late pithiness, non-forking roots.
Onion	Fresh: Bulb size, shape, colour, pungency, firmness, single centre, rings per bulb, ring thickness, narrow neck, dormancy, dry matter (>15% for white onion for dehydration; 9-13% in onion for fresh consumption). Low solid/high water content is good for fresh use. Dehydration: Snow white colour, globe shaped bulbs, thick neck, free from greening and moulds, high pungency and high TSS (>18%) and low reducing sugar that reduces caking and deterioration of the colour during storage.
Garlic	Bulb size index, bulb colour, clove colour, clove shape (sickle), number of cloves, clove diameter, TSS, dry matter
Leafy vegetables	Attractive leaf colour, succulent stem, less fiborous, crispiness; Higher vitamin A content and folate, good flavour, taste; Plants with lower level of antinutrients (nitrate, oxalate); Optimum leaf/stem ratio (>1.0)
Okra	Pod quality – medium length, not much thick, dry matter and weight and pods, smooth pod surface without much pubescence, number of ridges- preferably five, pod colour – dark green or shining green, and longest keeping quality, lodine content
Cucumber	Fruit colour – light green, green or white depending upon consumer's preference; Shape – uniformly cylindrical fruits without thin or crook neck or bulged blossom end; Size – depending upon consumers' preference and usage, preferably medium-long for slicing and small for pickling Spines- few or none, preferably white; Skin – smooth without warts; Seeds - few and immature at edible stage of fruits. Bitter free fruits
	Edible fruit without carpel separation
Pumpkin	Thick fruit flesh and small seed cavity; Round /oblong/flat round fruit shape; Small size fruits for nuclear families and big fruits for packing foods, orange flesh colour, rich in $\beta$ -carotene.
Muskmelon	Fruit quality- shape, size, epicarp/skin colour and surface (smooth, sutures, netted); Flesh thick and attractive colour; Seed cavity, preferably small; Sweet taste, musky flavour, juiciness and high TSS (Not less than 10%; 11-13 %).Good keeping quality and transportability.
Watermelon	Fruit shape (round/oblong), size/weight (large, medium or small), Skin/rind: (tough, thickness, resistance to cracking, colour green, dark green or light green, striped or without stripes); Intermediate fruit shape advantageous because long fruits are prone to gourd neck fruits and round fruits to 'hollow-heart'; Flesh: firm, colour – attractive red, pink or yellow Seeds: smaller and fewer. TSS (more than 10%), Transportability and shelf-life.
Bitter gourd	Fruit size: ranges from 7-40 cm but medium size fruits (10-15 cm) are preferred; Fruit shape: uniformly medium thickness but thick fruits are required for stuffing; Surface: warty (with tubercles), or smooth or smooth with continuous or broken ridges. Fruits having smooth surface and continuous smooth ridges are preferred in many places. Fruit colour: White, light to dark green. White are preferred in Tamil Nadu, Karnataka and Maharashtra. Green or light green in Bihar, eastern Utter Pradesh and West Bengal and dark green that having long shelf-life/keeping quality are preferred in most places. Less number of mature seeds at the time of marketable stage; Bitterness: small fruited common in West Bengal and Bihar are very bitter. Medium-bitter fruits are more common in many parts of country. Increase antidiabetic constituents.
Sponge gourd and Ridge gourd	Shape: Uniformly cylindrical, medium-long fruits; Colour – whitish green, light green and dark green in sponge gourd and light to dark green in ridge gourd; Size: 25-30 cm in smooth gourd and 30-35 cm in ridge gourd Non-bitter fruits; Tender and non-fibrous fruits at marketable stage

Contd...

Breeding and Genomic Investigations in Vegetable Crops-A review

Table	4	contd

Bottle gourd	Fruit shape varies as long, round, oval, club and dumble shaped; General preference for long, uniformly cylindrical, without neck and medium-long or spherical fruits are preferred; Surface : glossy green surface, pubescent with sparse hairs in immature fruits at marketable stage; Fruits should be non-bitter, non-fibrous flesh and less of immature soft seeds at marketable stage
Garden pea	Shelling percentage (30.7-56.4%); Pod colour, size, apex (blunt or pointed) and shape (straight or curved);
	or wrinkled (26-33% protein) or smooth (22-31% protein);
	Starch content in wrinkle seeded about 46.3%) and sweeter than smooth seeded (33.7%); Smooth seeded have hard texture and less flavour;
	Suitability for canning, dehydration (large sized wrinkled and dark green seeds) and freezing; Low content of antinutrients
Cowpea	Pods stringless, less fibrous, spongy inner pod walls with wide spaced seeds in the pods.
French bean	Fresh market: Pod shape- flat or round; Pod length/size, preferably smooth, stringless, straight, without curve, fleshy; Pod colour- light green, medium green or dark green; Pod wall fibre enough to retain its shape and fresh appearance for 7-10 days after harvest; Pods free from interlocular space and flat pods are preferred for fresh market, seed size/100 seed weight; Number of seeds per pod, seed colour, pod width (8.3-9.5 mm); Preference : High carbohydrate content, a pH of 8-8.5 and an alcohol insoluble solids content <8.5%; High protein content in seeds , Ca and Fe; Low content of antinutrients – tripsin inhibitor; Phytochemicals- L-3,4-dihydroxyphenylalanin (L-DOPA), C-DOPA, flavonoid quercetin, myrcetin, catechin, inositol hexaphosphate (IPC) and inositol pentakisphosphate. For canning – light or medium green, round fleshy, stringless pods with small white seeds, slogging free, short pods for whole pod processing.
	For freezing – Bright green, round or flat pods, stringless, small sized pods for freezing whole pod.

and fruit. Other genes *Ac* (convert delphinidin-3-rhamnoglucoside to nasunin), *Puc* (pigment under the calyx), *Sa* (anthocyanin stripe on the anthers), *Dil*, and *Dil*<sub>2</sub> (affect color intensity) and *R* (inhibits fruit pigmentation in certain genetic backgrounds) genes affects pigmentation in fruits and other parts.

**Chilli**: Fruit size (small>large; 30 genes), fruit shape (oval > elongated; polygenes), pungency (capsaicin by a single dominant gene *C* but degree is influenced by polygenes; high and dry temperature increases capaicin content) and fruit colour are red, brown and green (red > brown, single gene). Role of chlorophyll retainer gene [*cl*(*g*)] plays role in preventing complete degradation of chlorophyll. Chlorophyll and red pigment lycopene (genes *y*<sup>+</sup>) produce brown fruits and genetic constitution of yellow colour (*yycl*<sup>+</sup>*cl*<sup>+</sup>), brown (*y*<sup>+</sup>*y*<sup>+</sup>*clcl*), red (*y*<sup>+</sup>*y*<sup>+</sup>*cl*<sup>+</sup>*cl*<sup>+</sup>) and green (*yyclcl*). Yellow or orange mature fruits colour is dominant trait and governed by *Y* gene. Beta-carotene in fruits is governed by *B* gene which interacts with *t* gene for higher levels.

**Garden pea**: Pea have two main classes of storage proteins as 11S (legumin) and 7S (vicilin, convicilin) in wrinkled (26-33%) and smooth (23-31%) seed types. Sugar is also high in wrinkle seeded (10-

13%) (*RRrbrb*; *rrRbRb*) than round seeded (6-7%) (*RRRbRb*). But, they are low in methionine and also in cystine. *Pisum abyssinicum* and *P. fulvum* are good source of cystine. Inheritance of protein content is complex and polygenic governed trait.

Beans: In French bean, small seed size, dark pod colour, raw-pod quality, pod length, better holding capacity, percentage of pod fibre content and flesh firmness are important traits. Straight, smooth, round, uniform internal and external colour, flavour, texture, carpel separation, skin sloughing, interlocular cavitation, internal tissue breakdown. A single dominant gene is responsible for the inheritance of high Zn content in seeds (Cichy et al., 31). In Faba bean, seed protein, amino acid composition and antinutritive factors are important attributes and they have additive effect with some partial dominance. Larger string free pods, attractive cooking and taste qualities are quality traits in vegetable cowpea. While in Lima bean, Agglutining activity and HCN production are of major concern.

**Watermelon**: Fruit shape is qualitatively inherited trait with one pair of alleles (round: elongated) with modifiers and elongate spherical. Fruit size is governed by polygenes (25 genes) and Poole and Grimbell (181)

reported that 2-3 genes alongwith modifiers influence flesh colour. Red flesh colour (YY) is dominant over yellow (yy), white (WY) over canary yellow (C) and red, golden yellow flesh over red flesh. WR governed orange colour and white is epistatic to yellow. In *C. colosynthesis* x *C. lanatus*, white (Wf) colour is dominant over red flesh (wf) colour. Dark rind colour is dominant over light green or grey colour and stripped over solid light green colour. Total solids content is governed by three incompletely dominant genes. Bitterness in *C. colosyntheisis* is dominant over nonbitterness and governed by a single gene, however,  $Su^{Bi}$  acts suppresser of bitterness.

**Muskmelon**: Yield (earliness and concentration), fruit appearance (shape, size, colour, smoothness, flesh colour), flesh sweetness (TSS >9%), texture, aroma, firmness, colour and rind (hardness, thickness, netting).

**Pointed gourd:** Fruit colour, small seed content, greater pulp content and prolonged shelf-life are important quality traits. While in **bitter gourd** vitamin C, Iron, fruit colour, texture, tuberacles, taste and flavour are breeding targets.

**Leafy vegetables**: Attractive dark colour, high in nutrients such as iron, calcium, beta-carotene and antinutritive traits nitrate, oxalate, phytate and saponin. In coloured vegetables, high intensity of colour is a preferred trait.

**Radish**: Attractive colour, non-pithy, more in sugar and low in pungency (fresh – less; cooked/salted – high), elaboration of the cell, water content, pore extent. In turnip, two independent genes controls purple skin colour and flesh colour is monogenic and white is dominant over yellow.

**Beet root**: Colour is qualitative and quantitative and roots hypocotyls and petioles are red  $(R_/Y_)$ , yellow  $(rrY_)$ , white and red hypocotyls  $(R_yy)$ , yellow hypocotyl and white root (rryy). The *R* locus has five alleles  $(R, R^t, r, R^p \text{ and } R^h)$  and three alleles at *Y* locus  $(Y, Y^r \text{ and } y)$ . The red hypocotyl is  $R^{tt}Y$  and striped petiole is  $R^t$  without Y and  $Y^r$  for pigment only in roots. Quantitative differences in colour of red beet root are determined by the ratio of violet betacyanin to yellow beta-xanthin pigments which is governed by three alleles at *R* locus. High, medium and low ratio have violet, red and orange colour, respectively.

**Cucumber**: Skin colour, spine colour, presence or absence of warts and spines, fruit shape. Flesh firmness, skin tenderness and seed cavity. Bitter free (lacking cucurbitacin) is governed by *bi* gene. Parthenocarpy is governed by *Pc* and yellow green immature fruit colour and yellow flesh by *yg* and *yf* genes, respectively.

**Squash**: Single recessive allele (*n*) determines naked seed but modifying gene may influence it. Softness is

due to inhibition of thickening and lignification of the cell walls of testa. Orange skin and deep orange flesh colour are preferred traits.

**Onion**: Five genes *C*, *R*, *G*, *L*, *I* governed onion bulb colour and of them are, red ( $iiC_R_$ ), yellow ( $iiC_rr$ ), homozygous recessive white ( $iiccR_$ , iiccrr), homozyhous dominant white ( $II_{\_}$ ) and heterozygous dominant white or buff or off-white ( $Ii_{\_}$ ). Pink colour occurs when cross is attempted between two homozygous yellow bulbs which creates bulb colour. Further, two additional genes G (golden bulb) and L also causes variation in onion bulb colour. Golden colour bulbs developed during presence of a dominant allele at *G* locus. Three loci on chromosomes 3, 5 and 8 govern amount of fructans, fructose, glucose and sucrose and *Frc* on chromosome 8 is responsible for fructan content.

**Cabbage**: Red colour in cabbage is governed by several factors and quantitatively inherited. The  $F_1$  cross between green × dark red resulted pink plants. The gene *M* produces magenta and *S* gives purple on upper side of the leaf. Savoy leaf texture is high yielding, contains more dry matter, better flavoured and less gas producing and controlled by three or more genes. Storability and eating are contrasting traits because very late cultivars storage leaves are very hard and tough. Dry matter in early type (6%) is positively correlated with later maturity (10%) and is a quantitatively inherited trait.

Carrot: Important quality attributes are carotenoids, anthocyanins, lutein, fibre, texture, sugars, flavour, minerals and toxicants. Carrots have xylem, phloem and vascular cambium and their colouration is governed by different genes. Predominant colours are orange, red, black and pale for xylem. However, self-core roots are preferred but different colour combinations are made by combining gene combinations. Genes responsible for xylem colour were reported to be O for orange, P-1 and P-2 for purple and y for yellow. Y-1 and Y-2 governed differential xylem/phloem levels. Sugar quantity, total dissolved solids and dry matter content are quantitatively governed traits. A dominant locus Rs controls the ratio of reducing sugar (glucose and fructose) to sucrose, independent of total sugar. Harsh flavour is due to volatile terpenoids and quantitative trait, but mild flavour is dominant.

### High throughput techniques for quality assessment

Measurement of most of the quality traits is laborious, time consuming and costly practice. However, recent developments in tools and techniques in food analysis have opened new arena having high throughput capacity for such analysis which are rapid, cost effective and relatively easy or automated. Breeding and Genomic Investigations in Vegetable Crops-A review

	Quality traits	Number of genes	Type of Gene action	References
Potato	Skin colour	Digenic (D-R-)	Complementary <i>D_R_</i> Red; <i>D_rr</i> , White <i>dd R_</i> White; <i>dd rr</i> White	De Jong (45)
		Three independent loci	D (red), $R$ (tuber-specific regulation) and $P$ (purple)	Van Eck <i>et al</i> . (239)
		Single dominant gene	S. tuberosum anthocyanin 2 (Stan2) is an $R_2R_3$ MYB domain gene in the anthocyanin pathway that co-segregates with the D locus and is associated with the synthesis of red and purple anthocyanin pigments	Jung <i>et al</i> . (112)
		Single recessive gene	Dihydroflavonol 4-reductase ( $dfr$ ) co-segregates with the $R$ locus for red pigmentation	Zhang <i>et al</i> . (268)
		Single recessive gene	Flavonoid 3',5'-hydroxylase (f3'5'h), co-segregates with the $P$ locus	Jung <i>et al</i> . (111)
	Flesh colour	Monogenic	Colour is incompletely dominant to white, the intermediate is pale yellow	Jung <i>et al</i> . (112)
		Single dominant	Zeaxanthin accumulation (yellow flesh) <i>Chy2</i> gene	Wolters et al. (255)
	Tuber shape		Long tuber axis is dominant to short axis	De Jong (45)
Brinjal	Fruit colour	3 genes ( <i>P, X,</i> <i>Puc</i> )	Purple dominant to green	Thakur <i>et al</i> . (226)
			Green ( <i>Gm</i> ) incompletely dominant over white ( <i>gm</i> )	Choudhury (30)
			Green streaked purple > greenish white	More <i>et al</i> . (158)
		Three complementary genes	Pfa, $Pfb_1$ and $Pfb_2$ ; purple > green	Patil and More (175)
	Fruit shape	Monogenic	Round shape partially dominant to long ( <i>Ofa, Ofb1, Ofb2</i> and <i>Ofb3</i> )	Patidar (173)
	Fruit size	Monogenic	Large fruit size partially dominant to small fruit size	Patil and Moore (174)
Tomato	High B-carotene; low lycopene	Monogenic	<i>B</i> (high concentrations of b-carotene in orange-pigmented) <i>but</i> subject to influence by a modifier gene, $MoB$ , which segregated independently of <i>B</i>	Tomes <i>et al.</i> (231)
Chilli	Fruit colour	Three independent genes	<i>yy cl⁺cl⁺</i> yellow; <i>y⁺y⁺cl⁺cl⁺</i> Red; <i>y⁺y⁺clcl</i> brown; <i>yy clcl</i> green	Kormos and Kormos (124)
		Partial dominance	Bright red colour and purple colour over green	Ahmed <i>et al</i> . (1984)
	Capsanthin content	Monogenic dominant	С	Lippert (138)
	Pungency	Monogenic	Pungency dominant to non	Dempsey (49)
Garden Pea	Pod colour	Monogenic	Yellow ( <i>Gp</i> ); Blue-green ( <i>Dp</i> ); Purple ( <i>Pu</i> , <i>Pur</i> )	Swarup (222)

Table 5. Classical studies on Genetics of quality traits in different vegetable crops.

Contd...

