

Assessment of genetic diversity in guava

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ABSTRACT

An extensive survey was made to the diversity rich areas of Gujarat during the year 2016 and 2017 to earmark the elite guava genotypes having desirable traits in terms of shape, size, pulp colour, seed content, morphological and qualitative attributes of fruits. Twenty five genotypes of seedling origin were studied for their quantitative and qualitative characters. Fruit weight ranged between 53.50-318.50 g; length 4.09-9.53 cm; width 4.30-7.90 cm; length:width ratio 0.86-1.40; seed core length 2.70-5.25 cm, pulp thickness 1.00-2.25 cm; number of seeds fruit⁻¹ 55.30-600.00; seed weight fruit⁻¹ 0.94-9.12 g; 100 seed weight 0.66-2.95 g and seed texture was found medium soft in most of the genotypes. Similarly, the chemical quality attributes also varied significantly among all the different genotypes. Among the genotypes, the total soluble solids content varied between 10.80-16.33°B; acidity 0.28 to 0.70%; TSS:acidity ratio 20.37-39.82; pectin 0.88-1.42%; ascorbic acid 136.50-280.50 mg/100 g; reducing sugar 4.11-7.45%, non-reducing sugar 1.11-2.83% and total sugars 5.70-9.78% and high variability was also recorded in mineral contents of fruits which ranged from 11.48-17.48 P, 268.37-370.17 K, 16.31-23.18 Ca and 12.62-24.66 mg 100 g⁻¹ FW. Lycopene content in pink fleshed guava genotypes ranged from 0.67-2.43 mg 100 g⁻¹. Results of the study revealed that different genotypes exhibited wide range of diversity with respect to quality attributes under rain-fed semiarid conditions of Gujarat.

Key words: Psidium guajava, genotype, physico-chemical attributes.

INTRODUCTION

Guava is one the most delicious and popular fruits, widely grown in tropical and subtropical regions of India. It is rich source of vitamin C and minerals and is common raw material in fruit processing industry. Among most of the tropical and subtropical fruit trees guava plants exceed in adaptability, productivity, tolerance to adverse weather conditions and possibility of value addition makes quava an important fruit crop (Tiwari et al., 13). A large number of named cultivars are available in India, only a few like Allahabad Safeda and L-49, occupy the major area under its cultivation. Efforts have been made over past few decades to widen the genetic base through creating new variability or by utilizing natural variability for selection of elite variety (Tiwari et al., 13). In India, it is the fourth and fifth most important fruit crop by area and production, respectively. It occupies an area of 0.27 m ha with a total production of 3.67 mt. In Gujarat, it is mainly grown in Ahmadabad, Bhavnagar, Rajkot and Bharuch districts with a total production 140.80 thousand tonnes from an area of 10.80 thousand ha: the average productivity is 13.0 t ha⁻¹ (Anonymous, 1). Guava exhibits high levels of genetic diversity which is due to prevalence of seed propagation in these areas. Therefore, survey was undertaken to find out

MATERIALS AND METHODS

The diversity rich areas of semi-arid areas of Gujarat *viz.*, Panchmahals, Mahisagar, Vadodara Bharuch, and Bhavnagar were surveyed extensively

the diversity in fruit characteristics, and also to select elite seedlings from existing heterozygous seedling population having desirable horticultural traits. The physico-chemical attribute of the fruit are important as high TSS and titrable acidity in fruits along with red pulp are desirable for processing industry and low acidity and high TSS are desirable for fresh consumption (Corrêa et al., 4). For development of improved guava cultivars, a diverse gene pool is essential. Knowledge of the genetic diversity available and the origin of the cultivars would assist in the selection of parents for effective improvement programmes (Singh et al., 12; Hazarika et al., 6). In this regard, Yadav and Shankar (14) identified several elite seedling guava types based on bearing and fruit quality while surveying in Allahabad region. Development of nutrient rich cultivars has been a focus of fruit breeding studies (Corrêa et al., 4). Similar approach has been followed by Singh et al. (11) for identification of elite genotypes of wood apple seedling from Gujarat. Keeping above facts in view, an attempt was made to identify elite genotypes and their ex-situ establishment in field gene bank for further evaluation and crop improvement.