# Indian Journal of Horticulture, March 2020

	Quality traits	Number of genes	Type of Gene action	References
Turnip	Flesh colour	Monogenic	White dominant to yellow	Davey (43)
	Skin colour	Two independent loci	Both dominant genes give purple colours	Brar <i>et al</i> . (14)
Beetroot	Root colour	Digenic	Red is dominant to yellow or white. Intensity of red colour is influenced by minor genes. Further, 5 alleles at R locus: <i>R</i> , <i>Rt</i> , <i>r</i> , <i>Rp</i> and <i>Rh</i> 3 alleles at Y locus: <i>Y</i> , <i>Yr</i> and <i>y</i> $R_Y$ : Red roots, hypocotyls and petioles RrY - : Yellow roots, petioles and hypocotyl $R_yy$ : White roots with red hypocotyls <i>Rryy</i> : White roots and yellow hypocotyls	Watson and Gabelman (246); Keller (115); Pederson (176)
Carrot	Root colour	Digenic <i>P-Y-</i> and two modified <i>E_</i> & <i>I_</i>	Deep purple : <i>iiPPYYEE</i> ; Purple : <i>iiPPYYee</i> Diffused purple : <i>iiPPyyee</i> ; Yellow : <i>lippYYee</i> Red : <i>iiPPyyEE</i> ; Light red / orange : <i>iippyyee</i>	Kust (128); Buishand and Gabelman (17)
	Nutritional	Monogenic dominant	A ( $\alpha$ -carotene accumulation), <i>Io</i> (intense orange xylem, which may be an allelic form of A), $L_1$ and $L_2$ (lycopene accumulation), O (orange xylem, which may also be an allelic form of A)	Buishand and Gabelman (16)
	Root shape	Three gene ( <i>D,N,P</i> )	<ol> <li>Long or Desi : Long and tapay, <i>D-N-P</i></li> <li>Cylindrical type : Cylindrical, <i>dd</i>, <i>nn</i>, <i>p</i></li> <li>Chantenay type : Obvate / cuneate root, <i>dd</i>, <i>N-P</i>-</li> <li>Round type : Round root, <i>dd</i>,<i>N-</i>, <i>Pp</i></li> </ol>	Frimmel (72)
Radish	Root colour	3 genes and complimentary gene action	Red : $R_2r_2/R_3r_3$ /cc White : $r_2r_2/r_3r_3$ /CC Purple : $r_2r_2/R_3r_3$ /cc $R_2r_2/r_3r_3$ /Cc $R_2R_2/R_3R_3$ /CC : lethal and can not survive $R_2r_3r_3r_3$ /cc : double recessive could be lethal	Yi <i>et al.</i> (260); Tatsuzawa <i>et al.</i> , (225); Tatebe (224); Uphof (238)
	Root shape	Quantitative genes	Many genes	Tsuro <i>et al.</i> (234); Iwata <i>et al.</i> (103); Zaki <i>et al.</i> (261)
Beetroot	Root colour	Two independent loci	<i>R</i> locus has five alleles, viz., <i>R</i> , <i>Rt</i> , <i>r</i> , <i>Rp</i> , and <i>Rh</i> , whereas Y locus had three alleles, viz., <i>Y</i> , <i>Yr</i> , and <i>y</i>	Kellar (147); Watson and Gableman (246), Pederson (176)
Cauliflower	Curd colour	Monogenic	Orange (beta-carotene) dominant over white curd	Li <i>et al</i> . (134)
Cabbage	Leaf colour	Monogenic	Anthocyanin (A) development several alleles	Kwan (129); Magruder and Myers (144)
Cucumber	Fruit colour (ripe)	Digenic (R-C-)	<i>R</i> - <i>C</i> - (9) Red; <i>R</i> - <i>cc</i> (3) Orange; <i>rrC</i> - (3) Yellow; <i>rrcc</i> (1) Creamy	Hutchins (99)
	Flesh colour (ripe)	Digenic (V-W-)	<i>V-W-</i> (9) Diggy White; <i>V-ww</i> (3) Intense White <i>vv W-</i> (3) Intense Yellow: <i>vvww</i> (1) Orange	Kooistra (122)

Table 5 contd...

Contd...

Breeding and Genomic Investigations in Vegetable Crops-A review

Table 5 contd...

	Quality traits	Number of genes	Type of Gene action	References
Muskmelon	Flesh colour	Digenic	<i>gf::</i> Green flesh colour wf:: white flesh	Hughes(96); Clayberg (33); Imam <i>et al</i> . (101)
	Carotenoids	Monogenic	CmOr	Tzuri <i>et al</i> . (235)
Watermelon	Flesh colour	Monogenic	Wf – Y – White flesh Wfwfy – Red flesh Wfwfyy – Ref flesh C-Canary yellow B-Yellow flesh Y-coral red flesh y <sup>o</sup> - orange flesh y-salmon yellow	Wehner (247); Bang <i>et al.</i> (7)
Okra	Pod colour	Monogenic	White dominant to green	Jasim (104); Kolhe and D'Cruz (1121)
	Pod shape	Oligogenic	Angular dominant over round	Jasim and Fontenot (105)
Onion	Bulb colour	Three loci	<ul> <li><i>II</i> – Inhibitory gene – white colour</li> <li><i>Ii</i> – Off white or buff coloured bulb</li> <li><i>ii</i> – Other colour genes express</li> <li><i>iicc</i> – White</li> <li><i>iiC-R</i>- : Red colour</li> <li><i>iiC-rr</i> –: Yellow colour</li> </ul>	Rieman (190) Clarke <i>et al.</i> (32); El-Shafie and Davis (57); Jones and Peterson (109)

Few of them for phytochemicals/nutraceuticals are High performance liquid chromatography (HPLC) for non-volatile samples, Ultra Performance Convergence Chromatography (UPC<sup>2</sup>) for both qualitative and quantitative analysis of gas-phase, Ultra-performance convergence chromatography with a quadrupole time-of-flight mass spectrometry (UPCC-QTOF MS), Waters ACQUITY UPC<sup>2</sup> System for hydrophobic and chiral compounds, lipids, thermally-labile samples and polymers, Spin trapping technique for detection of short-lived free radicals, Gas chromatography (GC) for analyzing volatile compounds which vaporize without decomposition, Gas Chromatography-Mass Spectrometry (GC-MS) for identification and quantification of organic substances in compound matrices, Ultra-high performance liquid chromatography-triple quadrupole/ linear ion trap tandem mass spectrometry (UHPLC-QTRAP/MS/MS) for both small and large molecule analyses. Similarly for minerals, flame photometer, atomic absorption spectroscopy (AAS) for minerals, synchrotron radiation X-ray fluorescence (SXRF) for trace elements, chemical states and surface analyses, inductively coupled plasma-mass-spectrometry (ICP-MS) for low to ultra-los concentration of elements are

useful developments. However, these instruments are costly and technical demanding.

### Breeding vegetables for quality traits

While designing breeding for guality traits, the following points are necessary to consider: (i) source germplasm must have high density of target nutrient(s), (ii) biofortified variety must have wider adaptability with high content of target nutrients, (iii) processed forms of variety/hybrid must retain sufficient quality attribute i.e., nutrients, antoxidants, (iv) bioavailability of nutrients in human body must be high, and (v) acceptance of biofortified crops by target groups i.e., farmers, consumers. The flow chart for quality breeding is given in Fig. 2. Through conventional breeding, large numbers of varieties have been developed in different vegetable crops and some of them are rich in dietary minerals and nutraceuticals. Pigments present in vegetable varieties can be extracted and used as natural dyes, edible food colour and also for functional food development. A comprehensive breeding programme for quality improvement of vegetable need to involve evaluation of germplasm and identification of potential donors, pre-breeding and product enhancement to develop germplasm combining one or more micronutrients, transgressive segregation or heterosis for quality traits, search for promoters for increased bioavailability and reduced anti-nutrients in biofortified varieties, enhancing the density of desirable nutrients, bioactives, molecular breeding for handling complex quality traits and exploring gene editing approaches such as transgenics, RNA interference and CRISPR/Cas9.

### **Biofortification for quality traits**

It is a process of enrichment of health beneficial dietary nutrients in crop through conventional and molecular breeding, genetic and agronomic measures. Biofortification gives opportunity for on-site production of nutrient dense perishable



Fig. 1. Important steps in breeding for quality traits in vegetable crops. MAS- Marker assisted breeding, CBC- Conventional backcrossing, MABC –Marker assisted backcrossing, MAGP- Marker assisted gene pyramiding, MAGI- Marker assisted gene introgression.

Fig. 2. Important steps in breeding for quality traits in vegetable crops. MAS- Marker assisted breeding, CBC-Conventional backcrossing, MABC –Marker assisted backcrossing, MAGP- Marker assisted gene pyramiding, MAGI- Marker assisted gene introgression.

vegetables to serve target population in sustainable manner and to use the extra produce for extraction of desirable compound in industry related to pharmaceutical, cosmeceutical etc. Since 2003, HarvestPlus and its partners have demonstrated that this agriculture-based method of addressing micronutrient deficiency through plant breeding works well. Biofortification has been successfully attempted for vitamin A in vegetables such as sweet potato, cassava, sweet corn, tomato and cauliflower; for iron, millions of people in developing countries are now growing and consuming biofortified crops (Bouis and Saltzman, 13). They suggested three key challenges to reach one billion people by 2030: 1) Mainstreaming biofortified traits into public plant breeding programmes; 2) Building consumer demand, and 3) Integrating biofortification into public and private policies, programmes, and investments. However, long-term cost-effectiveness and its ability to reach rural population make it preferred choice to fight against micronutrient malnutrition.

## a) Biofortification for minerals in crops

Agronomic biofortification for minerals is effective through optimizing the application of mineral fertilizers and/or improving the solubilisation and mobilization of mineral elements in soil (Golubkina et al., 77). But, the extent of increase in mineral content varies with crop variety, soil and climatic condition and amount of nutrient supplied. Genetic biofortification is more stable and acceptable which can be attained through targeting genotypes and plant tissues. Thereby, enrichment of minerals in plants is possible by overcoming genetic barriers for metal accumulation in edible portion. Additionally, increase in concentrations of 'promoter' substances (vitamin C, β-carotene, cysteine-rich polypeptides and certain organic and amino acids which stimulate the absorption of minerals by the gut) and eliminate antinutrients (oxalate, tannins, phytate) which interfere with their absorption. The barriers for metal accumulation largely regulate absorption, transportation (xylem/ phloem loading) and redistribution to sink. In rhizosphere, absorption factors need changes in root morphology (i.e., primary root length and angle; lateral root length, number, density and diameter and root hairs length and density) to increase surface area and root-cell processes to augment solubility and movement of minerals (Welch, 248). Here, root-cell efflux of H<sup>+</sup>, metal-complexes and redox potential are crucial factors. Active and specific transporters and ion channels in rootcell plasma membrane also regulates absorption

pattern of minerals once they enter the apoplasm of root cells. Phloem/xylem loading and translocation factor their accumulation in edible plant organ. It is essential to decipher the molecular and physiological processes for minerals such as proteins for metal homeostasis and hyperaccumulation, transporters and candidates for cytoplasmic metal influx in roots, ATP powered ion pumps (H<sup>+</sup>-ATPases genes), ion channel proteins (aquaporins) and cotransporters in crop plants (Dunlop and Phung, 55; Grennan, 85), particularly vegetable crops where most of the crops have non-seed tissues as edible portion. This can be done through characterization and exploitation of genetic variation or gene discovery and directed gene modification. Genetic engineering led to overexpression of Ca2+/H+ antiporters (cation exchanger 1 antiporter- sCAX1) located in the vacuolar membrane to increase Ca content of transgenic crops like potato (Park et al., 172), carrots (Jeong and Guerinot, 107) and lettuce leaves (Park et al., 171). Besides, retention during processing and bioavailability in target population are important considerations.

#### b) Advanced approaches for quality improvement

To intensify the biofortification process in vegetables, the available opportunities in molecular breeding and related techniques such as genomics by sequencing (GBS), Genome Wide Association Study (GWAS) and single nucleotide polymorphism (SNPs) and techniques targeting genomic sequences such as RNA interference (RNAi), DNA-directed RNA interference (ddRNAi), CRISPR-associated protein-9 nuclease (CRISPR/Cas9), Targeting Induced Local Lesions in Genomes (TILLING) etc. can be explored. Use of TILLING for quality improvement in different vegetables is listed in Table 6. Okabe et al. (166) developed Micro-Tom TILLING platform and identified two allelic mutants of SIETR1 (Sletr-1-1 and Sletr-2) for reduced ethylene responses in tomato.

#### Molecular breeding for quality traits

Molecular markers enable unambiguous identification of lines/individuals in segregating population for handling of quality traits which otherwise difficult to phenotype due to want of technical expertise and sophisticated equipment. Generally, quality traits are outcome of different complex pathways and governed by quantitative traits with environmental influence. A number of genes governing quality attributes in vegetables are mapped using molecular markers (Table 7). A spontaneous '*Or*' mutant gene accumulates  $\beta$ -carotene in cauliflower (Li *et al.*,

Crop	Trait	Allele/Locus/Candidate gene	Method	Author
Tomato	Fruit shelf life	PG	TILLING	McCallum et al. (147)
	Fruit quality	Exp1	CSCE	Gady et al. (73)
	Fruit quality	PG, RIN, Gr, Rab11a, Exp1, Lcy-b, Lcy-e	TILLING	Minoia <i>et al.</i> (153)
	Fruit quality	PSY, Sus2	HRM	Gady et al. (74)
	Fruit quality	Exp1	TILLING	Colbert et al. (36)
	Fruit quality	10 genes	TILLING	Okabe <i>et al.</i> (208)
	Fruit quality	TBG4	TILLING	Hurst <i>et al.</i> (98)
	β-carotene synthase	Psy1	TILLING	Gady et al. (89)
Potato	Starch quality	Waxy	Sequencing	Muth <i>et al</i> . (163)
	Tuber colour	bch, dfr, f3'5'h	HRM	De Koeyer et al. (48)
Cassava	Starch quality	Waxy	TILLING	Tofino et al. (230)
French bean	Storage defense proteins	Lectin locus	TILLING	Lioi <i>et al.</i> (137)
Pea	Trypsin inhibitors	TI1	TILLING	Sonnante et al. (216)
Melon	Fruit quality	ACO1,PDS, DET, DHS	TILLING	Gonzalez et al. (78)
	Fruit shelf life	ACO1	TILLING	Dahmani-Mardas et al. (41)

 Table 6. TILLING approaches for different vegetable quality traits.

169). The introgression of this 'Or' gene into Indian cauliflower will lead to development of β-carotene rich cauliflower. Kalia et al. (114) developed Pusa KesariVitA-1 and promising introgression lines in cauliflower rich in beta-carotene content (8-20 ppm) in Indian cauliflower. Zhang et al. (263) found SCAR markers linked to "or" gene inducing betacarotene accumulation in Chinese cabbage. Zou et al. (272) performed fine mapping of or gene and identified BrPro1 molecular marker in the promoter region of Bra031539 (predicted to encode CRTISO, a carotenoid isomerase specifically required for carotenoid biosynthesis) that can be used for early identification of orange head materials. The eyeappealing orange cauliflower was first discovered in Bradford Marsh, Ontario, Canada in 1970. The orange cauliflower results from a spontaneous mutation of a single dominant gene designated as 'Or' for orange gene. This Or mutant was originally found in white curded autumn crop cv. Extra Early snowball. This trait is absent in Indian cauliflower where large population is suffering from carotene deficiency, however, biofortification of Indian cauliflower with beta-carotene enhancing native 'Or' gene to benefit Indian population.

**Genomics by sequencing (GBS):** The GBS is a simple, affordable and robust procedure for SNP discovery and mapping. This approach is suitable for population studies, germplasm characterization, breeding, and trait mapping in diverse organisms.

This procedure, which can be generalized to any species at a low per-sample cost, is based on high-throughput, next-generation sequencing of genomic subsets targeted by restriction enzymes (Elshire *et al.*, 58). Pereira *et al.* (177) identified 33 QTLs controlling fruit quality traits such as sugar and carotenoid content, fruit and seed morphology and major loci controlling external colour of immature fruit and mottled rind in melon using a RIL population and a GBS-based genetic map.

Genome Wide Association Study (GWAS): GWAS is also known as whole genome association study in which genetic variants in the entire genome are investigated in different individuals to see if any variant is associated with a particular trait. GWAS typically focuses on associations between single-nucleotide polymorphisms (SNPs) and traits. The GWAS have developed into a valuable approach for identifying the genetic basis of phenotypic variation. It has been used for understanding association between genetic variation and target trait. Sauvage et al. (204) performed GWAS using core collection of 163 tomato accessions composed of S. lycopersicum, S. lycopersicum var. cerasiforme, and Solanum pimpinellifolium to map loci controlling variation in fruit metabolites. They reported a total of 44 loci which were significantly associated with a total of 19 traits, including sucrose, ascorbate, malate, and citrate levels. The GWAS also found helpful in studying the evolution of different fruit quality traits (fruit shape, sweetness, flesh colour) in watermelon Breeding and Genomic Investigations in Vegetable Crops-A review

Quality	Traits	Gene/QTL	References
Tomato	Fruit weight	Fw2.2	Cong and Tanksley (38)
	Fruit shape	fas (fasciated)	Cong et al. (39)
	Fruit shape	o (ovate)	Liu <i>et al.</i> (139)
	Fruit shape	SUN	Xiao et al. (258)
	Sugar content	Lin5 (increased sugar content)	Fridman <i>et al</i> . (71)
	Vitamin C	Vtc9.1 (Higher vitamin C)	Stevens et al. (217)
	Shelf life	<i>Rin-</i> inhibited ripening (semi-dominant)	Vrebalov <i>et al</i> . (241)
		nor (non ripening)inhibited ripening (semi-dominant)	Moore <i>et al.</i> (157)
		<i>Nr (Never-ripe)</i> - inhibited ripening (dominant)	Wilkinson <i>et al.</i> (252)
		<i>Cnr</i> (Colorless non-ripening)- inhibited ripening (dominant)	Thompson <i>et al</i> . (227); Seymour <i>et al</i> . (207)
	Fruit colour/Carotenoids	B (Beta)-Yellow fruits	Ronen <i>et al</i> . (193)
		<i>og<sup>c</sup> (old gold-crimson)-</i> higher lycopene content	Ronen <i>et al</i> . (193)
		Del (Delta)-Orange fruits	Ronen <i>et al</i> . (194)
		r (yellow flesh)-Yellow fruits	Fray and Grierson (70)
		t (tangerine)-Orange fruits	Isaacson et al. (102)
		hp-2 (high pigment)- higher lycopene content	Mustilli <i>et al.</i> (162)
		Dg (dark green)-Higher lycopene content	Levin <i>et al.</i> (132)
		y-Uncoloured epidermis	Adato et al. (1)
		Apricot (at)	Jenkins and McKinney (106)
	Anthocyanins	Anthocyanin fruit (Aft)- anthocyanin in skin and outer pericarp	Giorgiev (76); Jones et al. (108)
		Atroviolacium (atv)	Clayberg (34)
		Aubergine (Abg)	Rick et al. (189)
Chilli	Pungency (capsaicin)	Сар, сар3.1, сар4.2, сар7.1, сар7.2	Ben Chaim et al. (9)
		Pun1	Stewart et al. (219)
		С	Blum <i>et al</i> . (10)
	Fruit shape	fs3.1, fs8.1, fs10.1 fs1.1, fs3.1, fs4.1 fs10.1	Rao <i>et al</i> . (186)
	Fruit weight	fw2.1, fw3.1, fw3.2, fw4.1, fw8.1, fw1.1, fw2.1, fw3.1, fw4.1, fw8.1, fw10.1, fw11.2	Rao <i>et al</i> . (186)
	Pericarp thickness	pt 3.1, pt4.1, pt8.1, pt10.1 perwd1.1, perwd3.1, perwd3.2, perwd6.1, perwd8.1, perwd11.1	Rao <i>et al.</i> (186)

Table 7. Gene and QTL mapping for improving vegetable quality.

Contd...

Quality	Traits	Gene/QTL	References
	Fruit Colour	Y-Yellow fruit colour	Lefebvre et al. (131)
		C2-Orange fruit colour	Huh <i>et al</i> . (97)
		Chlorophyll retainer (cl) –Brown fruits	Borovsky and Paran (12)
		A-Purple fruit colour	Borovsky <i>et al</i> . (11)
	Soft flesh and deciduous fruit	S	Rao and Paran (185)
Brinjal	Fruit weight	fw2.1, fw9.1, fw11.1	Frary <i>et al</i> . (68)
	Fruit shape	fl2.1-Ovate	Ku <i>et al</i> . (125)
		fs7.1, ovs4.1	Grandillo et al. (83)
	Anthocyanin	fap10.1	De Jong et al. (47)
	Fruit stripe	fst4.1	Grandillo and Tanksley (84)
	Fruit glossiness	fglo1.1, fglo6.1, fglo8.1, fglo9.1, fglo12.1	Frary <i>et al</i> . (68)
	Parthenocarpy	Cop3.1, Cop8.1	Miyatake <i>et al</i> . (155)
Onion	Carbohydrate	Frc 8 (Fructan content)	McCallum <i>et al</i> . (147)
	Bulb color	P (Pink colour)	Kim <i>et al</i> . (117)
		crb7-complementry recessive red	King <i>et al</i> . (118)
Cauliflower	Curd colour	β-carotene accumulation/Or gene	Li and Garvin (133)
		Pr-High anthocyanin content	Chiu <i>et al</i> . (32)
Cabbage	Head type	Glossy foliage (gl-1)	Kianian and Quiros (116)
	Head shape	Htd 3.1, Htd 8.1	Pang <i>et al</i> .(169)
Kale	Leaf colour	BoPr-Purple leaf	Liu <i>et al.</i> (140)
Beetroot	Sucrose content	13QTL	Trebbi (233)
Carrot	β-carotene	8QTL	Santos and Simon (198)
	δ-carotene	4QTL	Santos and Simon (199)
	Carotenoids	PSY	Santos et al. (199)
Watermelon	Lycopene	LCYB	Bang <i>et al</i> . (8)

Table 7 contd...

(Gou et al., 81) and draft genome sequence also open new area for molecular studies (Gou et al., 88). Colonna et al. (37) analyzed genomic diversity and novel genome wide association in Capsicum and identified four novel loci associated with the phenotypes determining the fruit shape, including a non-synonymous mutation in the gene Longifolia 1-like (CA03g16080). In tomato, Zhao et al. (269) performed GWAS using 775 tomato accessions and discovered 305 significant associations for sugars, acids, amino acids, and flavour-related volatiles. The potential of GWAS and genomic prediction for improving curdrelated traits in cauliflower was explored by Thorwarth et al. (228) using 174 accessions and identified 24 significant associations for curd-related traits. Accumulated capsaicinoid content and increased fruit size are important quality traits in Capsicum

annuum. Nimmakayala et al. (165) identified genomic segments linked to various fruit traits and capsaicin accumulation in *C. annuum* and generated 66,960 SNPs using GBS and reported SNPs in Ankyrin-like protein, IKI3 family protein, ABC transporter G family and pentatricopeptide repeat protein as markers for capsaicinoids.

**Transgenic approach:** Transgenic crops, commonly known as genetically modified (GM) crops enable plant breeders to bring favourable genes, often previously inaccessible, into elite cultivars, improving their value considerably. With the development of genetics and molecular biology, a large number of quality related genes such as those involved in pigmentation, biosynthesis of vitamins, minerals and flavour compounds, soluble carbohydrate metabolism, fruit colour, shape, size and position,

cell wall metabolism, shelf life have been identified in different vegetable crops. Genetic engineering enabled vegetable breeders to incorporate desired transgenes into elite cultivars, thereby improving their value, nutritional quality and other health benefits. Dias and Ortiz (52) reviewed recent advances in transgenic vegetable crops. This approach is more efficient and precise to incorporate genomic region which manifest desired improvement in target quality traits. Zinc fortified transgenic lettuce can be used to overcome its deficiency that severely impairs organ function. Transgenic lettuce with improved tocopherol and resveratrol composition and transgenic tomatoes with high folate levels are promising developments against deficiencies (Dias and Ortiz, 51; Diaz de la Garza et al. 53). Further, cyanide-free transgenic cultivars of cassava can be promising option to provide safe cassava (Siritunga and Sayre, 214). Romer et al. (192) developed transgenic tomato to enhance the carotenoid content and profile of tomato fruit. It has contributed to increase in β-carotene content about threefold, up to 45% of the total carotenoid content in cultivar "Ailsa Cray". Park et al. (171) demonstrated that lettuce expressing the deregulated Arabidopsis  $H^+/Ca^{2+}$  transporter *sCAX1* (cation exchanger 1) contained 25–32% more calcium than controls. Lu et al. (143) suggested that transgenic cauliflower with Or transgenesis associated with acellular process that triggers the differentiation of proplastids or other noncoloured plastids into chromoplasts for carotenoids accumulation. Wahlroos et al. (242) produced oilseed Brassica rapa with increased histidine content. Transgenic approach also helped in extending the shelflife of vegetable crops. For example, cytokinins are known to delay floral yellowing of plants and Chan et al. (25) used a transgene construct pSG766A to increase expression of isopentenyl transferase (key enzyme for cytokinin synthesis) in broccoli. The ipt transgene is triggered by the senescence-associated gene promoter (SAG-13) and this resulted into extending shelf-life to 7.5 - 8.5 d compared with 5.6 d for the nontransgenic line.

**RNA interference (RNAi) in quality breeding:** The RNA silencing is a gene regulatory mechanism that limits the transcript level by either suppressing transcription (TGS) or by activating a sequencespecific RNA degradation process [PTGS/RNA interference (RNAi)]. This approach was effectively used to alter the gene expressions for improving quality traits by increasing antioxidants in tomatoes or suppressing over expression of negative traits such as sinapate esters in canola and alpha-linolenic acid in soybean. Eady *et al.* (56) suppressed the lachrymatory factor synthase gene using RNAi

silencing and reduced lachrymatory synthase activity manifold. This silencing had shifted the trans-S-1propenyl-l-cysteine sulfoxide breakdown pathway so that more 1-propenyl sulfenic acid was converted into di-1-propenyl thiosulfinate which resulted into a marked increase in usually trace or nondetectable non-enzymatically produced zwiebelane isomer and other volatile sulfur compounds, di-1propenyl disulfide and 2-mercapto-3,4-dimethyl-2,3-dihydrothiophene.Further, Meli et al. (149) also used RNAi approach to extend shelf-life of tomato by blocking the expression of ACC oxidase gene and suppression of two ripening specific N-glycoprotein modifying enzymes,  $\alpha$ -mannosidase and  $\beta$ -D-Nacytlhexosaminidase (β-Hex). Targeting of three homologs of ripening genes 1-aminocyclopropane-1-carboxylate synthase (ACS) using chimeric RNAi-ACS construct also resulted into delayed ripening of fruits up to 45 days (Gupta et al., 89). Peters et al. (178) used RNAi technology to develop Dau c 1.01 and Dau c 1.02-silenced transgenic carrot plants show reduced allergenicity to patients with carrot allergy. The RNAi-mediated suppression of DET1 expression under fruit-specific promoters has recently shown to improve carotenoid and flavonoid levels in tomato fruits with minimal effects on plant growth (Williams et al., 253). Seedlessness is desirable factor and auxin and gibberellins are associated with parthenocarpy and suppression of auxin response factor 8 using RNAi resulted parthenocarpic fruits in tomato (De Jong et al., 46). Such fruits were also obtained by suppression of genes of the AUCSIA family coding for 53-amino-acid-long (protein or peptide) by RNAi (Molesini et al., 156).

#### CRISPR/Cas system in quality breeding

Clustered regularly interspaced short palindromic repeats (CRISPR)/CRISPR-associated protein 9 (Cas9) system is an efficient genome editing tool. It allows easy alteration in DNA sequences to modify gene function for obtaining desirable traits in crop plants as well. Wang et al. (244) reviewed use of CRISPR/cas in improvement of tomato attempted by different researchers through targeting a number of genes such as CLV3 and Ic (Fruits with increasing locule numbers) (Rodriguez-Leal et al., 199), PSY1 (vellow coloured fruits) (Filler et al., 66), MYB12 (pink colour fruits) and ANT2 (gene insertion) (purple colour fruits) (Deng et al., 50), Pl and ALC (long shelf life) (Uluisik et al., 237), MPK20 (Repression of genes controlling sugar metabolism) (Chen et al., 27) and SGR1, LCY-E, Blc, LCY-B1, LCY-B2 for increase in lycopene content (Li et al., 136).

Different genes of anthocyanin and carotenoid biosynthetic pathways including *Anthocyanin 1* 

(ANT1) (Cermak et al., 24), Phytoene desaturase (SIPDS), Phytochrome interacting factor (SIPIF4) (Pan et al., 167), and Phytoene synthase (PSY1) (Hayut et al., 92) have also been edited by CRISPR/ Cas9 in tomato. Parthenocarpy is an important trait in tomato with huge potential in processing industry and the trait has been obtained by editing two different genes namely knocking down Slagamous-like 6 (SIAGL6) gene (Klap et al. (119) and by mutating SIIAA9 gene (Ueta et al., 236). These reports in tomato open new opportunities for developing parthenocarpic fruits in other horticultural crops like watermelon, pointed gourd, bitter gourd, etc. where seedless or less seeded fruits are in demand.

There are few reports in potato which establish that CRISPR/Cas9 can be effectively utilized for multi-allelic mutagenesis in polyploid crops. Starch quality of potato is an important trait. Hexaploid potato containing only amylopectin was developed by mutating granule bound starch synthase (GBSS) gene using CRSIPR/Cas9 by Andersson et al. (5). Similarly ACETOLACTATE SYNTHASE1 (StALS1) gene has also been mutated in potato (Butler et al., 18).

Multiplexed CRISPR/Cas9 system has also been attempted to obtain knockout lines of multiple genes in a single transformation experiment, such experiments have application in metabolic engineering of vegetables. Li *et al.* (135) simultaneously targeted five key genes of  $\gamma$ -aminobutyric acid (GABA) shunt in tomato and significantly enhanced GABA accumulation in both the leaves and fruits.

#### Biosynthetic pathway analysis in vegetable crops

In different vegetable crops various studies have been carried out on the expression profiling of genes involved in different nutrient biosynthetic pathways. These studies carried out on different tissues under various conditions or in different genotypes or in the comparison of mutants and wild-type have led to the identification of candidate genes linked to various nutritional traits.

Glucosinolates (GSL) are the most studied secondary metabolites in brassica crops (Ramchiary *et al.*, 183; Feng *et al.*, 65). Twenty five putative GSL biosynthetic and degradation genes have been identified in broccoli through Transcriptome RNAseq analysis (Gao *et al.*, 75). In a separate study by Guo *et al.* (87) transcriptomes of high-GSL and low-GSL *Brassica alboglabra* sprouts were compared by RNA-seq analysis which revealed that level of myrosinase in high-GSL material was significantly lower than that of lower-GSL one. Such studies have also shown that the GSL profile of *B. oleracea* has undergone extensive variations compared with that of *B. rapa* as the type of major GSL vary in both these spp. As gluconapin and glucobrassicanapin are the major GSLs in *B. rapa* (Cartea *et al.*, 24; Wiesner *et al.*, 251), glucoraphanin, gluconapin, progoitrin, and sinigrin in *B. oleracea* whereas broccoli (*B. oleracea* var. *italica*) has a higher level of the anticancer precursor glucoraphanin compared with the other vegetable Brassicas (Sarikamis *et al.* 201).

Vegetables get their red, orange, and yellow colour due to the presence of phytonutrients called carotenoids. Vegetables contain different carotenoids such as beta-carotene, lutein, zeaxanthin, neoxanthin, violaxanthin, and folate, which have important antioxidant, anticancer and pro-vitaminA properties (Kopsell and Kopsell, 126).

The Or gene that is responsible for the orange colour has been extensively studied in relation to its function in the carotenoid biosynthesis pathway in Brassicas (Lu et al., 143; Zhou et al., 270; Zhang et al., 265). Zhang et al. (267) reported that the orange head phenotype of the Br-or mutant is attributable to the insertion of a large genomic fragment at the 3' end of the *BrCRTISO* gene and the differential expression of a number of transcription factor genes. In cauliflower (B. oleracea var. botrytis), the semi-dominant Or gene mutant induces carotenoid (dominant beta-carotene) accumulation in the leaf bases and curd shoot meristems but not in the leaves (Li et al., 134). The relationship between carotenoid accumulation and the expression of biosynthesis genes was investigated in different tissues (Jung et al., 113). The BrPSY, BrPDS, BrZDS, BrLCYE, BrCHXB, and BrZEP genes were upregulated in the flowers and leaves in which lutein and beta-carotene were abundant. Additionally, Br300K microarray analysis has indicated that the downregulation of three carotene degradation genes, BrNCED3, BrNCED4, and BrNCED6, are associated with the carotenoid content in Chinese cabbage yellow leaves (Jung et al., 113).

Clotault et al. (35) investigated the expression of eight genes (phytoene synthase -PSY1 and PSY2, phytoene desaturase -PDS, ζ-carotene desaturase-ZDS1 and ZDS2, lycopene  $\varepsilon$ -cyclase-LCYE, lycopene β-cyclase -LCYB1, and zeaxanthin epoxidase-ZEP) encoding carotenoid biosynthesis enzymes during the development of white, yellow, orange, and red carrot roots. All eight genes were expressed in the white cultivar even though it did not contain carotenoids. The high expression of genes encoding LCYE and ZDS noted in yellow and red cultivars, respectively was found to be consistent with the accumulation of lutein and lycopene, respectively. Campos et al. (20) characterized and established role of plastid terminal oxidase gene DcPTOX during growth and development stages in different colour roots. Carotenoids are produced in both carrot (*Daucus carota*) leaves and reserve roots, and high amounts of  $\alpha$ -carotene and  $\beta$ -carotene accumulate in the latter. In some plant models, the presence of different isoforms of carotenogenic genes is associated with an organ-specific function. Carrot has also been reported to harbour two *Lcyb* genes, of which *DcLcyb1* is expressed in leaves and storage roots during carrot development, correlating with an increase in carotenoid levels. Moreno *et al.* (159) indicated that *DcLcyb1* does not possess an organ specific function and modulate carotenoid gene expression and accumulation in carrot leaves and storage roots.