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to identify elite guava genotypes during the fruiting season of 2016 and 2017. The detailed about germplsm and their source of collection are given in Table 1. Matured fruits were collected from selected trees to study the physico-chemical characteristics. Ten fruits were randomly selected from all the directions for recording the data and brought to the laboratory of Central Horticultural Experiment Station, Vejalpur, Panchmahal (Godhra), Gujarat. For measuring physical parameters like fruit weight, seed weight and fruit size were recorded as per standard procedures with the help of an electronic balance and vernier caliper respectively. The fruit shape, fruit shape at stalk end and pulp colour were described with the help of standard descriptors for guava prescribed by Rodrguez et al. (9). Seed hardness was observed organoleptically. Juice was filtered through filter paper, thereafter, juice samples were subjected to determine the following parameters; titratable acidity (% of citric acid) using N/10 NaOH and phenolphthalein as indicator, total soluble solids (TSS°Brix) using hand refractometer and ascorbic acid (mg 100 g⁻¹ pulp) using a dye (2, 6-dichlorophenol indophenol), pectin, sugar contents and Lycopene content (mg100 g⁻¹) according to the standard method of AOAC (2). To determine the mineral contents on fresh weight (FW) basis in fruit pulp, 3g fruit pulp passed through nitricperchloric (9:4) digestion. Potassium, phosphorus, magnesium and calcium were determined by the method described by Bhargava and Raghupathi (3) and expressed as mg 100 g⁻¹ FW. The data were statistically analyzed as per method outlined by Gomez and Gomez (5).

RESULTS AND DISCUSSION

Results on the fruit morphological characters showed wide range of variation amongst different guava genotypes. In all studied genotypes, considerable variability was observed in fruit shape, seed content, peel and pulp colour (Table 1). Fruit morphological characters are complex trait, which depends upon genetic makeup and edapho-climatic conditions and their interaction. The rich diversity in these characters may be due to highly heterozygous and diverse genetic background of parents (Singh *et al.*, 10). Diverse fruit characters in wood apple were also observed by Singh *et al.* (11) while making surveys in Gujarat.

The analysis of variance of 25 guava genotypes studied in this investigation revealed significant differences in various physico-chemical characters of the fruits (Table 2 & 3). The fruit weight of different genotypes ranged between 53.50 and 318.50 g being maximum in GG-8 (318.50 g) followed by GG-7 (292.50 g), GG-19 (262.25 g) and GG-21 (253.66 g). However, the minimum fruit weight was observed in GG-12 (53.50 g) followed by GG-22 (88.20 g) and GG-9 (98.50 g). The diverse fruit weight in different guava genotypes has been reported by Yadav and Shankar (14) in guava, Hazarika et al. (6) in hatkora, Singh et al. (10) in aonla and Singh et al. (11) in wood apple genotypes. Among the different genotypes studied, the maximum fruit length was recorded in GG-21 (9.95 cm), followed by GG-8 (9.53 cm), while the minimum fruit length was recorded in GG-22 (4.09 cm) followed by GG-16 (5.20 cm). The fruit width varied from 4.30 cm in GG-12 followed by 4.50 cm in GG-22 to 8.25 cm in GG-8 followed by 7.90 cm in GG-7. However, highest fruit length: width ratio was recorded in GG-11 (1.45) followed by GG-21 (1.40), whereas it was lowest in GG-15 (0.86). The length of seed core varied from 2.5 to 5.45 cm. The maximum seed core length was recorded in GG-5 (5.45 cm) followed by GG-8 (5.25 cm), whereas, the minimum seed core length was measured in GG-12 (2.50 cm) followed by GG-22 (2.70 cm). The highest pulp thickness was observed in GG-8 (2.30 cm) and least in GG-12 (1.00 cm). The number of seed fruit⁻¹ varied between 55.30 to 609 and being the maximum in GG-25 (609.0) followed by GG-20 (600.0) and GG-20 (510.0) and the minimum number of seed fruit⁻¹ was recorded in GG-12 (55.30) followed by GG-9 (98) among the studied genotypes of guava. The maximum seed weight fruit⁻¹ was recorded in GG-20 (9.12 g) followed by GG-13 (8.16 g), while it was found minimum in GG-12 (0.94 g) followed by GG-19 (1.62 g) and GG-23 (1.85 g). Among the different genotypes, 100 seed weight was recorded highest in fruits of GG24 (2.95 g) and it was lowest in GG16 (0.66 g). It is established fact that the softness of seed influences the guality of fruit, which was found soft in GG-18, GG-19, GG-22 and GG-23; medium soft in GG-1, GG-3, GG-4, GG-5, GG-6, GG-8, GG-10, GG-11, GG-14, GG-15, GG-16, GG-17 and GG-21; medium hard in GG-2, GG-7, GG-9, GG-15 and GG-16 and hard in GG-12, GG-13, GG-20, GG-24 and GG-25. Similar variations in fruit size, seed core dia, pulp thickness and seed content among diverse guava genotypes were reported by Yadav and Shankar (14) and Singh et al. (12).