Phytoene synthase catalyzes the first committed step in the carotenoid biosynthesis pathway, and its overexpression is the main driving force in the orange phenotype. Promoter analysis showed that *DcPSY* genes have diverged substantially in their regulatory sequences after gene duplication. Expression levels of DcPSY1 and DcPSY2 were generally positively correlated with carotenoid content during root development. Wang et al. (245) reported that DcPSY1 makes an important contribution to carotenoid accumulation in the leaves and is important for photosynthesis and photoprotection, but they are not the determining factors of root colour. Fraser et al. (69) analysed tomato fruit, at five stages of development, for their carotenoid and chlorophyll (Chl) contents, in vitro activities of phytoene synthase, phytoene desaturase, and lycopene cyclase, as well as expression of the phytoene synthase (Psy) and phytoene desaturase (Pds) genes. They reported highest carotenoid in ripe fruit while carotenogenic enzymic activities in green fruit. Of these enzymes, Psy was located in the plastid stroma whereas metabolism of phytoene was associated with plastid membranes during fruit development stages. Transcription of Psy and Pds is regulated developmentally, with expression being considerably elevated in chromoplast-containing tissues. The absence of detectable Psy and Pds *mRNA* in green tissues despite their high enzyme activities support the hypothesis of divergent genes encoding these enzymes. Enfissi et al. (59) performed expression analysis of 4, 4' carotenoid oxygenase (crtW) and 3, 3' hydroxylase (crtZ) from marine bacteria in tomato which showed low level production of ketocarotenoids in ripe fruit but over production of lycopene (~3.5 mg/g DW) along with delayed ripening. Su et al. (220) demonstrated that Carotenoid accumulation during tomato fruit ripening is modulated by the auxin-ethylene balance.

Anthocyanins are flavonoid pigments that are responsible for the red, purple and blue colours in

plants. These provide antioxidant and anti-cancer benefits to humans, as well as protection of plant DNA from UV light damage. Several vegetable crops including purple cauliflower, red cabbage, purple heading Chinese cabbage, purple capsicum and carrots are rich in anthocyanin in their stems, curds, heading leaves, fruits or roots (Chiu *et al.*, 29; Lorizzo *et al.*, 142; Sankhari *et al.*, 196; Singh *et al.*, 254; Stommel *et al.*, 218).

Genes involved in anthocyanin biosynthesis have been identified and characterized in A. thaliana and include 24 structure genes, 16 transcription factor genes, and one transport gene, as well as some miRNAs that participate in anthocyanin synthesis, regulation, and transport (Walker et al., 243; Matsui et al., 146; Gou et al., 80). 73 genes orthologous to these 41 A. thaliana anthocyanin biosynthesis genes have been identified in B. rapa, (Guo et al., 86). Differentially expressed genes that are involved in anthocyanin synthesis have been identified through comparative transcriptional analysis of green and purple lines/cultivars/mutants of Brassica vegetables. Almost all anthocyanin biosynthesis-related genes are upregulated in purple cultivars such compared with white ones (Zhang et al., 264). A very high upregulation of BoPAP1 gene, which encodes a R2R3 MYB transcription factor is believed to induce extensive anthocyanin accumulation in purple kale by activating structural genes under low temperature stress (Zhang et al., 262). BoMYB2, an ortholog of AtPAP2 or MYB113, together with BobHLHs has been associated with anthocyanin synthesis in purple cauliflower (B. oleracea var. botrytis) mutant (Chiu and Li, 28), however, a different pattern of expression of these genes was reported in Sicilian purple by Singh et al. (Singh et al., 209). The BrMYB2 and BrTT8 are two regulatory factors of anthocyanin accumulation in purple heading Chinese cabbage line rich in anthocyanin (He et al., 93; Zhang et al., 264).

A wide array of tissue-specific anthocyanin pigmentation is observed in *Capsicum annuum* L. (pepper). In order to determine the genetic basis for tissue-specific pigmentation, Stommel *et al.* (262) studied the expression of anthocyanin biosynthetic (*Chs, Dfr,* and *Ans*) and regulatory (*Myc, Myb<sub>A</sub>*, and *Wd*) genes in flower, fruit, and foliar tissue from pigmented and nonpigmented *C. annuum* genotypes. Biosynthetic gene transcript levels were significantly higher in anthocyanin-pigmented tissue than in nonpigmented tissues. *Myb<sub>A</sub>* and *Myc* transcript levels were also substantially higher in anthocyaninpigmented floral and fruit tissues. They demonstrated that differential expression of *C. annuum Myb<sub>A</sub>* as well as Myc occurs was coincident with anthocyanin accumulation in *C. annuum* flower and fruit tissues. In contrast to the situation in flowers and fruit, differential expression of  $Myb_A$  and Myc was not observed in foliar tissue, suggesting that different mechanisms contribute to the regulation of anthocyanin biosynthesis in different parts of the *C. annuum* plant.

Purple carrots can accumulate large quantities of anthocyanins in their roots. Lorizzo *et al.* (180) performed fine mapping combined with gene expression analyses to identify candidate genes controlling anthocyanin pigmentation in the carrot root and petiole. They identified a cluster of six *MYB* transcription factors, denominated *DcMYB6* to *DcMYB11*, associated with the regulation of anthocyanin biosynthesis. No anthocyanin biosynthetic genes were present in this region. Comparative transcriptome analysis indicated that upregulation of *DcMYB7* was always associated with anthocyanin pigmentation in both root and petiole tissues, whereas *DcMYB11* was only upregulated with pigmentation in petioles.

In addition to the pigments and secondary metabolites biosynthetic pathways of vitamins have also been studied in vegetable crops. The expression profiles of vitamin C synthetic pathway-related genes have been investigated in various studies. In non-heading Chinese cabbage, the upregulation of three *d*-mannose//-galactose pathway genes (*PMI*, *GME*, and *GGP*) and downregulation of ascorbate oxidase (*AAO*) was correlated with the high vitamin C contents (Ren *et al.*, 187). Additionally, the overexpression of a monodehydroascorbate reductase gene (*MDHA*) in non-heading Chinese cabbage and tobacco reduces its ascorbate level, thereby indicating its negative regulatory function (Ren *et al.*, 188).

# Pre-breeding and introgressiomics for nutraceutical enrichment

The pre-breeding is transferring useful gene(s) from exotics/wild (unadapted sources) types into agronomically acceptable background/breeding materials. It aims to generate conventionally usable new base population/genetic stock which is/are expected to have merit to be included in ordinary breeding programme. It is a bridge between gene pool and crop improvement. There are different approaches to pre-breeding, namely Introgression, Incorporation, wide crossing, somatic hybridization, genetic transformation and others. The germplasm of vegetable crops useful for biofortification/quality traits is given in Table 8. Introgressiomics is a related term to pre-breeding and Prohens *et al.* (182) defined it as a mass scale systematic development of plant

genome fragments obtained from (mostly wild) crop relatives into the genetic background of crops that may allow developing new generations of cultivars with improved properties'. They described five clear steps: (i) Identification of CWRs to be used in the programme; (ii) Hybridization and backcrossing of the crop with a number (as large as possible) of crop wild relatives from different genepools using special techniques when needed; (iii) Development of multiple special introgression populations containing introgressed fragments from one or several CWRs using genomic tools; (iv) Creating repositories of the introgressiomics populations and materials and databases with phenotypic and genomic information; and lastly, and (v) Moving the materials into breeding pipelines. Traka et al. (232) developed three highglucoraphanin F, broccoli hybrids through genome introgression from the wild species Brassica villosa. The high-glucoraphanin broccoli hybrids contained 2.5-3 times the glucoraphanin content of standard hybrids due to enhanced sulphate assimilation and modifications in sulphur partitioning between sulphur-containing metabolites. All of the highglucoraphanin hybrids possessed an introgressed B.villosa segment which contained B.villosa Myb28 allele. Xishuangbanna gourd (C. sativus L. var. xishuangbannanesis) is a potential donor source of beta-carotene (700 µg/100 g fresh weight) for use in enrichment of commercial cucumber (Cucumis sativus L) which contains beta-carotene in range of 22-48 µg/100 g fresh weight. Endocarp quantity of β-carotene (QbC) in cucumber was governed by a single recessive gene ore on chromosome 3 (Cuevas et al., 40). But, this need further investigation for some commercially undesirable traits (e.g., poor fruit quality including fruit shape, length, and spine colour). Orange colour intensity (hue; QbC) and dispersion (uniform colour) in endocarp/mesocarp tissue are likely under polygenic control.

materials and populations carrying introgressions of

Glucosinolates are functional compounds which affect health due the activities by degradation products such as sulphoraphane which acts against tumor inhibition through different mechanisms such as induction of phase 2 response, anti-inflammatory activity, antibiotic against *H. pylori*, cell cycle arrest and apoptosis and variable effects on phase I. Hence, development of broccoli rich in health beneficial glucoiberin (3- MSP) and glucoraphanin (4-MSB) was attained by Faulkner *et al.* (64) and Mithen *et al.* (154) using *Brassica villosa*. They identified two QTLs having major effect on glucosinolate content alongwith linked microsatellite marker O112-F02 for QTL-2. Glucosinolates content was high when QTL1 was from *B. villosa*, although it was highest (10  $\mu$ mol/g dry weight) when both QTL1 and QTL2 were from *B. villosa* followed by QTL1 from *B. villosa* and QTL2 in heterozygous state. Interestingly, MSB content and sulforaphane content of soup after 90 s cooking was increased 89% in 48/13 × BR9 (*B. villosa* based hybrid).

Wild species are used to enhance bioavailability of antioxidants in vegetables such as Willits et al. (254) enhanced guercetin in fruit flesh and peel in tomato using S. pennellii and anthocyanin from S. chilense (dominant allele Anthocyanin fruit, AFT) and S. cheesmanie (recessive allele Atroviolacium, atv) and S. lycopersicoides (dominant allele Aubergine, ABG). Fruits of either AFT-/atvatv and ABG-/atvatv hybrids showed high production of anthocyanins in the peel (Mes et al., 152; Gonzali et al., 79). Similarly, flavonoids content increased in Allium sativum using A. ursinum and A. victorialis (Wu et al., 257). However, classical breeding has limitation therefore molecular markers are ideal to handle such complex to measure traits and provide quick tool that allows breeders to accelerate and reduce the costs of selection programmes. In onion, Kim et al. (117) developed a codominant PCR-based marker linked to the DFR-A gene, known to be involved in the last steps of anthocyanin biosynthesis. They proposed the use of this marker to expedite the screening of

heterozygous red onions in segregating populations, thereby eliminating the need for time-consuming progeny tests. Further, specific biochemical pathways can be modified through the activation or repression of specific genes.

**Omics approaches in biofortification:** Omics represents collective technologies used to understand the roles, relationships, and actions of the various types of molecules that make up the cells of an organism. This leads to the development of new breeding tools including precise cross breeding and genetic engineering. The use of omics tools is in infancy for nutraceutical research in vegetable crop. Xiaonan *et al.* (259) reviewed the use of omics technologies to analyze the genome sequence, transcripts, proteins, and metabolites involved in phytonutrient biosynthesis/ degradation for nutritional enrichment.

# Minor vegetable crops as source of dietary nutrients and nutraceuticals

Minor vegetables are important source of food and dietary needs of tribal and vulnerable communities in remote forest and rural areas (Vijay Bhaskar *et al.*, 240; Singh *et al.* 211). Although, no attempt have been made except few of these underexplored crops for yield or quality traits they are inherently found to be rich in various minerals (Ca, Fe, Zn) and dietary microelements such as vitamins and amino acids

Crops	Wild relatives/ accessions/ landraces/ varieties	Nutrients
Tomato	S. pimpinellifolium, Caro Red (Rugers x S. hirsutum)	Vitamin A
	Caro Rich, F-7045, VRT-35, CGT, VRT-5	Beta carotene
	High pigment mutants (hp), Crimpson (og), Pusa Rohini	Lycopene
	S. pennellii IL6-2, IL7-2	Phenolics
	S. pennellii IL12-4	Ascorbic acid
	S. chilense and atroviolacium (atv) from S. cheesmaniae	Anthocyanin
Chilli	C. annuum var. IC: 119262(CA2), Bayadaggi kaddi	Ascorbic acid
Paprika	KTPL-19	Capsanthin
Cucumber	Xishuangbanna gourd (C. sativus var. Xishuangbananesis)	Beta carotene
Muskmelon	Honey dew32	Ascorbic acid
	Canary yellow	Flavons (Naringenin chalcone)
Spine gourd	Momordica dioca	Protein
	M. chochinchinenesis	Lycopene
Bitter gourd	DRAR-1, DVBTH-5	Beta carotene
	DRAR-1, DVBTG-5	Ascorbic acid
Sweet potato	Resisto, Zambezi, Chiwoko	Beta carotene
Cassava	UMUCASS 44, UMUCASS 45 and UMUCASS 46	Vitamin A
Broccoli	Brassica villosa	Glucosinolates

Table 8. Crop wild relatives rich in quality traits useful for breeding.

(Singh et al., 210). They are hardy for biotic and abiotc stresses, require less care and low inputs, easily accessible in natural habitats or home gardens and are a rich source of micronutrients (Flyman and Afolayan, 67). Despite their resilience to climate change factors and first line of foods for primitive and indigenous tribes still they are neglected vegetables. Certain antinutrients like nitrate, phytate and oxalate content in these vegetables is a matter of concern, however, they can be removed by common cooking practices (Aletor and Adeugun, 4; Singh et al., 213) but still the breeding genotypes hacing low values of antinutrients and high content of nutritional content could be preferred option. Underutilized vegetables also have potential for nutraceuticals such as gac (Momordica cochinchinensis) contain very high lycopene 380 ( $\mu$ g g<sup>-1</sup> fresh fruit) than commonly known sources like tomato (31  $\mu$ g g<sup>-1</sup>), watermelon (41  $\mu$ g g<sup>-1</sup>) (Aoki *et al.* 6). Similarly, aril fraction of teasel gourd (an underutilized cucurbitaceous vegetable) also contain high amount of lycopene and have potential for commercial extraction (Singh et al. 212) from arils. Hence, they could make an important contribution to combating micronutrient malnutrition as well as providing food security.

#### Antinutritional factors and their elimination

Some of the vegetables contain chemical compounds which are: (i) fatal/toxic to human,

and (ii) interfering agents to dietary nutrients and enzymes and called as 'anti-nutritional factors'. Some of them are oxalates, nitrate, phytate, saponin, tannins, glycosides which affect human health. It is important for consumers and researchers to understand the importance of these chemicals and their impacts on human health and available methods for their assessments. Anti-nutritional compounds in vegetables are given in Table 9. In faba bean, condensed vicine, convicine and condensed tannins are major antinutrients and two recessive genes, *zt-1* and *zt-2* interrupt the anthocyanin biosynthetic pathway and cause zero-tannin content and promote white-flowered plants. The removal through marker assisted breeding was attempted by Gutierrez et al. (90) by developing SCAR markers for zt-2 and *VfTTG1* gene for a loss of pigmentation in flowers and absence of tannins. Vicine-convicine content may decrease up to 20 times due to presence of the vcallele that is linked to colourless hilum in the seeds. Another gene vcr is also reported to reuduce 20-fold reduction in vicine-convicine concentration with respect to wild-type faba bean similar to vc<sup>-</sup> (Ramsay et al., 184). Phytate is another antinutrient in plant but, low phytate affects germination, emergence, stress tolerance and yield. Hence, biotechnological approaches have been suggested to remedy this problem, including embryo-specific silencing of an ABC transporter responsible for phytic acid

Table 9. Antinutrients in plant foods that reduce nutrient bioavailability impairing health.

Antinutrients	Effect	Dietary source
Phytic acid	Binds minerals K, Mg, Ca, Fe, Zn	Legumes and cereals
Trypsin inhibitor	Reduces the activity of the enzyme trypsin and other closely related enzymes that help digest protein	Legumes, cereals and potato
Hemaglutinin, eg. Lectin	Interfere with cells lining the gastrointestinal tract causing acute symptoms, can bind metals and some vitamins	Legumes
Polyphenolics, tannins	Form complexes with iron, zinc, copper that reduces mineral absorption	Beans, tea, coffee, sorghum
Cyanogens or glycoalkaloids	Inhibit acetylcholinestrase activity which impair nerve transmission, can damage cell membranes	Cassava, peas, beans
Oxalic acid	Binds calcium to prevent its absorption	Spinach leaf, amaranth, rhubarb, portulaca, colocasia, elephant foot yam
Solanine	Can be toxic, affect gastro intestinal and nervous system	Green parts of potato tubers
Saponins	May irritate the gastrointestinal tract and interfere with nutrient absorption	Soybeans, peas, sugar beets, pea nuts
Goitrogens	Suppress thyroids function	Brassica, alliums foods
Cadmium, mercury, lead	May have toxic effects, e.g High levels of Hg impair brain development	Contaminated leafy vegetables
Glycosides	Liberate toxic hydrocynic acid with enzymic action	Tapioca leaves
Dioscorine	Toxic alkaloid	Yam

accumulation (Shi *et al.*, 208) or by Silencing of a key phytate transporter and engineering of increased phytase activity in seeds (Campion *et al.*, 19). Tripsin inhibitor content in pea seeds is governed by QTL closely linked to *TI* genes at the *Tri* locus (Domoney *et al.*, 54). Zivanov *et al.* (271) used *Pisum elatius* to reduce low trypsin inhibitor and found a marker At13/ At5 for use in breeding.

#### **Bioavailability studies**

Horvitz et al. (95) investigated higher level of serum lycopene and beta-carotene concentrations in human beings while fed with red carrot in comparison to white carrot, tomato paste and white carrot+tomato paste. Morris et al. (160) reported that fractional absorption of calcium from control carrots was 48.8% for females and 56.9% for males, compared with 42.1% for females and 43.8% for males for the sCAX1expressing carrots. However, fractional Ca absorption is lower in the sCAX1-expressing carrots, the total Ca absorbed per 100 g of fresh carrots is 45.9% higher for females and 38.7% higher for males from the sCAX1-expressing carrots. Therefore, the sCAX1expressing carrots contain more bioavailable Ca in both the mouse and human models. Bioavailability studies on beta-carotene biofortified cassava in Kenyan children showed increased level of serum retinol concentration over the control and serum betacarotene concentration over both control and betacarotene supplement. Further, prevalence of vitamin A deficiency was reduced to 29% with yellow cassava while it was still at higher levels in control (34%) and beta-carotene supplement (33%) (Talsma et al., 223). The mineral absorption increases from food plant alongwith increase in 'mineral promoters' such as ascorbic acid and vitamin A and also by reducing antinutritional compounds namely oxalate, nitrate, phytate, saponin, tannins and tripsin inhibotors in vegetables such as leafy vegetables, legume and tuber crops (Platel and Srinivasan, 180).

# CONCLUSION

Promoting a nutrient rich crop fitting to growing conditions in target communities is relatively less time demanding compared to developing varieties and challenging the deficiencies. Fortification of nonperishable public food is option but its affordability and mass availability to masses at reasonable prices (on consumer side) will remain a challenge. Except 'iodised salt' we hardly found a big breakthrough showing larger implication on public health. Biofortification using classical breeding or new breeding approaches always face challenges from traditional food culture point of view, so participatory mode is always a preferred mode.

Biofortified crops have potential to increase delivery of minerals to vulnerable communities pro rata to their contribution to the diet, without a change in behaviour. However, before planning biofortification programme, it is essential to understand the issues related to, such as (i) feasibility of breeding nutrient/ nutraceutical/pharmaceutical/bioactive compounds dense varieties, (ii) chances of adoption of new genotypes by farmers, (iii) target nutrient content for breeding, (iv) impact on nutritional status, (v) costeffectiveness of new genotypes, and (vi) consumers acceptance of biofortified foods (Nestel et al., 2006). Breeding vegetable for guality traits needs adequate attention to meet the diverse need of consumers' taste, health and nutrition. Identify vegetables for target nutraceuticals, other industrial compounds and develop holistic programme by PPP mode with breeder and health expert's participation. Adequate focus should be given to explore the nutritional and health potential of underutilized vegetables particularly in the areas/communities of their preference. Exploit wild species and land races for genes/QTLs for nutraceuticals and other bioactive compounds. Molecular tools have great potential to support conventional breeding in developing vegetable varieties high in nutraceuticals/ pharmaceuticals/ bioactive compounds. Use of 'omics' approaches to understand complexities of biosynthetic pathway is also emerging option to use for tailoring the genotypes for plan vaccine, nutraceutical and aromatic compounds.

#### **REFERENCES:**

- Adato, A., Mandel, T., Mintz-Oron, S. *et al.* 2009. Fruit-surface flavonoid accumulation in tomato is controlled by a *SIMYB12*-regulated transcriptional network. *PLoS Genet.* 5: e1000777.
- Agrawal, A.A. and Kurashige, N.S. 2003. A role for isothiocyanates in plant resistance against the specialist herbivore *Pieris rapae*. *J. Chem. Ecol.* 29: 1403-15.
- Al Said, F.A., Al Farsi, K., Khan, I.A., Ali, A., Khan, M.M. and Iqbal, Q., 2014. Evaluation of adaptability and nutritional quality of 54 tomato accessions grown in Oman. *J. Food, Agric. Environ.* 12: 40-50.
- Aletor, V. A. and Adeogun, O. A. 1995. Nutrient and antinutrients components of some tropical leafy vegetables. *Food Chem.* 53: 375–79.
- Andersson, M., Turesson, H., Nicolia, A., Fält, A. S., Samuelsson, M. and Hofvander, P. 2017.

Efficient targeted multiallelic mutagenesis in tetraploid potato (*Solanum tuberosum*) by transient *CRISPR-Cas9* expression in protoplasts. *Plant Cell Rep.* **36**: 117–28.

- Aoki, H., Kieu, N.T.M., Kuze, N., Tomisaka, K. and Chuyen, N.V. 2002. Carotenoid pigments in GAC fruit (*Momordica cochinchinensis* SPRENG). *Biosci. Biotechnol. Biochem.* 66: 2479-82.
- Bang, H., Davis, A., Kim, S., Leskovar, D. and King, S. 2010. Flesh color inheritance and gene interactions among canary yellow, pale yellow, and red watermelon. *J. American Soc. Hort. Sci.* 135: 362-68.
- Bang, H., Kim, S., Leskovar, D. and King, S. 2007. Development of a codominant CAPS marker for allelic selection between canary yellow and red watermelon based on SNP in lycopene β-cyclase (LCYB) gene. *Mol. Breed.* 20: 63–70.
- Ben Chaim, A., Paran, I., Grube, R. *et al.* 2001. QTL mapping of fruit related traits in pepper (*Capsicum annuum*). *Theor. Appl. Genet.* **102**: 1016–28.
- Blum E, Liu K, Mazourek M, Yoo EY, Jahn M. and Paran I. 2002. Molecular mapping of the *C* locus for presence of pungency in Capsicum. *Genome*, 45: 702–05.
- Borovsky Y, Oren-Shamir M, Ovadia R, De Jong W. and Paran I. 2004. The A locus that controls anthocyanin accumulation in pepper encodes a MYB transcription factor homologous to Anthocyanin 2 of Petunia. *Theor. Appl. Genet.* **109**: 23–29.
- 12. Borovsky, Y.and Paran, I. 2008. Chlorophyll breakdown during pepper fruit ripening in the chlorophyll retainer mutation is impaired at the homolog of the senescence-inducible stay-green gene. *Theor. Appl. Genet.* **117**: 235–40.
- Bouis, H.E. and Saltzman, A. 2017. Improving nutrition through biofortification: a review of evidence from HarvestPlus, 2003 through 2016. *Global Food Sec.* 12: 49–58.
- Brar, J.S., Gill, H.S. and Nandpuri, K.S. 1969. Inheritance of qualitative characters in turnips (*Brassica rapa*). *J. Res. PAU*, 6: 907.
- 15. Brown, A. F., Yousef, G. G., Jeffery, E. H., Klein, B. P., Wallig, M. A., Kushad, M. M. *et al.* 2002.

Glucosinolate profiles in broccoli: Variation in levels and implications in breeding for cancer chemoprotection. *J. American Soc. Hort. Sci.* **127**: 807–13.

- Buishand, J. G. and Gabelman, W.H. 1979. Investigations on the inheritance of color and carotenoid content in phloem and xylem of carrot roots (*Daucus carota* L.). *Euphytica*, **28**: 61I-32.
- Buishand, J.G. and Gabelman, W.H. 1980. Studies on the inheritance of root color and carotenoid content in red × yellow and red × white crosses of carrot (*Daucus carota* L.). *Euphytica*, 29: 241-60.
- Butler, N.M., Baltes, N.J., Voytas, D.F., and Douches, D.S. 2016. Gemini virus mediated genome editing in potato (*Solanum tuberosum* L.) using sequence specific nucleases. *Front. Plant Sci.* 7: 1045. doi: 10.3389/fpls.2016.01045
- Campion, B., Sparvoli, F., Doria, E., Tagliabue, G., Galasso, I., Fileppi, M., Bollini, R. and Nielsen E. 2009. Isolation and characterisation of an Ipa (low phytic acid) mutant in common bean (*Phaseolus vulgaris* L.) *Theor. Appl. Genet.* **118**: 1211–21.
- Campos, M.D., Campos, C., Cardoso, H.G., Simon, P.W., Oliveira, M., Nogales, A. and Arnholdt-Schmitt, B., 2016. Isolation and characterization of plastid terminal oxidase gene from carrot and its relation to carotenoid accumulation. *Plant Gene*, 5: 13-21.
- Cartea, M. E., Velasco, P., Obregón, S., Padilla, G. and De Haro, A. 2008. Seasonal variation in glucosinolate content in *Brassica oleracea* crops grown in northwestern Spain. *Phytochem*. 69: 403–10.
- Cartea, M.E., de Haro, A., Obregon, S., Soengas, P. and Velasco, P., 2012. Glucosinolate variation in leaves of *Brassica rapa* crops. *Plant Foods Hum. Nutr.* 67: 283–88.
- 23. Castaneda-Ovando, A., Ma de Lourdes, P. H., Ma Elena, P.H., Rodriguez, J.A. and Carlos, A.G.V. 2009. Chemical studies of anthocyanins: A review. *Food Chem.* **113**: 859-71.
- Cermak, T., Baltes, N.J., Cegan, R., Zhang, Y., and Voytas, D.F. 2015. High frequency, precise modification of the tomato genome. *Genome Biol.* 16: 232. doi: 10.1186/s13059-015-0796-9

- Chan, L. F., Chen L.F., Lu, H.Y., Lin, C.H., Huang, H. C., Ting, M.Y., *et al.* 2009. Growth, yield and shelf-life of isopentenyltransferase (*ipt*)-gene transformed broccoli. *Canadian J. Plant Sci.* 89: 701-11.
- Chandra, A.K., Mukhopadhyay, S., Lahari, D. and Tripathy, S. 2004. Goitrogenic content of Indian cyanogenic plant foods and their *in vitro* anti-thyroidal activity. *Indian J. Med. Res.* **119**: 180-85.
- Chen, L., Yang, D., Zhang, Y., Wu, L., Zhang, Y., Ye, L., Pan, C., He, Y., Huang, L., Ruan, Y.L. and Lu, G. 2018. Evidence for a specific and critical role of mitogen-activated protein kinase 20 in unito-binucleate transition of microgametogenesis in tomato. *New Phytol.* **219**: 176–94.
- 28. Chiu, L.W. and Li, L. 2012. Characterization of the regulatory network of *BoMYB2* in controlling anthocyanin biosynthesis in purple cauliflower. *Planta*, **236**: 1153–64.
- 29. Chiu, L.W., Zhou, X., Burke, S., Wu, X., Prior, R.L. and Li, L. 2010. The purple cauliflower arises from activation of a *MYB* transcription factor. *Plant Physiol*, **154**: 1470–80.
- 30. Choudhury, H. L. 1972. Genetical studies in some West African Solarium melongena, Canadian J. Genet. Cytol. 14: 441.
- Cichy, K.A., Forster, S., Grafton, K.F. and Hosfield, G.L., 2005. Inheritance of seed zinc accumulation in navy bean. *Crop Science*, 45: 864-70.
- Clarke, A. E., Jones, H. A. and Little, T. M. 1944. Inheritance of bulb color in onion. *Genetics*, 29: 569.
- Clayberg, C. 1992. Interaction and linkage test of flesh color genes in *Cucumis melo* L. *Rep. Cucurbit Genet. Coop.* 15: 53.
- Clayberg, C.D. 1972. Preliminarymapping of three chromosome 7 genes. *Tomato Genet Coop. Rep.* 22: 4.
- Clotault, J., Peltier, D., Berruyer, R., Thomas, M., Briard, M., Geoffriau, E., 2008. Expression of carotenoid biosynthesis genes during carrot root development. *J. Exp. Bot.* **59**: 3563–73.
- 36. Colbert, T.G., Hurst, S.R. and Slade, A.J. 2011. Tomatoes that soften more slowly postharvest

due to non-transgenic alterations in an expansin gene. US Patent Application 20110113507.

- Colonna, V., D'Agostino, N., Garrison, E., Albrechtsen, A., Meisner, J., Facchiano, A., *et al.* 2019.Genomic diversity and novel genome-wide association with fruit morphology in *Capsicum*, from 746k polymorphic sites. *Sci Rep.* **11**: 9:10067. doi: 10.1038/s41598-019-46136-5.
- Cong, B., and Tanksley, S.D. 2006. FW2.2 and cell cycle control in developing tomato fruit: a possible example of gene cooption in the evolution of a novel organ. Plant Mol. Biol. 62: 867–80.
- Cong, B., Barrero L.S.and Tanksley, S.D. 2008. Regulatory change in YABBY-like transcription factor led to evolution of extreme fruit size during tomato domestication. *Nature Genet.* 40: 800–04.
- 40. Cuevas, H.E., Song, H., Staub, J.E. and Simon, P.W., 2010. Inheritance of beta-caroteneassociated flesh color in cucumber (*Cucumis sativus* L.) fruit. *Euphytica*, **171**: 301.
- Dahmani-Mardas, F., Troadec, C., Boualem, A., Lévêque, S., Alsadon, A.A., Aldoss, A.A *et al.* 2010. Engineering melon plants with improved fruit shelf life using the TILLING Approach. *PLoS One*, 5: e15776.
- Das, L., Bhaumik, E., Raychaudhuri, U. and Chakraborty, R. 2012. Role of nutraceuticals in human health. *J. Food Sci. Tech.* 49: 173-83.
- 43. Davey, V. M. 1931. Colour inheritance in swedes and turnips and its bearing on the identification of commercial stocks. *Scottish. J. Agric.* **14**: 303.
- 44. Davis, A.R., King, S.R., Jarret, R.L., Levi, A. and Tadmor, Y.A., 2004. Lycopene and Total Carotenoid Content as Descriptors for Citrullus lanatus: Limitations and Preliminary Trials. *Rep. Cucurbit Genet. Coop.* **27**: 34.
- 45. De Jong, H. 1991. Inheritance of anthocyanin pigmentation in the cultivated potato a critical-review. *American Potato J.* **68**: 585–93.
- De Jong, M., Wolters-Arts, M., Feron, R., Mariani, C. and Vriezen, W. H. 2009. The Solanum lycopersicum auxin response factor 7 (SIARF7) regulates auxin signaling during tomato fruit set and development. *Plant J.* **5**: 160–70.

- 47. De Jong, W.S., Eannetta, N.T, De Jong, D.M. and Bodis, M. 2004. Candidate gene analysis of anthocyanin pigmentation loci in the Solanaceae. *Theor. Appl. Genet.* **108**: 423–32.
- De Koeyer, D., Douglass, K., Whitney, A.S., Nolan, L., Song, Y. and De Jong W. 2010. Application of high-resolution DNA melting for genotyping and variant scanning of diploid and autotetraploid potato. *Mol. Breed.* 25: 67–90.
- Dempsey, A.H. 1962. Inheritance studies of certain fruit and plant characters in *Capsicum frutescens*. *Abstracts in Plant Breeding Abstracts*, 31: Entry No 785.
- Deng, L., Wang, H., Sun, C., Li, Q., Jiang, H., Du, M., Li, C.B. and Li, C. 2018. Efficient generation of pink-fruited tomatoes using CRISPR/Cas9 system. *J. Genet. Genom.* 45: 51–54.
- Dias, J.S. and Ortiz, R. 2012. Transgenic Vegetable Crops: Progress, Potentials and Prospects. *Plant Breed. Rev.* 35: 151-46.
- 52. Dias, J.S. and Ortiz, R. 2014. Advances in Transgenic Vegetable and Fruit Breeding. *Agric. Sci.* **5**: 1448-67.
- 53. Diaz de la Garza, R.I., Gregory, J.F. and Hanson A.D. 2007. Folate biofortification of tomato fruit," *Proc. Natl. Acad. Sci. USA*, **104**: 4218- 22.
- Domoney, C., Welham, T., Sidebottom, C. and Firmin, J.L. 1995. Multiple isoforms of *Pisum* trypsin inhibitors result from modification of two primary gene products. *FEBS Lett.* 360:15–20
- 55. Dunlop, J. and Phung, T. 2002. Transporter genes to enhance nutrient uptake: Opportunities and challenges. *Plant Soil*, **245**: 15-122.
- Eady, C.C., Kamoi, T., Kato, M., Porter, N.G., Davis, S., Shaw, M., *et al.*. 2008. Silencing onion lachrymatory factor synthase causes a significant change in the sulfur secondary metabolite profile. *Plant Physiol.* **147**: 2096-2106.
- 57. El-Shafie, M. and Davis, G. 1967. Inheritance of bulb colour in *Allium cepa. Hilgardia*, **9**: 607.
- Elshire, R.J., Glaubitz, J.C., Sun, Q.; Poland, J. A., Kawamoto, K., Buckler, E. S., Mitchell, S. E. 2011. A robust, simple genotyping-by-sequencing (GBS) approach for high diversity species. *PLoS One*, 6: e19379.