The data presented in Table 3 showed significant variations in chemical quality attributes and mineral contents of fruits. The TSS of fruit juice ranged from 10.80 to 16.33 °B, being the highest in GG-24 (16.33 °B) followed by GG-21 (16.0 °B), while the minimum TSS was recorded in GG-19 (10.80 °B). The titrable acidity in fruit juice ranged from 0.28 to 0.70%, being the highest in GG-24 (0.70%) followed by GG-21 (0.60%) and GG-20 (0.56%) and the minimum

Genotypes	Latitude	Longitude	Elevation	Place of	Shape	Peel colour	Pulp colour	Shape at	Calyx	Longitudinal	Fruit
				collection				stalk end	cavity	ridges	surface
GG-1	22049'46.01"	73047'41.74"	153.62 m	Bodidra	Subglobose	Yellowish green	White	Rounded	Small	Absent	Smooth
GG-2	22052'56.78"	73049'05.27"	142.04 m	Rampur	Round	Green	Red	Rounded	Medium	Present	Rough
GG-3	22034'45.49"	73036'51.12"	113.99 m	Paroli	Round	Yellow	White	Broadly rounded	Small	Absent	Smooth
GG-4	22034'44.80"	73036'51.29"	113.39 m	Paroli	Round	Orange grrenish	Creamish white	Broadly rounded	Small	Absent	Smooth
GG-5	22034'48.71"	73036'51.61"	113.53 m	Paroli	Round	Yellow	Creamish white	Rounded	Broad	Absent	Smooth
GG-6	22053'54.15"	73048'05.94"	140.21 m	Rampur	Pyriform	Yellowish green	White	Pointed	Small	Present	Rough
GG-7	22052'54.39"	73048'02.64"	136.55 m	Rampur	Subglobose	Dark yellow	White	Rounded	Small	Absent	Rough
GG-8	22045'09.73"	73038'23.27"	127.41 m	Godhra	Pyriform	Yellow green	White	Rounded	Small	Present	Rough
6-99	22045'08.03"	73038'27.68"	127.40 m	Godhra	Conical	Yellow green	Pink	Pointed	Small	Absent	Smooth
GG-10	22045'04.32"	73038'19.61"	129.84 m	Godhra	Round	Yellowish green	White	Rounded	Broad	Present	Rough
GG-11	22031'18.71"	73040'38.59"	155.14 m	Zingiri	Pyriform	Green	White	Necked	Small	Absent	Smooth
GG-12	22034'43.41"	73036'53.17"	112.47 m	Paroli	Obovate	Yellow	Light pink	Rounded	Small	Absent	Smooth
GG-13	22051'50.48"	73049'05.60"	150.88 m	Kasanpur	Conical	Greenish yellow	White	Pointed	Small	Absent	Smooth
GG-14	22018'46.67"	73025'02.13"	61.57 m	Waghodiya	Conical	Pale yellow	Light pink	Pointed	Small	Present	Smooth
GG-15	21043'20.43"	71056'46.15"	41.45 m	Sihor	Round	Grey yellow	Pale pink	Broadly	Small	Absent	Smooth
	"00 00,070 PC	"00 0012077	AF 44	, o H O		Vellen				A 6 0 0 4	4100000
GG-16	21043'08.68"	71057'02.33"	45.41 m	Sinor	Kound	Yellow	Any	Kounded	Broad	Absent	Smooth
GG-17	23019'03.03"	72044'44.62"	90.83 m	Chandrala	Oblong	Greenish yellow	Creamish white	Necked	Small	Absent	Smooth
GG-18	22054'45.46"	73052'25.84"	159.72 m	Kuazar	Round	Orange grrenish	Yellow white	Rounded	Broad	Present	Smooth
GG-19	23012'39.36"	73033'22.94"	109.73 m	Veerpura	Oblong	Yellow	Creamish white	Rounded	Small	Present	Rough
GG-20	23012'14.45"	73033'56.05"	107.59 m	Veerpura	Subglobose	Yellowish	Pale pink	Rounded	Small	Absent	Rough
56.21	21042'35 0E"	71064'44 40"	67 06 m	Moto curbbo	Conical	Vallow	Jain oleO	Dobariod	Cmall	Abcont	doilod
66-22		/3096/26.9/	182.27 m	Pipliya	Kound	Green	White	Broadly rounded	Small	Absent	Smooth
GG-23	21043'06.07"	71057'05.84"	47.24 m	Sihor	Conical	Whish yellow	Creamish	Pointed	Small	Absent	Smooth
GG-24	22051'21.03"	73045'39.73"	164.59 m	Rampur	Round	Greenish yellow	Pale pink	Truncate	Small	Present	Bumpy
GG-25	22050'49.55"	73045'02.19"	168.55 m	Rampur	Subglobose	Pale yellow	Creamish light pink	Rounded	Broad	Absent	Rough