- Enfissi, E., Nogueira, M., D'Ambrosio, C., Stigliani, A. L., Giorio, G., Misawa, N., and Fraser, P. D. 2019. The road to astaxanthin production in tomato fruit reveals plastid and metabolic adaptation resulting in an unintended high lycopene genotype with delayed over-ripening properties. *Plant Biotechnol. J.* **17**: 1501–13.
- 60. FAO, 2002. Joint FAO/WHO Working Group Report on Drafting Guidelines for the Evaluation of Probiotics in Food London, Ontario, Canada, April 30 and May 1, 2002.
- 61. FAO/WHO, 2003. FAO/WHO launch expert report on diet, nutrition and prevention of chronic diseases. FAO, Rome, *23 April 2003.*
- 62. FAOSTAT, 2018. Food and Agriculture Statistics. Food and Agriculture Organization, Rome. Accessed on 24 April, 2020.
- Farnham, M.W., Wilson, P.E., Stephenson, K.K. and Fahey, J. W. 2004. Genetic and environmental effects on glucosinolate content and chemoprotective potency of broccoli. *Plant Breed.*123: 60–65.
- 64. Faulkner, K., Mithen, R. and Williamson, G. 1998. Selective increase of the potential anticarcinogen 4-methylsulphinylbutyl glucosinolate in broccoli. *Carcinogenesis*, **19**: 605-09.
- Feng, J., Long, Y., Shi, L., Shi, J., Barker, G. and Meng, J. 2012. Characterization of metabolite quantitative trait loci and metabolic networks that control glucosinolate concentration in the seeds and leaves of *Brassica napus*. *New Phytol.* **193**: 96–108.
- Filler H. S., Melamed Bessudo, C. and Levy, A.A. 2017. Targeted recombination between homologous chromosomes for precise breeding in tomato. *Nature Commun.* 8: 15605
- Flyman, M.V. and Afolayan, A.J. 2006. The suitability of wild vegetables for alleviating human dietary deficiencies. *South African J. Botany*, **72**: 492-97.
- Frary, A., Doganlar, S., Daunay, M.C. and Tanksley, S.D. 2003. QTL analysis of morphological traits in eggplant and implications for conservation of gene function during evolution of solanaceous species. *Theor. Appl. Genet.* **107**: 359–70.
- 69. Fraser, Mark, R., Truesdale, Colin, R., Bird, Wolfgang Schuch, and Peter M. Bramley. 1994.

Carotenoid Biosynthesis during Tomato Fruit Development' Evidence for Tissue-Specific Gene Expression Paul D. *Plant Physiol.* **105**: 405-13.

- Fray, R. G. and, Grierson, D. 1993. Moleculargenetics of tomato fruit ripening. *Trends Genet*. 9: 438-43.
- 71. Fridman, E., Pleban, T. and Zamir, D. 2000. A recombination hotspot delimits a wild-species quantitative trait locus for tomato sugar content to 484 bp within an invertase gene. *Proceed. Nat. Acad. Sci.USA*, **97**: 4718–23.
- 72. Frimmel, F. 1938. Karottenzuchtung. *Zuchter*, **10**: 181–85.
- Gady, A.L, Hermans, F.W., Van de Wal, M.H., van Loo, E.N., Visser, R.G. and Bachem, C.W. 2009. Implementation of two high through-put techniques in a novel application: detecting point mutations in large EMS mutated plant populations. *Plant Methods*, **5**: 13.
- 74. Gady, A.L.F, Vriezen, W.H., Van de Wal M.H.B.J., Huang, P., Bovy, A.G., Visser, R.G.F. and Bachem, C.W.B. 2011. Induced point mutations in the phytoene synthase 1 gene cause differences in carotenoid content during tomato fruit ripening. *Mol. Breed.* DOI 10.1007/s11032-011-9591-9.
- 75. Gao, J., Yu, X., Ma, F. and Li, J. 2014. RNAseq analysis of transcriptome and glucosinolate metabolism in seeds and sprouts of broccoli (*Brassica oleracea* var. *italica*). *PLoS One*, **9**: e88804.
- 76. Giorgiev, C. 1972 Anthocyanin fruit tomato. *Tomato Genet. Coop. Rep.* **22**: 10.
- 77. Golubkina, N., Zamana, S., Seredin, T., Poluyarinov, P. *et al.* 2019. Effect of selenium biofortification and beneficial microorganism inoculation on yield, quality and antioxidant properties of shallot bulbs. Plants (Basel), 8: 102. doi: 10.3390/plants8040102.
- Gonzalez, M., M. Xu, C. Esteras, C. Roig, A.J. Monforte, C. Troadec, M. Pujol, F. Nuez, A. Bendahmane, J. Garcia-Mas, and B. Pico. 2011. Towards a TILLING platform for functional genomics in Piel de Sapo melons. *BMC Res Notes* 4: 289.
- 79. Gonzali, S., Mazzucato, A. and Perata, P. 2009. Purple as a tomato: towards high anthocyanin tomatoes. Trends Plant Sci. 14: 237–41.

- 80. Gou, J.Y., Felippes, F.F., Liu, C.J., Weigel, D. and Wang, J.W. 2011. Negative regulation of anthocyanin biosynthesis in *Arabidopsis* by a miR156- targeted SPL transcription factor. *Plant Cell*, **23**: 1512–22.
- Bragnani M. 2013. The EU regulation 1151/2012 on quality schemes for agricultural products and foodstuffs. *European Food Feed Law Rev.* 8: 376-85.
- Grandillo S., Tanksley S. D. 1996 Analysis of horticultural traits differentiating the cultivated tomato from the closely related species *Lycopersicon pimpinellifolium*. Theor. Appl. Genet. **92**: 935–951.
- Grandillo, S., Ku, H.M. and Tanksley, S.D. 1999. Identifying the loci responsible for natural variation in fruit size and shape in tomato. *Theor. Appl. Genet.* **99:** 978–87.
- 84. Grennan, A.K. 2009. Identification of Genes Involved in Metal Transport in Plants. *Plant Physiol.* **149**: 1623–24.
- 85. Guo, N., Cheng, F., Wu, J., Liu, B., Zheng, S., Liang, J. and Wang, X. 2014. Anthocyanin biosynthetic genes in *Brassica rapa*. *BMC Genom*. **15**: 426.
- Guo, R., Huang, Z., Deng, Y., Chen, X., Xuhan, X. and Lai, Z., 2016. Comparative transcriptome analysis reveal a special glucosinolate metabolism mechanism in *Brassica alboglabra* sprouts. *Front. Plant Sci.* 7: 1497.
- 87. Guo, S., Zhao, S., Sun, H., Wang, X., Wu, S., Lin, T., Ren, Y., Gao, L., Deng, Y., Zhang, J. and Lu, X., 2019. Resequencing of 414 cultivated and wild watermelon accessions identifies selection for fruit quality traits. *Nature genetics*, pp.1-8.
- Guo, S., Zhang, J., Sun, H., Salse, J., Lucas, W.J., Zhang, H. *et al.* 2013. The draft genome of watermelon (*Citrullus lanatus*) and resequencing of 20 diverse accessions. *Nature Genet.* 45: 51-58.
- Gupta, A., Pal, R. K. and Rajama, M. V. 2013. Delayed ripening and improved fruit processing quality in tomato by RNAi-mediated silencing of three homologs of 1-aminopropane-1carboxylate synthase gene. *J. Plant Physiol.* **170**: 987–95.
- 90. Gutierrez, N., Avila, C.M., Moreno, M.T. and Torres, A.M. 2008. Development of SCAR

markers linked to zt-2, one of the genes controlling absence of tannins in faba bean. *Aust. J. Agric. Res.* **59**: 62–68.

- 91. Habauzit, V. and Morand, C. 2012. Evidence for a protective effect of polyphenols-containing foods on cardiovascular health: an update for clinicians. *Ther. Adv. Chronic. Dis.* **3**: 87–106.
- 92. Hayut, S.F., Bessudo, C.M. and Levy, A. A. 2017. Targeted recombination between homologous chromosomes for precise breeding in tomato. *Nat. Commun.* 8:15605. doi: 10.1038/ncomms15605
- Hazra, P. and Som. M.G. 1996. Technology for Vegetable Production and improvement. Naya Prokash, Kolkata, India.
- 94. He, Q., Zhang, Z. and Zhang, L., 2016. Anthocyanin accumulation, antioxidant ability and stability, and a transcriptional analysis of anthocyanin biosynthesis in purple heading Chinese cabbage (*Brassica rapa* L. ssp. *pekinensis*). J. Agric. Food Chem. 64: 132–45.
- 95. Horvitz, M.A., Simon, P.W. and Tanumihardjo, S.A., 2004. Lycopene and β-carotene are bioavailable from lycopene 'red'carrots in humans. *Eur. J. Clin. Nutr.* 58: 803.
- 96. Hughes, M. 1948. The inheritance of two characters of *Cucumis melo* and their interrelationship. *Proc. American Soc. Hort. Sci.* **59**: 399–402.
- 97. Huh, J.H., Kang, B.C., Nahm, S.H., Kim, S., Ha, K.S., Lee, M.H. and Kim, B.D. 2001. A candidate gene approach identified phytoene synthase as the locus for mature fruit color in red pepper (*Capsicum spp.*). *Theor. Appl. Genet.* **102**: 524–30.
- Hurst, S., Loeffler, R., Steine, D.L., Amen, M.N.A. and Vafeados D. 2011. Non-transgenic tomato varieties having increased shelf life postharvest due to alterations in betagalactosidase 4. US Patent Application. 20110159168.
- 99. Hutchins, A.E. 1940. Inheritance in the cucumber. *J. Agr. Res.* **60**: 117-28.
- ICMR, 2010. Dietary Guidelines for Indian Populations. Indian Council of Medical Research, New Delhi/National Institute, of Nutrition, Hyderabad.
- 101. Imam, M.K.L., Abo-Bakr, M.A. and Hanna, H.Y. 1972. Inheritance of some economic characters in crosses between sweet melon and

snake cucumber. I. Inheritance of qualitative characters. *Assiut. J. Ag. Sco.* **3**: 363–80.

- 102. Isaacson, T., Ronen, G., Zamir, D. *et al.* 2002. Cloning of tangerine from tomato reveals a carotenoid isomerase essential for the production of beta-carotene and xanthophylls in plants. *Plant Cell*, **14**: 333–42.
- 103. Iwata H., Satoshi Niikura, Seiji Matsuura, Yasushi Takano and Yasuo Ukai. 2004. Genetic control of root shape at different growth stages in radish (*Raphanus sativus* L.). *Breed. Sci.* 54: 117-24.
- 104. Jasim, A. J. 1967. Inheritance of certain characters in okra (*H. esculentus* L.), *Diss. Abstr. Sect.* **28**: 3, 45.
- 105. Jasim, A. J. and Fontenot, J. F. 1967. Inheritance of certain characters in okra (*H. esculentus*), *Diss. Abstr. Sect.* **28**: 211.
- 106. Jenkins, J.A. and Mackinney, G. 1955. Carotenoids of the apricot tomato and its hybrids with yellow and tangerine. *Genetics*, **40**: 715–20.
- 107. Jeong, J. and Guerinot, M.L. 2008. Biofortified and bioavailable: the gold standard for plantbased diets. *Proc. Natl. Acad. Sci. USA* **105**:1777–78.
- 108. Jones, C.M., Mes, P. and Myers, J.R., 2003. Characterization and inheritance of the Anthocyanin fruit (Aft) tomato. *J. Hered.*, **94**(6): 449-456.
- 109. Jones, H. A. and Peterson, C. 1952. Complementary factors for light-red bulb color in onions, *Proc. Am. Soc. Hort. Sci.* **59**: 457.
- 110. Jourdan, M., Gagné, S., Dubois-Laurent, C., Maghraoui, M., Huet, S., Suel, A., Hamama, L., Briard, M., Peltier, D. and Geoffriau, E., 2015. Carotenoid content and root color of cultivated carrot: a candidate-gene association study using an original broad unstructured population. *PLoS One*, **10**: e0116674.
- 111. Jung, C.S., Griffiths, H.M., De Jong, D.M., Cheng, S., Bodis, M. and De Jong, W. S. 2005. The potato *P* locus codes for flavonoid 3', 5'hydroxylase. *Theor. Appl. Genet.* **110**: 269–75.
- Jung, C.S., Griffiths, H.M., De Jong, D.M., Cheng, S., Bodis, M., Kim, T.S. and De Jong, W.S. 2009. The potato developer (D) locus encodes an R<sub>2</sub>R<sub>3</sub> MYB transcription factor that

regulates expression of multiple anthocyanin structural genes in tuber skin. *Theor. Appl. Genet.* **120**: 45–57.

- Jung, H.J., Manoharan, R.K., Park, J.I., Chung, M.Y., Lee, J., Lim, Y.P., Hur, Y. and Nou, I.S. 2014. Identification of yellow pigmentation genes in *Brassica rapa* ssp. *pekinensis* using Br300 microarray. *Int. J. Genom.* **10**: 11.
- 114. Kalia, P., Muthukumar, P., Soi, S. and Shilpi 2018. Marker assisted introgression of the *Or* gene for enhancing β-carotene content in Indian cauliflower. *Acta Hort*. **1203**: 121-27.
- 115. Keller, W. 1936. Inheritance of some major colour types in beets. J. Agric. Sci. (Cambridge), **52**: 27.
- 116. Kianian, S.F. and Quiros, C.F. 1992b. Genetic analysis of major multigene families in Brassica oleracea and related species. *Genome*, **35**: 516–27.
- 117. Kim, S., Yoo, K.S. and Pike, L.M. 2005. Development of a co-dominant, PCR-based marker for allelic selection of the pink trait in onions (*Allium cepa*), based on the insertion mutation in the promoter of the anthocyanidin synthase gene. *Theor. Appl. Genet.* 110:1167
- 118. King, J.J., Bradeen, J.M., Bark, O., McCallum, J.A. and Havey, M.J. 1998a. A low-density genetic map of onion reveals a role for tandem duplication in the evolution of an extremely large diploid genome. *Theor. Appl. Genet.* **96**: 52–62.
- 119. Klap, C., Yeshayahou, E., Bolger, A. M., Arazi, T., Gupta, S. K., Shabtai, S., *et al.* 2017. Tomato facultative parthenocarpy results from SIAGAMOUS-LIKE 6 loss of function. *Plant Biotechnol. J.* **15**: 634–47. doi: 10.1111/ pbi.12662
- 120. Koch, T.C. and Goldman, I.L., 2005. Relationship of carotenoids and tocopherols in a sample of carrot root-color accessions and carrot germplasm carrying Rp and rp alleles. *J. Agric. Food Chem.*, **53**(2):325-331.
- 121. Kolhe, A. K. and D'Cruz, R. 1966. Inheritance of pigmentation in okra. *Indian J. Genet. Plant Breed.* **23:** 112.
- 122. Kooistra, E. 1971. Inheritance of flesh and skin colors in powdery mildew resistant cucumbers (*Cucumis sativus* L.). *Euphytica*, **20**: 521-23.

- 123. Kopsell, D.A., Kopsell, D.E. 2006. Accumulation and bioavailability of dietary carotenoids in vegetable crops. *Trends Plant Sci.* 11: 499–507.
- 124. Kormos, J. ancl Kormos, K.1960. Die Genetischen Typen der carotenoid - systeme de paprika fruit. *Acta Sci*. Hung, **6**: 305\_ te (c.f.
- 125. Ku, H.M., Doganlar, S., Chen K.Y. and Tanksley, S.D. 1999. Genetic basis of pear-shaped fruit in tomato. *Theor. Appl. Genet.* **99**: 844–50
- 126. Kumari, A., Chawla, N. and Dhatt, A.S., 2018. Genotypic Differences for anthocyanins in different parts of eggplant (*Solanum melongena* L.). *Int. J. Adv. Res. Biol. Sci.* **5**:22-15.
- 127. Kushad, M. M., Brown, A. F., Kurilich A. C., Juvik, J. A., Klein, B. P., Wallig, M. A. and Jeffery, E. H. 1999. Variation of glucosinolates in vegetable crops of *Brassica oleracea. J. Agric. Food Chem.* **47:** 1541–48.
- 128. Kust, A.F. 1970. Inheritance and differential formation of color and associated pigments in xylem and phloem of carrot (*Daucus carota* L.) Ph.D. Thesis, Univ. of Wisconsin: pp.42.
- 129. Kwan, C. C. 1934. Inheritance of some plant characters in cabbage, *Brassica oleracea* var. *capitata. J. Agric. Assoc.* China No. 126–127: 81–127.
- 130. Kyriacou, M.C. and Rouphael, Y. 2018. Towards a new definition of quality for fresh fruits and vegetables. *Sci. Hortic.*, 234: 463-469
- Lefebvre, V., Kuntz, M., Camara, B. and Palloix, A., 1998. The capsanthin-capsorubin synthase gene: a candidate gene for the *y* locus controlling the red fruit colour in pepper. *Plant Mol. Biol.*, **36**(5): 785-789.
- 132. Levin, I., Frankel, P., Gilboa, N. *et al.* 2003. The tomato dark green mutation is a novel allele of the tomato homolog of the DEETIOLATED1 gene. *Theor. Appl. Genet.* **106**: 454–60.
- 133. Li, L. and Garvin, D.F. 2003. Molecular mapping of *Or*, a gene inducing β-carotene accumulation in cauliflower (*Brassica oleracea* L. var. *botrytis*). *Genome*, **46**: 588–94.
- 134. Li, L., Paolillo, D.J., Parthasarathy, M.V., DiMuzio, E.M. and Garvin, D.F 2001. A novel

gene mutation that confers abnormal patterns of b-carotene accumulation in cauliflower (*Brassica oleracea* var. *botrytis*). *Plant J.* **26**: 59–67.