Table 1. Topography of place of collection of guava genotypes and their fruit characteristics.

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Lycopene (mg 100 2.12 0.97 2.43 2.32 0.67 1.86 1.14 1.57 4. g_¹) 1.7 ł Seed texture Medium hard Medium hard Medium hard Medium hard Medium hard Medium soft Hard Hard Hard Soft Hard Hard Soft Soft soft 100 seed weight (g) 0.016 0.046 2.15 1.28 2.69 1.73 1.86 1.53 2.1 1.72 0.80 1.70 1.62 1.31 1.36 0.66 1.59 1.53 1.72 1.54 1.69 2.18 1.68 2.95 1.61 2.77 1.55 1.71 veight / fruit (g) Seed 20.85 8.16 5.35 2.35 4.25 2.76 3.98 2.10 2.80 0.94 2.95 4.73 3.23 9.12 2.70 4.12 1.85 2.60 7.79 0.46 2.93 2.90 2.89 3.67 1.62 4.51 5.32 1.31 No. seed/ 158.40 06.20 69.35 215.03 315.40 74.50 182.30 189.00 477.00 510.00 225.70 395.00 361.12 298.80 212.00 95.00 800.00 160.40 89.00 110.60 309.00 164.71 88.00 98.00 55.30 19.46 fruit 6.85 4.82 :hickness Pulp 17.29 1.45 1.25 I.40 I.32 1.80 2.25 2.30 1.20 1.75 1.30 1.00 1.40 1.30 1.70 1.50 1.83 1.65 2.10 1.70 1.75 1.20 1.65 1.20 0.15 0.44 (cm) 4 1.66 Seed core length (cm) 5.25 4.15 2.50 3.90 2.80 4.20 3.70 4.40 3.60 2.70 3.30 4.10 4.22 4.13 4.00 4.40 5.45 3.87 4.60 3.50 5.20 4.30 5.20 4.20 4.30 0.58 SS ı Length: width 1.15 1.13 1.06 1.45 1.30 0.95 1.38 1.40 1.19 0.99 0.93 0.99 1.02 0.86 0.87 0.06 0.19 9.82 0.94 0.88 .28 1.17 1.37 0.91 .03 1.01 1.21 1.21 Width 6.40 7.12 7.90 8.25 5.30 4.30 7.05 5.50 6.04 7.43 7.67 6.10 7.13 4.50 6.05 6.20 6.26 0.56 14.94 (cm) 6.90 7.41 5.91 6.07 7.20 7.32 1.61 6.93 7.01 Length 14.05 8.39 9.16 9.30 7.86 5.60 7.20 6.20 5.20 6.47 9.33 8.40 9.95 4.09 7.18 6.17 0.58 (cm) 6.16 6.82 6.33 7.10 9.53 7.32 6.37 6.50 6.0 8.61 1.67 Weight (g) 292.50 204.73 198.35 187.75 108.50 155.16 202.20 262.25 187.45 169.45 58.60 187.30 155.34 173.20 234.75 318.50 163.07 186.50 184.20 253.66 137.50 123.11 98.50 53.50 88.20 1.63 4.63 1.60 Genotypes CD (0.05) GG-16 GG-18 GG-19 GG-10 GG-15 GG-20 GG-25 GG-12 GG-13 GG-14 GG-17 GG-22 GG-23 GG-24 SEm± GG-21 GG-11 GG-8 6-99 GG-5 9-99 GG-7 GG-1 GG-2 66-3 GG-4 CV%