- 135. Li, R., Li, R., Li, X., Fu, D., Zhu, B., Tian, H., Luo, Y. and Zhu, H. 2018. Multiplexed CRISPR/ Cas9-mediated metabolic engineering of γ-aminobutyric acid levels in *Solanum lycopersicum*. *Plant Biotechnol. J.* **16**: 415–27.
- 136. Li, X., Wang, Y., Chen, S., Tian, H., Fu, D., Zhu, B., Luo, Y. and Zhu, H. 2018. Lycopene is enriched in tomato fruit by CRISPR/Cas9mediated multiplex genome editing. *Front. Plant Sci.* **9**: 559.
- 137. Lioi, L., Galasso, I., Lanave, C., Daminati, M.G., Bollini, R. and Sparvoli, F. 2007. Evolutionary analysis of the APA genes in the Phaseolus genus: wild and cultivated bean species as sources of lectin-related resistance factors? *Theor. Appl. Genet.* **115**: 959–70.
- 138. Lippert, L. F. 1975. Hererosis and combining ability in chilli peppers by dialiet analysis. *Crop Sci.* **15**: 323-25.
- 139. Liu, J.P., Van Eck, J., Cong, B. *et al.* 2002. A new class of regulatory genes underlying the cause of pear-shaped tomato fruit. *Proc. Natl. Acad. Sci. USA*, **99**: 13302–06.
- 140. Liu, X., Gao, B., Han, F., Fang, Z., Yang, L, Zhuang, M. *et al.* 2017. Genetics and fine mapping of a purple leaf gene, *BoPr*, in ornamental kale (*Brassica oleracea* L. var. *acephala*). *BMC Genomics*, **18**: 230
- 141. Longvah, T., Ananthan, R., Bhaskarachary, K. and Venkaiah, K. 2017. Indian Food Composition Tables 2017. National Institute of Nutrition, Hyderabad.
- 142. Lorizzo Massimo, Cavagnaro Pablo, F., Bostan Hamed, Zhao Yunyang, Zhang Jianhui and Simon Philipp W. **2019**. A cluster of MYB transcription factors regulates anthocyanin biosynthesis in carrot (*Daucus carota* L.) root and petiole. *Front. Plant Sci.* **9**: 1927
- 143. Lu, S., Van Eck, J., Zhou, X., Lopez, A.B., O'Halloran, D.M., Cosman, K.M., Conlin, B.J. *et al.* 2006. The cauliflower *Or* gene encodes a DnaJ cysteine-rich domain-containing protein that mediates high levels of beta-carotene accumulation. *Plant Cell*, **18**: 3594–3605.

- 144. Magruder, R. and C. H., Myers, 1933. The inheritance of some plant colors in cabbage. *J. Agric. Res.* **47**: 233–48.
- 145. Maligeppagol, M., Chandra, G. S., Navale, P. M., Deepa, H., Rajeev, P. R. *et al.* 2013. Anthocyanin enrichment of tomato (*Solanum lycopersicum* L.) fruit by metabolic engineering. *Curr. Sci.* **105**: 72-80.
- 146. Matsui, K., Umemura, Y., Ohme-Takagi, M., 2008. AtMYBL2, a protein with a single MYB domain, acts as a negative regulator of anthocyanin biosynthesis in *Arabidopsis*. *Plant J.* **55**: 954–67.
- 147. McCallum, C.M., Slade A.J., Colbert, T.G., Knauf, V.C. and Hurst, S. 2008. Tomatoes having reduced polygalacturonase activity caused by non-transgenic mutations in the polygalacturonase gene. US Patent 7393996.
- 148. McCallum, J., Clarke, A., Pither-Joyce, M., Shaw, M., Butler, R., Brash, D., Scheffer, J., Sims, I., van Heusden, S., Shigyo M and Havey, M. 2006. Genetic mapping of a major gene affecting onion bulb fructan content. *Theor. Appl. Genet.* **112**: 958–67
- 149. Meli, V. S., Ghosh, S., Prabha, T. N., Chakraborty, N., Chakraborty, S. and Datta, A. (2010). Enhancement of fruit shelf life by suppressing N-glycan processing enzymes. Proc. Natl. Acad. Sci. USA, **107**: 2413–18.
- Menard, R., Larue, J., Silue, D., Thouvenot, D. 1999. Glucosinolates in cauliflower as biochemical markers for resistance against downy mildew. *Phytochem.* 52: 29–35.
- 151. Mes, P.J., Boches, P., Myers, J.R. *et al.* 2008. Characterization of tomatoes expressing anthocyanin in the fruit. *J. American Soc.Hort. Sci.* 133: 262–69.
- 152. Michael, J. and Havey 2020. Genetic Mapping of Chartreuse Bulb Color in Onion. *J. Am. Soc. Hort. Sci.* **145**: 110–19.
- 153. Minoia, S.A., Petrozza, O., D'Onofrio, F., Piron, G., Mosca, G., Sozio, F., Cellini, A. and Bendahmane F Carriero. 2010. A new mutant genetic resource for tomato crop improvement by TILLING technology. *BMC Res Notes*, **3**: 69.

- 154. Mithen, R., Faulkner, K., Magrath, R., Rose, P., Williamson, G. and Marquez, J., 2003. Development of isothiocyanate-enriched broccoli, and its enhanced ability to induce phase 2 detoxification enzymes in mammalian cells. *Theor. Appl. Genet.* **106**: 727-34.
- 155. Miyatake, K., Saito, T., Negoro, S. *et al.* 2012. Development of selective markers linked to a major QTL for parthenocarpy in eggplant (*Solanum melongena* L.). *Theor. Appl. Genet.* **124**: 1403-13.
- 156. Molesini, B., Pandolfini, T., Rotino, G. L., Dani, V. and Spena, A. 2009. *Aucsia* gene silencing causes parthenocarpic fruit development in tomato. *Plant Physiol*. **149**: 534–48.
- 157. Moore, S., Vrebalov, J., Paxton, P. and Giovannoni, J. 2002. Use of genomics tools to isolate key ripening genes and analyse fruit maturation in tomato. J. Exp. Bot. 53: 2023–30.
- 158. More, D. C, Patil, S. B. and Nimbalkar, V. S. 1982. Inheritance of some characters in brinjal cross SM<sub>2</sub> × Nimbkar Green Round. *J. Maharashtra Agric. Univ.* 7: 243.
- 159. Moreno, J.C., Pizarro, L., Fuentes, P., Handford, M., Cifuentes, V. and Stange, C. 2013. Levels of lycopene β-cyclase 1 modulate carotenoid gene expression and accumulation in *Daucus carota*. *PLoS One*, 8: e58144.
- Morris, J., Hawthorne, K.M., Hotze, T., Abrams, S.A. and Hirschi, K.D. 2008. Nutritional impact of elevated calcium transporter activity in carrots. *Proc. Natl. Acad. Sci. USA*, **105**: 1431–5.
- Mou, B., 2008. Evaluation of oxalate concentration in the US spinach germplasm collection. *HortSci.* 43: 1690-93.
- 162. Mustilli, A.C., Fenzi, F., Ciliento, R. *et al.* 1999. Phenotype of the tomato high pigment-2 mutant is caused by a mutation in the tomato homolog of *DEETIOLATED1*. *Plant Cell*, **11**: 145–57.
- 163. Muth, J., S. Hartje, R. M. Twyman, H. R. Hofferbert, E. Tacke, and D. Prüfer. 2008. Precision breeding for novel starch variants in potato. *Plant Biotech. J.* 6: 576–84.
- 164. Natesh, H.N., Abbey, L. and Asiedu, S.K., 2017. An overview of nutritional and antinutritional factors in green leafy vegetables. *Hort. Int. J.* **1**: 00011.

- 165. Nimmakayala, P., Abburi, V. L., Saminathan, T., Alaparthi, S. B., Almeida, A., Davenport, B., Nadimi, M., Davidson, J. *et al.* 2016. Genomewide diversity and association mapping for capsaicinoids and fruit weight in *Capsicum annuum* L. *Sci. Rep.* **6**: 38081.
- 166. Okabe, Y., Saito, E.T., Matsukura, C., Ariizumi, T., Mizoguchi, T. and Ezura, H. 2011. Tomato TILLING technology: development of a reverse genetics tool for the efficient isolation of mutants from Micro-Tom mutant libraries. *Plant Cell Physiol.* doi:10.1093/ pcp/pcr134.
- 167. Pan, C., Ye, L., Qin, L., Liu, X., He, Y., Wang, J., et al. 2016. CRISPR/Cas9- mediated efficient and heritable targeted mutagenesis in tomato plants in the first and later generations. Sci. Rep. 6: 24765. doi: 10.1038/srep24765
- 168. Pandey, M., Verma, R. K. and Saraf, S. A. 2010. Nutraceuticals: new era of medicine and health. *Asian J. Pharm. Clin. Res.* **3**: 11-15.
- 169. Pang, W., Kim, Y. Y., Li, X., Choi, S. R., Wang, Y., Sung, C. K., Im, S., Ramchiary, N., Zhou, G. and Lim, Y. P. 2015. Anatomic characteristics associated with head splitting in cabbage (*Brassica oleracea* var. *capitata* L.). *PloS one*, **10:** e0142202.
- 170. Panthee, D.R., Labate, J.A. and Robertson, L.D., 2013. Evaluation of tomato accessions for flavour and flavour-contributing components. *Plant Genet. Resour.* **11**:106-13.
- 171. Park, S., Elless, M., Park, J., Jenkins, A., Lim, W., Chambers I.V. E. and Hirschi, K. 2009. Sensory analysis of calcium-biofortified lettuce. *Plant Biotech. J.* **7**: 106-17.
- 172. Park, S., Kang, T.S., Kim, C.K. *et al.* 2005. Genetic manipulation for enhancing calcium content in potato tuber. *J. Agric. Food Chem.* **53**: 5598–5603.
- 173. Patidar, D. 2015. DUS and qualitative characters inheritance studies of brinjal (*Solanum melongena* L.) genotypes. *Trends Biosci.* 178-80.
- 174. Patil S.K., and Moore D.C. 1983. Inheritance studies in brinjal. *J. Maharashtra Agric. Univ.* 8: 43-45.
- 175. Patil, S. K. and More, D. C. 1983. Genetics of pigmentation in brinjal, *J. Maharashtra Agric. Univ.* **8**: 126.

- 176. Pederson, A. 1944. The colours of beets (*Beta vulgaris* L.). *K. Vet-of-Landbokojsk Arsskr.*, p. 60.
- 177. Pereira, L., Ruggieri, V., Pérez, S., Alexiou, K.G., Fernández, M., Jahrmann, T., Pujol, M. and Garcia-Mas, J. 2018. QTL mapping of melon fruit quality traits using a high-density GBS-based genetic map. *BMC Plant Biol.* 18: 324
- 178. Peters, S., Imani, J., Mahler, V., Foetisch, K., Kaul, S., Paulus, K.E., Scheurer, S., Vieths, S. and Kogel, K.H. 2011. Dau c 1.01 and Dau c 1.02-silenced transgenic carrot plants show reduced allergenicity to patients with carrot allergy. *Transgenic Res.* **20**: 547-56.
- 179. Pilch, S. 1987. Physiological effects and health consequences of dietary fibre. Bethesda, M.D.: Life Science Research Office. Federation of American Societies for Experimental Biology. (Abstr.)
- 180. Platel, K. and Srinivasan, K., 2016. Bioavailability of micronutrients from plant foods: an update. *Crit Rev. Food Sci. Nutri.* **56**: 1608-19.
- Poole, C.F. and Grimball, P.C. 1945. Interaction of sex, shape, and weight genes in watermelon. *J. Agric. Res.* **71**: 533-52.
- 182. Prohens, J., Gramazio, P., Plazas, M., Dempewolf, H., Kilian, B. *et al.* 2017. Introgressiomics: a new approach for using crop wild relatives in breeding for adaptation to climate change. *Euphytica*, **213**:158 DOI 10.1007/s10681-017-1938-9
- 183. Ramchiary, N., Bisht, N.C., Gupta, V., Mukhopadhyay, A., Arumugam, N., Sodhi, Y.S., Pental, D. and Pradhan, A.K., 2007. QTL analysis reveals context-dependent loci for seed glucosinolate trait in the oilseed *Brassica juncea*: Importance of recurrent selection backcross scheme for the identification of 'true' QTL. *Theor. Appl. Genet.* **116**: 77–85.
- 184. Ramsay G., Griffiths D.W. and Dow N.D. 1991. Spontaneous and induced variation in levels of vicine and convicine in faba beans. *Aspects Appl. Biol. Prod. Prot. Legumes*, **27**: 43-47
- 185. Rao, G.U. and Paran, I. 2003. Polygalacturonase: a candidate gene for the soft flesh and deciduous fruit mutation in Capsicum. *Plant Mol. Biol.* 51: 135–41

- 186. Rao, G.U., Ben Chaim, A., Borovsky, E. and Paran, I. 2003. Mapping of yield related QTLs in pepper in an interspecific cross of *Capsicum annuum* and *C. frutescens. Theor. Appl. Genet.* **106**: 1457–66.
- 187. Ren, J., Chen, Z., Duan, W., Song, X., Liu, T., Wang, J., Hou, X. and Li, Y. 2013. Comparison of ascorbic acid biosynthesis in different tissues of three non-heading Chinese cabbage cultivars. *Plant Physiol. Biochem.* **73**: 229–36.
- 188. Ren, J., Duan, W., Chen, Z., Zhang, S., Song, X., Liu, T., Hou, X. and Li, Y. 2015. Overexpression of the monodehydroascorbate reductase gene from non-heading Chinese cabbage reduces ascorbate level and growth in transgenic tobacco. *Plant Mol. Biol. Rep.* 33: 881–92.
- 189. Rick, C.M., Cisneros, P., Chetelat, R.T. and DeVerna, J.W. 1994. Abg –A gene on chromosome 10 for purple fruit derived from S. *lycopersicoides*. Tomato Genet. Coop. Rep. 44: 29–30.
- 190. Rieman, G. H. 1931. Genetic factors for pigmentation in the onion and their relation to disease resistance. *J. Agric. Res.* **42**: 251.
- 191. Rodríguez-Leal, D., Lemmon, Z. H., Man, J., Bartlett, M. E. and Lippman, Z. B. 2017. Engineering quantitative trait variation for crop improvement by genome editing. Cell, 171:470–80
- Romer, S., Fraser, P.D., Kiano, J.W., Shipton, C.A., Misawa, N., Schuch, W. and Bramley, P.M. 2000. Elevation of the pro-vitamin A content of transgenic tomato plants. *Nature Biotech.* 18: 666–69.
- 193. Ronen, G., Carmel-Goren, L., Zamir, D. and Hirschberg, J. 2000. An alternative pathway to β-carotene formation in plant chromoplasts discovered by map-based cloning of Beta and old-gold color mutations in tomato. *Proc. Natl. Acad. Sci. USA*, **97**:11102–07.
- 194. Ronen, G.L., Cohen, M., Zamir, D. and Hirschberg J. 1999. Regulation of carotenoid biosynthesis during tomato fruit development: expression of the gene for lycopene epsiloncyclase is down-regulated during ripening and is elevated in the mutant Delta. *Plant J.* **17**: 341–51.

- 195. Ruggieri, V., Francese, G., Sacco, A., D'Alessandro, A., Rigano, M.M., Parisi, M., Milone, M., Cardi, T., Mennella, G. and Barone, A. 2014. An association mapping approach to identify favourable alleles for tomato fruit quality breeding. *BMC Plant Biol.*, 14: 337.
- 196. Sankhari, J.M., Thounaojam, M.C., Jadeja, R.N., Devkar, R.V. and Ramachandran, A.V. 2012. Anthocyanin-rich red cabbage (*Brassica oleracea* L.) extract attenuates cardiac and hepatic oxidative stress in rats fed an atherogenic diet. *J. Sci. Food Agric.* **92**: 1688–93.
- 197. Santos, C.A. and Simon, P.W., 2004. Merging carrot linkage groups based on conserved dominant AFLP markers in F2 populations. *J. Am. Soc. Hortic. Sci.*, **129**,: 211-217.
- 198. Santos, C.A.F. and Simon, P.W. 2002a. QTL analyses reveal clustered loci for accumulation ofmajor provitamin A carotenes and lycopene in carrot roots. *Mol. Genet. Genom.* 268: 122–29
- 199. Santos, C.A.F., Senalik, D.A. and Simon, P. W. 2005. Path analysis suggests phytoene accumulation is the key step limiting the carotenoid pathway in white carrot roots. *Genet. Mol. Biol.*, **28**: 287-293
- 200. Sapir, M. Shamir, M.O., Ovadia, R., Reuveni, M., Evenor, D., Tadmor, Y. *et al.* 2008. Molecular aspects of *anthocyanin fruit* tomato in relation to *high pigment-1. J. Hered.* **99**: 292-303.
- 201. Sarikamis, G., Marquez, J., MacCormack, R., Bennett, R.N., Roberts, J. and Mithen, R., 2006. High glucosinolate broccoli: a delivery system for sulforaphane. *Mol. Breed.* 18(3):219-228.
- 202. Sarpras, M., Gaur, R., Sharma, V., Chhapekar, S.S., Das, J., Kumar, A., Yadava, S.K., Nitin, M., Brahma, V., Abraham, S.K. and Ramchiary, N., 2016. Comparative analysis of fruit metabolites and pungency candidate genes expression between Bhut Jolokia and other *Capsicum* species. *PLoS One*, **11**: e167791. Doi:10.1371/ journal.pone.0167791
- 203. Saubade, F., Hemery, Y.M., Guyot, J.P. and Humblot, C. 2016. Lactic acid fermentation as a tool for increasing the folate content of foods. *Crit. Rev. Food Sci. Nutr.* **57**: 3894-3910.