Table 2. Physical quality attributes and lycopene content of guava genotypes.

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Genotypes	(aº) SST	Acidity	TSS:	Ascorbic acid	Reducing	Non-reducing	Total	Pectin	Mineral	I content (mg	mg 100 g ⁻¹	FW)
		(%)	acidity	(mg/100g)	sugar (%)	sugar (%)	sugar (%)	(%)	٩.	×	Ca	Mg
GG-1	12.24	0.35	34.97	222.20	6.25	2.35	8.72	1.27	15.41	362.97	23.19	24.66
GG-2	11.30	0.42	26.90	163.50	5.80	2.47	8.40	1.33	13.43	329.15	19.33	15.29
GG-3	15.20	0.40	38.00	230.50	7.45	2.07	9.63	0.92	14.18	322.88	19.02	21.93
GG-4	13.20	0.42	31.40	275.72	4.21	1.95	6.27	1.11	16.27	295.93	17.67	14.29
GG-5	11.15	0.28	39.82	243.10	4.29	1.98	6.38	0.97	14.47	339.61	20.84	15.01
GG-6	14.20	0.54	26.29	192.20	7.35	2.15	9.61	1.04	14.84	283.57	17.06	18.93
GG-7	15.50	0.45	34.44	280.50	7.61	2.06	9.78	1.07	13.96	315.47	19.64	13.57
GG-8	12.20	0.31	39.35	235.73	7.05	1.11	8.22	0.95	14.77	339.26	19.83	12.69
GG-9	15.30	0.49	31.22	167.80	5.70	1.63	7.42	1.24	17.48	308.18	18.28	21.57
GG-10	12.16	0.33	36.84	227.50	4.74	2.73	7.62	1.15	12.71	279.79	16.87	20.38
GG-11	13.50	0.43	31.39	189.56	5.97	2.27	8.36	1.02	13.18	273.11	17.55	20.53
GG-12	12.45	0.41	30.36	167.52	4.35	1.28	5.70	1.08	13.11	299.55	18.86	22.71
GG-13	15.25	0.52	29.32	274.77	5.77	2.53	8.44	1.13	15.32	268.37	16.31	12.62
GG-14	11.50	0.41	28.04	195.63	5.54	1.27	6.88	1.29	14.53	312.42	19.49	22.99
GG-15	13.10	0.38	34.47	239.30	4.82	2.75	7.72	1.42	13.25	319.17	20.53	23.09
GG-16	13.00	0.45	28.88	189.60	5.32	1.32	6.71	1.31	14.43	335.63	20.64	23.14
GG-17	14.00	0.37	37.83	240.50	7.15	1.09	8.20	1.22	15.35	288.71	17.32	20.18
GG-18	14.30	0.50	28.60	203.75	6.30	2.37	8.80	0.98	11.48	272.95	16.54	17.34
GG-19	10.80	0.53	20.37	200.50	4.11	2.18	6.41	1.09	13.66	345.19	21.12	23.66
GG-20	13.50	0.56	24.10	177.32	5.18	1.20	6.45	1.17	12.58	303.86	18.07	21.17
GG-21	16.00	09.0	26.66	220.75	6.57	1.60	8.26	1.39	11.59	366.15	22.16	15.72
GG-22	13.00	0.35	34.28	136.50	6.01	2.83	8.99	0.95	15.81	291.46	18.46	21.54
GG-23	12.40	0.38	32.63	215.45	6.82	1.26	8.15	1.20	12.83	370.17	21.36	16.84
GG-24	16.33	0.70	23.32	249.00	7.33	1.99	9.43	0.96	13.58	333.28	20.53	20.86
GG-25	13.50	0.46	29.34	176.02	5.17	1.22	6.46	0.88	13.38	289.39	17.36	20.32
SEm±	0.54	0.026	2.23	1.63	0.50	0.06	0.50	0.02	0.59	1.46	0.73	0.77
CD (0.05)	1.55	0.074	6.40	4.64	1.51	0.19	1.50	0.06	1.68	4.16	2.07	2.19
CV%	7.06	10.26	12.48	15.28	15.60	5.93	11.63	3.26	7.27	0.80	6.58	6.93

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acidity was noted in GG-5 (0.28%). The TSS:acid ratio among different accessions varied from 39.82 in GG-5 followed by 39.35 in GG-8 to 20.37 in GG-19 followed by 23.32 in GG-24. The maximum ascorbic acid content was recorded in GG-7 (280.50 mg 100 g^{-1}), followed by GG-4 (275.72 mg 100 g^{-1}), while lowest ascorbic acid was recorded in GG-22 (136.50 mg 100 g⁻¹). GG-7 had the maximum reducing (7.61%) and total sugar (9.78%) followed by GG-6 (7.35 & 9.61%), while the minimum reducing and total sugar was recorded in GG-19 (4.11%) and GG-12 (5.70%) respectively. The highest non-reducing content was obtained in GG-22 followed by GG-10, and it was the minimum in GG-17 (1.09%). Similar kind of variation is recorded by Yadav and Shankar (14) and Corrêa et al. (4) in guava. The pectin content differed significantly among the different genotypes and ranged from 0.88 to1.39 %. The highest pectin content was recorded in GG-15 (1.42%) followed by GG-21 (1.39%) and the lowest was observed in GG-25 (0.88%). Guava fruit is one of the best sources of food grade pectin, which is used for making good quality jelly (Patel et al., 8). This finding is in agreement with the observations of Singh et al. (12) and Yadav and Shankar (14) in guava. Similarly, sugar contents also varied significantly among the genotypes. The highest value for reducing sugar (7.61%) and total sugars (9.78%) were recorded in GG-7, while, the lowest reducing sugar (4.11%) and total sugars were recorded in GG-19 and GG-12 respectively. The maximum non-reducing content was obtained in GG-22(2.83%) followed by GG-13 (2.53%) and it was the minimum in GG-17 (1.09%). The highest lycopene content was also observed in pink pulped guava genotypes which ranged from 0.67 mg 100 g⁻¹ in GG-25 to 2.43 mg 100 g⁻¹ in GG-21 (Pandey et al., 7). Various workers have recorded similar kind of variation in guava genotypes, which may be due to different genetical constitution of the individual genotypes (Yadav and Shankar, 14; Patel et al., 8).

There was a significant difference among the genotypes with respect to mineral contents of guava fruits on fresh weight (FW) basis (Table 3). Among the 25 guava genotypes, the highest level of phosphorus was found in GG-9 (17.48 mg 100 g⁻¹) followed by GG-4 (16.27 mg 100 g⁻¹), and the lowest was recorded in GG-18 (11.48 mg 100 g⁻¹). The potassium content ranged from 268.37-370.17mg 100 g⁻¹ being the maximum in GG-23 (370.17 mg 100 g⁻¹) followed by GG-1 (362.97 mg 100 g⁻¹) and it was minimum in GG-13 (268.37 mg 100 g⁻¹). The highest calcium content was recorded in GG-1 (23.19 mg 100 g⁻¹) followed by GG-21(22.16 mg 100 g⁻¹), while the lowest was recorded in GG-13 (16.31 mg 100 g⁻¹).