- 204. Sauvage, C., Segura, V., Bauchet, G., Stevens, R., Do, P.T., Nikoloski, Z., *et al.* 2014. Genomewide association in tomato reveals 44 candidate loci for fruit metabolic traits. *Plant Physiol.* **165**:1120-32.
- 205. Savadogo, A., Flibert, G. and Tapsoba, F. 2016. Probiotic microorganisms involved in cassava fermentation for Gani and Attieke production. *J. Advan. Biotech.* **6**: 858-68.
- 206. Schonhof, I., Krumbein, A. and Brückner, B. 2004. Genotypic effects on glucosinolates and sensory properties of broccoli and cauliflower. *Food Nahrung*, **48**:25-33.
- 207. Seymour, G.B., Manning, K., Eriksson, E.M. *et al.* 2002. Genetic identification and genomic organization of factors affecting fruit texture. *J. Exp. Bot.* **53**: 2065–71.
- 208. Shi, J., Wang, H., Schellin, K., Li, B., Faller, M., Stoop, J.M., Meeley, R.B., Ert, D.S., Ranch, J.P. and Glassman, K. 2007. Embryo-specific silencing of a transporter reduces phytic acid content of maize and soybean seeds. *Nature Biotech.* 25: 930–37.
- 209. Singh, S., Kalia, P., Meena, R.K., Mangal, M., Islam, S., Saha, S. and Tomar, B.S. 2020. Genetics and expression analysis of anthocyanin accumulation in curd portion of sicilian purple to facilitate biofortification of Indian Cauliflower. *Front. Plant Sci.* **10**: 1766. doi: 10.3389/fpls.2019.01766
- Singh, S., Singh, D. R., Salim, K. M., Srivastava, A., Singh, L. B., and Srivastava, R. C. 2011. Estimation of proximate composition, micronutrients and phytochemical compounds in traditional vegetables from Andaman & Nicobar Islands. *Int. J. Food Sci. Nutr.* 62: 765–73.
- 211. Singh, S., Singh, L.B., Singh, D.R., Chand, S., Ahmed, S.Z., Singh, V.N. and Roy, S.D., 2018. Indigenous underutilized vegetables for food and nutritional security in an island ecosystem. *Food Sec.* **10**: 1173-89.
- 212. Singh, S., Swain, S., Nisha, M., Banu, V.S., Singh, D.R. and Roy, S.D., 2015b. Changes in lycopene, total carotenoid and anti-radical activity in teasel gourd [*Momordica subangulata* ssp. *renigera* (G. Don) de Wilde] fruit fractions at different stages of maturity. *Ind. Crops Prod.* **73**:154-63.

- 213. Singh, S., Swain, S., Singh, D.R., Salim, K.M., Nayak, D. and Roy, S.D., 2015a. Changes in phytochemicals, anti-nutrients and antioxidant activity in leafy vegetables by microwave boiling with normal and 5% NaCl solution. *Food Chem.* **176:** 244-53.
- 214. Siritunga, D. and Sayre, R.T. 2003. Generation of cyanogen-free transgenic cassava. *Planta*, **217**: 367-73.
- 215. Som MG and Hazra P (1996). Vegetable Crops. Naya Prokash, Kolkata, India.
- 216. Sonnante G, De Paolis A, Pignone D (2005) Bowman–Birk in-hibitors in lens: identification and characterization of twoparalogous gene classes in cultivated lentil and wild relatives. Theor Appl Gen 110:596–604
- Stevens, R., Page, D., Gouble, B. *et al.* 2008. Tomato fruit ascorbic acid content is linked with monodehydroascorbate reductase activity and tolerance to chilling stress. *Plant, Cell Environ.* 31: 1086–96.
- 218. Stewart Jr, C., Kang, B.C., Liu, K., Mazourek, M., Moore, S.L., Yoo, E.Y. *et al.* 2005. The Pun1 gene for pungency in pepper encodes a putative acyltransferase. *Plant J.*, **42**(5): 675-688.
- 219. Stommel, J. R., Lightbourn, G. J., Winkel, B. S., and Griesbach, R. J. 2009. Transcription Factor Families Regulate the Anthocyanin Biosynthetic Pathway in *Capsicum annuum*. *J. American Soc. Hort. Sci.* **134**: 244-51.
- 220. Su, L., Diretto, G., Purgatto, E. *et al.* 2015. Carotenoid accumulation during tomato fruit ripening is modulated by the auxin-ethylene balance. *BMC Plant Biol.* **15:** 114, doi. org/10.1186/s12870-015-0495-4
- 221. Swaminathan, S., Hemalatha, R., Pandey, A., Kassebaum, N.J., Laxmaiah, A., Longvah, T. *et al.* 2019. The burden of child and maternal malnutrition and trends in its indicators in the states of India: the Global Burden of Disease Study 1990–2017. *Lancet Child Adolesc. Health*, **3**: 855-70.
- 222. Swarup V. (2014). Vegetable Science and Technology in India. Kalyani Publishers, India.
- Talsma, E., Brouwer, I., Verhoef, H., Mbera, G., Mwangi, A., Demir, A., Maziya-Dixon, B., Boy, E., Zimmermann, M., Melse-Boonstra, A. 2016.

Biofortified yellow cassava and vitamin A status of Kenyan children: a randomized controlled trial. *American J. Clin. Nutr.* **103**: 258–67.

- 224. Tatebe T. 1938. On inheritance of root color in *Raphanus sativus* Linn. *Japanese. J. Genet.* **14**: 39-50.
- 225. Tatsuzawa, F., Toki, K., Saito, N., Shinoda, K., Shigihara, A. and Honda, T. 2008. Anthocyanin occurrence in the root peels, petioles and flowers of red radish (*Raphanus sativus* L.). *Dyes Pigm.* **79**: 83-88.
- 226. Thakur, M.R., Singh, K. and Singh, J. 1969. Inheritance of some qualitative characters in brinjal (*Solanum melongena* L.). *J. Res. PAU*, **6**: 769-75.
- 227. Thompson, A.J., Tor, M., Barry, C.S. *et al.* 1999, Molecular and genetic characterization of a novel pleiotropic tomato-ripening mutant. *Plant Physiol.* **120**: 383–90.
- 228. Thorwarth, P., Yousef, E.A.A. and Schmid, K. J. 2018. Genomic Prediction and Association Mapping of Curd-Related Traits in Gene Bank Accessions of Cauliflower. *G3* (Bethesda), **8**:707-18.
- 229. Tigchelaar, E.C., Janick, J. and Erickson HT. 1968. The genetics of anthocyanin coloration in eggplant (*Solanum melongena* L.). *Genetics*, **60**:475–91.
- Tofino, A., Cabal, D., Ceballos, H., Pérez, J.C. and H. M. Romero. 2009. Validation of TILLING in the evaluation of inbred progenies of irradiated cassava (*Manihot esculenta* Crantz). *Agron. Colomb.* 27: 313-21.
- 231. Tomes, M.L., Quackenbush, F.W. and McQuistan, M. 1954. Modification and dominance of the gene governing formation of high concentrations of beta-carotene in the tomato. *Genetics*, **39**: 810–17.
- 232. Traka, M.H., Saha, S., Huseby, S. and *et al.* 2013. Genetic regulation of glucoraphanin accumulation in Beneforté® broccoli. *New Phytol.* **198**: 1085-95.
- 233. Trebbi, D. 2005. Genetic analysis of sucrose accumulation in sugar beet (*Beta vulgaris* L.). Dissertation, Michigan State University.
- 234. Tsuro, M., Keita, Suwabe, Nakao, Kubo, Satoru, Matsumoto, Masashi and Hirai. 2008.

Mapping of QTLs controlling root shape and red pigmentation in radish, *Raphanus sativus* L. *Breed. Sci.* **58**: 55-61.

- 235. Tzuri, G., Zhou, X., Chayut, N., Yuan, H., Portnoy, V. *et al.* 2015. A 'golden' SNP in *CmOr* governs fruit flesh color of melon (*Cucumis melo*). *Plant J.* 82: 267–79.
- 236. Ueta, R., Abe, C., Watanabe, T., Sugano, S. S., Ishihara, R., Ezura, H. *et al.* 2017. Rapid breeding of parthenocarpic tomato plants using CRISPR/Cas9. *Sci. Rep.* **7**: 507. doi: 10.1038/s41598-017-00501-4
- 237. Uluisik, S., Chapman, N.H., Smith, R., Poole, M., Adams, G., Gillis, R.B. *et al.* 2016. Genetic improvement of tomato by targeted control of fruit softening. *Nature Biotech.* **34**: 950-52.
- 238. Uphof, J.C. 1924. On Mendelian factors in radishes. *Genetics*, **9**: 292-304
- 239. van Eck, H.J., Jacobs, J.M., van den Berg, P.M., Stiekema, W.J. and Jacobsen E. 1994. The inheritance of anthocyanin pigmentation in potato (*S. tuberosum* L.) and mapping of tuber skin color loci using RFLPs. *Heredity*, **73**: 410–21.
- 240. Vijaya Bhaskar, A.V., Nithya, D.J., Raju, S. et al. 2017. Establishing integrated agriculturenutrition programmes to diversify household food and diets in rural India. Food Secur. 9: 981.
- 241. Vrebalov, J., Ruezinsky, D., Padmanabhan, V., White, R., Medrano, D., Drake, R. *et al.* 2002. A MADS-box gene necessary for fruit ripening at the tomato ripening inhibitor (Rin) locus. *Science*, **296**: 343–46.
- 242. Wahlroos, T., Susi, P., Solovyev, A., Dorokhov, Y., Morozov, S, Atabekov, J. and Korpela, T. 2004. Increase of histidine content in *Brassica rapa* subsp. *oleifera* by over-expression of histidine-rich fusion proteins. *Mol. Breed.* 14: 455-62.
- 243. Walker, A.R., Davison, P.A., Bolognesi-Winfield, A.C., James, C.M., Srinivasan, N. *et al.* 1999. The *TRANSPARENT TESTA GLABRA1* locus, which regulates trichome differentiation and anthocyanin biosynthesis in *Arabidopsis*, encodes a WD40 repeat protein. *Plant Cell*, **11**: 1337–50.

- 244. Wang, D., Zhang, C., Wang, B., Li, B., Wang, Q., Liu, D. *et al.* 2019. Optimized CRISPR guide RNA design for two high-fidelity Cas9 variants by deep learning. *Nature Commun.* **10**: 1-14.
- 245. Wang, H., Ou, C. G., Zhuang, F. Y., and Ma, Z. G. 2014. The dual role of phytoene synthase genes in carotenogenesis in carrot roots and leaves. *Mol. Breed.: New Strategies In Plant Improv.* **34**: 2065–79.
- 246. Watson, J.F. and Gabelman, W.H. 1984. Genetic analysis of betacyanine, betaxanthine and sucrose concentrations in roots of table beet. *J. American Soc. Hort. Sci.* **109**: 386.
- 247. Wehner, T.C. 2007. The gene list for watermelon. *Cucurbit Genet. Coop. Rpt.* **30**: 96–120.
- 248. Welch, R.M. 1995. Micronutrient nutrition of plants. Crit. Rev. Plant Sci. 14: 49-82.
- 249. Welch, R.M. and Graham, R.D. 2004. Breeding for micronutrients in staple food crops from a human nutrition perspective. *J. Exp. Bot.* **55**: 353-64.
- 250. White, P.J. and Broadley, M.R. 2009. Biofortification of crops with seven mineral elements often lacking in human diets–iron, zinc, copper, calcium, magnesium, selenium and iodine. *New Phytologist*, **182**: 49-84.
- 251. Wiesner, M., Zrenner, R., Krumbein, A., Glatt, H. and Schreiner, M. 2013. Genotypic variation of the glucosinolate profile in pak choi (*Brassica rapa* ssp. *chinensis*). *J. Agric. Food Chem.* **61**: 1943–53.
- 252. Wilkinson, J.Q., Lanahan, M.B., Yen, H.C., Giovannoni, J.J. and Klee H.J. 1995. An ethylene-inducible component of signaltransduction encoded by Never-Ripe. *Science*, **270**: 1807–09.
- 253. Williams, M., Clark, G., Sathasivan, K. and Islam, A.S. 2004. RNA Interference and its application in crop improvement. *Plant Tissue Cult. Biotech.* 1-18. doi=10.1.1.98.1069.
- 254. Willits, M.G., Kramer, C.M., Prata, R.T.N. *et al.* 2005. Utilization of the genetic resources of wild species to create a nontransgenic high flavonoid tomato. *J. Agric. Food Chem.* 53: 1231–36.
- 255. Wolters, A.M., Uitdewilligen, J.G.A.L., Kloosterman, B.A., Hutten, R.C.B., Visser,

R.G.F. and van Eck, H.J. (2010). Identification of alleles of carotenoids pathwa genes important for zeaxanthin accumulation in potato tubers. *Plant Mol. Biol.* **73**: 659-71.

- 256. World Food Summit, 1996. Report of the *World Food Summit*. Food and Agriculture Organization of the United Nations. Rome, 13-17.
- 257. Wu, H., Dushenkov, S., Ho, C.T. *et al.* 2009. Novel acetylated flavonoid glycosides from the leaves of *Allium ursinum*. *Food Chem.* 115: 592–95.
- 258. Xiao, H., Jiang, N., Schaffner, E. *et al.* 2008. A retrotransposon-mediated gene duplication underlies morphological variation of tomato fruit. *Science*, **319**: 1527–30.
- 259. Xiaonan, L.I., Wenxing, P.A. and Zhongyun, P.I. 2017. Omics meets phytonutrients in vegetable *Brassicas*: For nutritional quality breeding. *Hort. Plant J.* **3**: 247-54.
- 260. Yi,I G., June-Sik, Kim, Jeong, Eun Park, Hosub, Shin, Seung, Hwa Yu, Suhyung, Park, Jin and Hoe Huh. 2018. MYB1 transcription factor is a candidate responsible for red root skin in radish (*Raphanus sativus* L.). *PLoS One*, **13**: e0204241.
- 261. Zaki, H.E.M., Shuji, Yokoi, Yoshihito and Takahata. 2010. Identification of genes related to root shape in radish (*Raphanus sativus*) using suppression subtractive hybridization. *Breed. Sci.* **60**: 130-38.
- 262. Zhang Y., Jung C. S., De Jong W. S. (2009). Genetic analysis of pigmented tuber flesh in potato. *Theor. Appl. Genet.* 119 143–150.
- 263. Zhang, B., Hu, Z., Zhang, Y., Li, Y., Zhou, S. and Chen, G., 2012. A putative functional *MYB* transcription factor induced by low temperature regulates anthocyanin biosynthesis in purple kale (*Brassica oleracea* var. *acephala* f. *tricolor*). *Plant Cell Rep.* **31**: 281–89.
- 264. Zhang, F., Wang, G., Wang, M., Liu, X., Zhao, X., Yu, Y., Zhang, D. and Yu, S. 2008. Identification of SCAR markers linked to or, a gene inducing beta-carotene accumulation in Chinese cabbage. *Euphytica*, **164**: 463–71.

- 265. Zhang, J., Yuan, H., Fei, Z., Pogson, B.J., Zhang, L. and Li, L. 2015a. Molecular characterization and transcriptome analysis of orange head Chinese cabbage (*Brassica rapa* L. ssp. *pekinensis*). *Planta*, **241**: 1381–94.
- 266. Zhang, J.X., Li, H.X., Zhang, M.K., Hui, M., Wang, Q., Li, L. and Zhang, L.G., 2013. Fine mapping and identification of candidate *Br-or* gene controlling orange head of Chinese cabbage (*Brassica rapa* L. ssp. *pekinensis*). *Mol. Breed.* 32: 799–805.
- 267. Zhang, Y., Chen, G., Dong, T., Pan, Y., Zhao, Z., Tian, S. and Hu, Z. 2014. Anthocyanin accumulation and transcriptional regulation of anthocyanin biosynthesis in purple bok choy (*Brassica rapa* var. *chinensis*). *J. Agric. Food Chem.* **62**: 12366–76.
- 268. Zhang, Y., Hu, Z., Zhu, M., Zhu, Z., Wang, Z., Tian, S. and Chen, G. 2015b. Anthocyanin accumulation and molecular analysis of correlated genes in purple kohlrabi (*Brassica oleracea* var. *gongylodes* L.). *J. Agric. Food Chem.* **63**: 4160–69.
- 269. Zhao, J., Sauvage, C., Zhao, J. et al. 2019. Meta-analysis of genome-wide association studies provides insights into genetic control of tomato flavor. *Nature Commun*. 10: 1534. https://doi.org/10.1038/s41467-019-09462-w
- 270. Zhou, X., Van Eck, J. and Li, L. 2008. Use of the cauliflower *Or* gene for improving crop nutritional quality. *Biotechnol. Annu. Rev.* **14**: 171–90
- 271. Zivanov, D., Vasiljevic, S., Nikolic, Z., Đordevic, V., Ramazanova, R., Milosevic, B. and Petrovic, G. 2018. A potential use of wild pea as a source of lower trypsin inhibitor activity. *Mol. Breed.* 38: 108.
- 272. Zou, C.L., Zheng, Y., Wang, P., Zhang, X., Wang, Y.H., Liu, Z.Y. and Feng, H., 2016. Fine mapping and characterization of the or gene in Chinese cabbage (*Brassica rapa* L. ssp *pekinensis*). *Genet. Mol. Res.* 15.

Received : December, 2019; Reviewed : February 2020; Revised : February, 2020; Accepted : March 2020