The magnesium content ranged between 12.62-24.66 mg 100 g⁻¹ being the maximum in GG-1 (24.66 mg 100 g⁻¹) followed by GG-19 (23.66 mg 100 g⁻¹) and minimum in GG-13 (12.62 mg 100 g⁻¹). Corrêa *et al.* (4) found wider variation for fruit mineral contents in various guava genotypes of seedling origin. This finding is also substantiated by the results reported by Singh *et al.* (11) in wood apple.

In general, the result of the study showed wider diversity for various physico-chemical characters especially for high TSS:acidity ratio and mineral contents. Based on the results, it has been observed that, among all the genotypes of guava collected from different locations of Gujarat, GG-1, GG-4, GG-15, GG-21, GG-23 and GG-24 having the desirable physico-chemical characters for consumers and breeders.

REFERENCES

- 1. Anonymous. 2014. Indian Horticulture Data Base. 2014. NHB, Ministry of Agriculture, Govt. of India, Gurugram. pp. 42-48.
- AOAC. 1980. Official Methods of Analysis of Association of Official Agricultural Chemists, Association of Official Agricultural Chemists, Benjamin Franklin Station, Washington, DC, USA.
- Bhargava, B.S. and Raghupathi, H.B. 1993. Analysis of plant materials for macro and micronutrients. In: *Methods of Analysis of Soils, Plants, Water and Fertilizers*. Tandon, H.L.S. (ed.), Fertilizer Development and Consultation Organization, New Delhi, pp. 49-82.
- Corrêa, L.C., Santos, C.A.F. and Lima, G.P.P. 2012. Chemical and biochemical characterization of guava and araçá fruits from different regions of Brazil. *Acta Hort.* 959: 103-9.
- Gomez, K.A. and Gomez, A.A. 1984. Statistical Procedure for Agricultural Research, 2nd edn., John Wiley and Sons Inc., New York.
- Hazarika, T.K., Lalawmpuii, B. and Nautiyal, B.P. 2013. Studies on variability in physicochemical characters of hatkora (*Citrus* macroptera Mont.) collections of Mizoram. *Indian J. Hort.* **70**: 480-84.
- 7. Pandey, D., Pandey, A.K. and Yadav, S.K. 2016. Evaluation of newly developed guava cultivars & selections under Lucknow conditions. *Indian J. Hort.* **73**: 334-38.

- Patel, R.K., Maiti, C.S., Deka, B.C., Deshmukh, N.A. and Nath, A. 2013. Changes in sugars, pectin and antioxidants of guava (*Psidium guajava*) fruits during fruit growth and maturity. *Indian J. Agric. Sci.* 83: 1017-21.
- Rodríguez-Medina, N.N., Fermin, G.A., Valdés-Infante, J., Velásquez, B., Rivero, D., Martínez, F., Rodríguez, J. and Rohde, W. 2010. Illustrated descriptors for guava (*Psidium guajava*). Acta Hort. 849: 103-10.
- Singh, A.K., Singh, P.P., Singh, S., Bhargava. R. and Makwana, P. 2016. Variability in morphological and physico-chemical traits of aonla (*Emblica officinalis*) genotypes collected from north-eastern region of India. *Indian J. Agric. Sci.* 86: 992-97.
- 11. Singh, A.K., Singh, S., Yadav, V. and Sharma, B.D. 2016. Genetic variability in wood apple

(*Feronia limonia*) from Gujarat. *Indian J. Agric. Sci.* **86**: 1504-08.

- Singh, D., Gill, M.I.S., Boora, R.S. and Arora, N.K. 2015. Genetic diversity analysis in guava (*Psidium guajava*) on the basis of morphological and physico-chemical traits. *Indian J. Agric. Sci.* 85: 678-83.
- Tiwari, J.P., Bisen, B. and Mishra, D.S. 2003. Improvement of guava using indigenous strains. In: *Role of Indigenous Germplasm In Improvement of Horticultural Crop*. Singh, R., Ram, H.H. and Tiwari, J.P. (eds.), Bishen SMPS Publ & Dis of Scientific Books, Dehra Dun, pp. 227-34.
- Yadav, L.P. and Shankar, G. 2007. Exploiting genetic diversity in guava (*Psidium guajava* L.) from Allahabad and surrounding areas. *Pt. Gen. Reso. News Lett.* **149**: 14-16.

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